Adrenoceptors, Uncoupling Proteins, and Energy Expenditure

SHEILA COLLINS,†* WENHONG CAO,* KIEFER W. DANIEL,* TONYA M. DIXON,†
ALEXANDER V. MEDVEDEV,* HIROKI ONUMA,* AND RICHARD SURWIT*

*Departments of Psychiatry and Behavioral Sciences, and †Pharmacology, Duke University Medical Center, Durham, North Carolina 27710

Interest in the biology of adipose tissue has undergone a revival in recent years with the discovery of a host of genes that contribute to the regulation of satiety and metabolic rate. The catecholamines have long been known to be key modulators of adipose tissue lipolysis and the hydrolysis of triglyceride energy stores. However, more recent efforts to understand the role of Individual adrenergic receptor subtypes expressed in adipocytes and their signal transduction pathways have revealed a complexity not previously appreciated. Combined with this interest in the modulation of adipocyte metabolism is a renewed focus upon brown adipose tissue and the mechanisms of whole body thermogenesis in general. The discovery of novel homologs of the brown fat uncoupling protein (UCP) such as UCP2 and UCP3 has provoked intensive study of these mitochondrial proteins and the role that they play in fuel metabolism. The story of the novel UCPs has proven to be intriguing and still incompletely understood. Here, we review the status of adipose tissue from inert storage depot to endocrine organ, interesting signal transduction pathways triggered by β-adrenergic receptors in adipocytes, the potential of these receptors for discriminating and coordinated metabolic regulation, and current views on the role of UCP2 and UCP3 based on physiological studies and gene knockout models.

[Exp Biol Med Vol. 226(11):982-990, 2001]

Key words: β-receptors; adipose tissue; brown fat; UCP; obesity

Introduction: Adipose Tissue As a Storage Depot and Thermogenic Organ

Old and New Ideas about the Role of Body Fat. In days of yore, to be well-endowed with a layer of fat was to be wealthy, respected, attractive, even godlike (Fig. 1). In more recent times, it has been more popular to strive for the least amount of body fat, a concept glamorized by fashion, art, and even medicine, given that overweight or

1535-3702/01/22611-0982\$15.00
Copyright © 2001 by the Society for Experimental Biology and Medicine

obesity is linked to increased risk for a host of serious medical complications such as coronary artery disease, diabetes, and hypertension (1). However, when taken to the extreme, as in lipodystrophy, it is clear that the absence of adipose tissue is as detrimental as having too much fat (2). What then is the role of adipose tissue and how is its metabolism regulated?

Until this past decade of the 1990s, adipose tissue has been largely considered to be an inert storage depot, with access to stored triglycerides being gated by the adrenergic receptors and their ability to stimulate lipolysis. The discovery of leptin as an adipose-derived hormone that "reports" on the status of energy reserves to other organs of the body, including the central nervous system, gave us new appreciation for adipose tissue biology. It is now also known that several cytokines and growth factors are secreted from adipose tissue and may play significant roles in insulin resistance, and cell differentiation and growth. In this sense, adipose tissue has now achieved status as an endocrine organ. Moreover, although we most commonly think of adipose tissue as something to shed by dieting and exercising, a form of adipose tissue known as brown fat could be considered to be, as once referred to in a New York Times Science Times piece, the "Type of Body Fat That Fights Obesity" (3).

The rich and varied history of brown adipose tissue as an anatomically discreet tissue includes early speculations in the 17th century that it was part of the thymus, and a century later that it was an endocrine organ involved in blood formation or a form of fat acting as a reservoir for certain nutrients (4). It was only in 1961 that brown adipose tissue was proposed to be thermogenic (5, 6). Since then, an immense body of work has shown that this tissue is uniquely capable of responding to various environmental stimuli to generate heat from stored metabolic energy. In response to sympathetic nervous system activation, brown adipose tissue undergoes an orchestrated hyperplastic and hypertropic expansion, increased blood flow, and recruitment of lipid and carbohydrate fuels for oxidative metabo-

¹ To whom requests for reprints should be addressed at Duke University Medical Center, Box 3557, Durham, NC 27710. E-mail: colli008@mc.duke.edu

lism (7, 8). A critical element in brown fat thermogenesis is its mechanism for dissipation of the mitochondrial proton gradient, a mechanism that involves the brown fat-specific mitochondrial uncoupling protein (UCP) (9), also called "thermogenin" (10). This mitochondrial protein, now termed UCP1, allows controlled proton leakage for the purpose of heat generation at the expense of coupled ATP production (Fig. 2). UCP1 is regulated at both the transcriptional level and at the level of the mitochondrion. As illustrated in Figure 2 and discussed in greater detail by Ricquier and Bouillaud (11), uncoupling in brown fat mitochondria is activated by free fatty acids that are released as a result of hormone-stimulated lipolysis. The cloning of brown fat UCP from rodents provided the opportunity to investigate the molecular mechanism of this mitochondrial uncoupling and the regulation of UCP gene expression by hormonal stimulation (12-14). When transgenic mice expressing UCP1 under the control of the adipose-specific aP2 promoter were generated such that UCP1 was expressed in both white and brown adipose tissue, these animals were remarkably resistant to obesity (15, 16). However, complicating the interpretation of these results was the fact that mice with a targeted disruption of the UCP1 gene were distinctly coldsensitive, but not obese (17).

Thermogenic Defects in Rodent Models of Obesity. From the earliest studies of the *ob/ob* (obese) mouse (now called leptin-deficient C57BL/6J Lep^{ob}), there was evidence that these mice were not only obese, hyperglycemic, and hyperinsulinemic, but were also extremely sensitive to cold (18). The blunted capacity for adrenergic stimulation of lipolysis in their adipose tissue (see below) probably also hindered activation of UCP1 function by free

fatty acids (Fig. 2). Studies of other monogenic obesity models and hypothalamic lesioned rodents have all indicated a complex set of neural and endocrine abnormalities, culminating in the loss of homeostatic mechanisms controlling both food intake and metabolic efficiency (19).

Adrenergic Receptors and Adipose Tissue Metabolism

Three β-Adrenergic Receptor Subtypes in Adipocytes. The ability of norepinephrine to stimulate lipolysis and thermogenesis in adipocytes is controlled largely by the β -adrenergic receptors (β ARs). They are members of the large family of G protein-coupled receptors that regulate an assortment of intracellular second messenger systems, including cAMP, phospholipid hydrolysis, ion fluxes, and mitogen-activated protein (MAP) kinase cascades. Early studies to pharmacologically characterize the adrenergic receptors in adipose tissue concluded that there was a single βAR subtype present (20–22), but further pharmacological studies could not clearly define which subtype(s) it was. Molecular cloning eventually led to identification of the β₃AR, and it was confirmed that this new receptor was the target of novel lipolytic and thermogenic β-agonists developed by Arch, Cawthorne, and colleagues (23). We now know that all three βAR subtypes: $\beta_1 AR$, β_2 AR, and β_3 AR, are expressed in white and brown adipocytes (24-27). Unlike β₁AR and β₂AR, which are broadly expressed, expression of the β₃AR gene in adipocytes is a function of differentiation (28, 29). Similar to certain other adipocyte-specific genes (30, 31), the differentiationdependent transcription of the \(\beta_3 AR \) gene requires expression of C/EBP α for both its induction and maintenance (29).

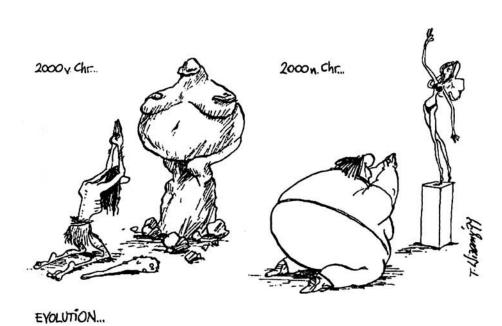


Figure 1. "Evolution" of our concepts about the value of body fat.

Mechanism of Uncoupling Protein Action in Mitochondria

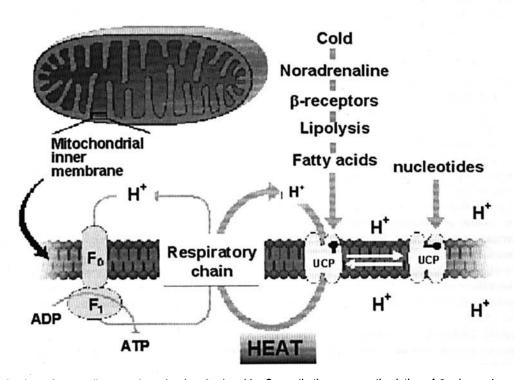


Figure 2. Mechanism of uncoupling protein action in mitochondria. Sympathetic nervous stimulation of β-adrenergic receptors on brown adipocytes occurs in response to cold temperature or overfeeding (89), leading to lipolysis and activation of the thermogenic mediator, UCP1. Binding of free-fatty acids liberated from lipolysis stimulates the action of UCP1, releasing the tonic inhibition by guanine nucleotides. This "uncoupling" of mitochondrial respiration from ATP production results in an increased net expenditure of caloric energy per ATP produced, with heat generated as the by-product.

Impaired Adipose Tissue Adrenergic Signaling in Obesity. For many years it was known that obese C57BL/6J Lepob and C57KsJ LepRdb mice exhibited a marked inability to effectively mobilize triglycerides from white adipose tissue (32-35). These animals were also unable to recruit brown adipose tissue for thermogenesis in response to cold temperature-induced adrenergic stimulation (35, 36), indicating that adrenergic mechanisms regulating metabolism in both white and brown fat were affected in obesity. Although defects in sympathetic outflow have been shown to be associated with obesity (37-39), other experiments clearly indicated that there was impaired B-adrenergic receptor function at the level of the adipocyte itself, independent of the availability of catecholamines (34). Investigators tried to determine the nature of the molecular defect in adipocytes from obese animals. The components of the adrenergic signal transduction pathway (at least those that were known at the time and that did not include β_3AR) were examined, and despite a severe blunting of the β-adrenergic response, adenylyl cyclase itself and other downstream effectors of the lipolytic process did not differ between lean and obese animals (34, 40, 41). Collectively, these results led to the conclusion that the signal transduction mechanism of the BAR(s) must be defective. However, because the classical βAR radioligands such as cyanopindolol only bind the $\beta_3 AR$ very weakly, this receptor went undetected, and estimates of $\beta_1 ARs$ and $\beta_2 ARs$ were also distorted

In the first study (25) comparing expression and activity of all three βAR subtypes in the genetically obese C57BL/6J Lep^{ob}, we found a marked decrease in expression of the β_3AR , as well as that of the β_1AR . Combined with a series of pharmacologic analyses, it was found that these reductions in BAR expression corresponded functionally to the impaired stimulation of cAMP production, and they appeared to be largely responsible for the defects in catecholamine-stimulated lipolysis observed in the C57BL/6J Lep^{ob} mouse. Similar findings of depressed β₃AR mRNA levels in the Zucker fatty (fa/fa) rat were reported by Muzzin et al. (42), but their relationship to changes in the function of the receptor was not examined in that study. In an extension of these studies, we found significant deficits in expression and function of adipocyte BARs in essentially every model of obesity that we have examined, including obesity induced by high-fat feeding in non-mutant mice (43).

Selective β_3AR Agonists as Potential Thermogenic and Antiobesity Agents. Beginning with the first reports by Arch and colleagues (23) that atypical β -ad-

renergic ligands had thermogenic and weight-reducing properties in C57BL/6J Lepob mice, there has been great interest in trying to understand their biochemical and physiological effects. In most species studied, including some nonhuman primates, β₃AR-agonist treatment is associated with increased oxygen consumption, decreased white adipose tissue mass, and increased density of brown adipocytes expressing UCP1 within typical white adipose depots (44-48). From our studies in various inbred strains of mice, the relative success of β₃-agonists as an anti-obesity therapy appears to parallel the extent of this expansion of brown adipocytes (46). Others reported similar effects of cold exposure as well as of acute β₃-agonist stimulation in a series of recombinant inbred strains (49). Importantly, we have also observed that the beneficial effects of β₃AR agonists to decrease adipose tissue mass and improve glycemic control in mouse models of obesity and diabetes can persist, even after many weeks of chronic treatment (46). This apparent lack of desensitization is rather unusual, particularly since tachyphylaxis is a hallmark of most receptor systems. Perhaps β₃AR activation and stimulation of down-stream effectors can continue because the β_3AR is neither a target for phosphorylation (50) nor does it bind β -arrestin (51), an accessory protein involved in G protein-coupled receptor desensitization (52).

Many studies have now documented the potent antiobesity and anti-hyperglycemic properties of selective β_3AR agonists in a variety of animal models (23, 45, 46, 48, 53). These observations have fueled intense investigation of these drugs as potential obesity and/or diabetes therapies for humans. We have recently discussed some of the issues surrounding the abundance and activity of β_3AR in humans elsewhere (54) and so we will not address this issue here.

Novel Signaling Properties of the β₃AR. Over 25 years ago, Rodbell and colleagues (55, 56) made the observation that there was an unusual, biphasic stimulation of cAMP production in adipocytes in response to the BAR agonist isoproterenol, the inhibitory phase of which could be relieved by pretreatment of adipocytes with pertussis toxin (57). With the cloning of the β_3AR gene and the development of highly selective β₃AR agonists (23, 58), it was postulated that this novel adipocyte-specific BAR may be responsible for the biphasic adenylyl cyclase response in adipocytes (59). In fact, we previously noted that despite the relatively high level of expression of the β₃AR in adipocytes, the efficiency of its coupling to stimulation of adenylyl cyclase was low (25). We recently reported that the β₃AR is simultaneously coupled to both Gs and Gi, with the consequent activation of the protein kinase A (PKA) and MAP kinase (ERK1/2) pathways, respectively (60). More recent work in our laboratory now shows that novel sequence elements within the \(\beta_3AR\) itself are responsible for the direct recruitment to the receptor of SH3 domaincontaining signaling molecules such as c-Src, and that this interaction is required to trigger the ERK cascade (51) (Fig. 3). These results highlight two major points. First, in terms of mechanisms of G protein-coupled receptor signaling, the recognition that such receptors can directly bind and activate members of the so-called growth factor signaling cascade defines a new paradigm in signal transduction crosstalk. Second, these interactions at the β_3AR suggest that this member of the βAR family could utilize this unique ability to immediately transmit signals to two different signaling cascades to more tightly regulate certain transcriptional and metabolic responses (what we like to refer to as the "safe deposit box" mechanism whereby specialized responses can only be elicited when both "keys" are turned simultaneously).

From a more physiologic perspective, we now wish to understand if the combined activation of the PKA and MAP kinase pathways could underlie the β_3AR agonist-dependent appearance of thermogenically active brown adipocytes in white fat depots, as well as the potential for integrated regulation of certain metabolic actions of β -agonists such as activation of hormone-sensitive lipase and the general program of lipogenesis versus lipolysis, as suggested in Figure 4. Further, more recent data indicate an interesting role for p38 MAP kinases in the β -agonist and cAMP-dependent regulation of UCP1 expression in brown adipocytes (61).

Ucp2 And Ucp3: Links to Resting Metabolic Rate, Fuel Metabolism, or Signal Transduction?

Evolution of Ideas about the UCP Homologs. The discovery of UCP2 and UCP3, both of which exhibit significant sequence and domain structural homology to UCP1 and the capacity to uncouple mitochondrial respiration in model systems (62–66) immediately led to the notion that they might be the explanation for the relative inefficiency of oxidative respiration seen in most cell types and thus, influence metabolic rate and fuel utilization. The structural homology between these UCPs, their close chromosomal locations to one another, and basic features about their regulation and expression in various rodent models and human populations have been recently reviewed (67–69).

The UCP2 and UCP3 genes reside in a chromosomal location on distal mouse chromosome 7 that is coincident with a quantitative trait locus (QTL) linkage to hyperinsulinemia and high plasma leptin levels (62). This close linkage relationship suggests that either or both of these UCPs could be related to this QTL. (Alternatively, the QTL could be linked to an unidentified gene in the region.) The finding that expression of UCP2 was specifically elevated in white adipose tissue in murine strains that are relatively resistant to the development of diet-induced obesity and diabetes, but not those which are obesity prone (70), plus the absence of such differences in UCP3 in any tissue, led us to reason that if UCP2 were an uncoupler of oxidative respiration, its activity would be consistent with increased energy expenditure and resistance to obesity. However, soon thereafter, several groups reported that fasting and/or starvation led to substantial increases in expression of both UCP2 and UCP3

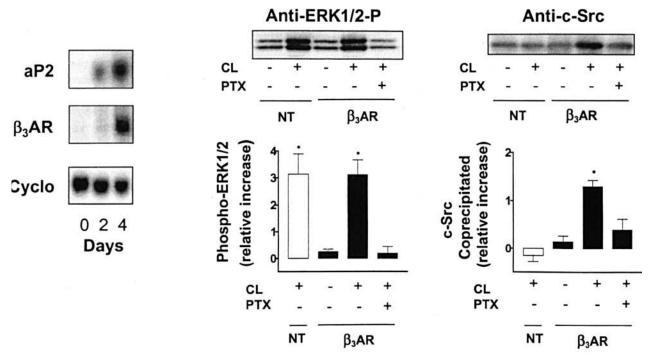


Figure 3. kb_3AR recruits c-Src in a Gi protein- and agonist-dependent manner in differentiated C3H10T1/2 adipocytes. C3H10T1/2 cells were transfected with hemaglutinin (HA)-tagged β_3AR and were differentiated as described (90). Immunoprecipitation assays (IP) were performed with anti-HA antibody. (A) Expression of aP2 and β_3AR mRNA as a function of differentiation on the indicated days. Levels of aP2 are maximal by Day 4. Cyclophilin RNA (cyclo) is a control for the Northern blot (45, 91). (B) The level of phosphorylated ERK1/2 (ERK1/2-P) in cell lysates and quantification of three independent experiments (mean \pm SD). (C) The level of c-Src coprecipitated with β_3AR and quantification of three independent experiments (mean \pm SD). Reprinted from Ref. 50.

in skeletal muscle and adipose tissue (71, 72). Dulloo and colleagues (73-75) showed that blockade of the fastinginduced rise in free fatty acids completely prevented the increase in UCP2 and UCP3 mRNA. Together, these findings suggested that these UCPs were participating in the metabolic adaptations required during the fasted state, adaptations that require a switch from predominantly glucose to predominantly fatty acids as a fuel source. The molecular mechanisms regulating expression of UCP2 and UCP3 genes are currently under investigation, but it has been postulated that this expression is increased by fatty acids. In support of this idea are reports that ligands for peroxisome proliferating-activated receptors (PPARs) γ and α increased UCP2 expression in adipocyte cell cultures (76–78). More recently, we have found that the PPARs regulate transcription of the UCP2 gene by an indirect mechanism involving the recruitment of other transcription factors (79).

Insights from UCP Transgenic and "Knockout" Models. Faced with studying proteins whose function was uncertain, several investigators approached this dilemma by generating mice with either targeted disruptions or tissue-specific overexpression of UCP genes. The first reports came from UCP3 knock-out mice (80, 81). These mice were neither obese nor diabetic, although there was some evidence for increased mitochondrial production of reactive oxygen species (ROS). In contrast, mice with robust over-

expression of UCP3 in skeletal muscle were hyperphagic, had decreased adipose tissue mass, and had increased glucose disposal (82). However, a recent report from Martin Brand's laboratory (83) indicates that UCP2 is not an uncoupler when expressed at physiologic levels and suggests that overexpression models may not be representative of physiological conditions. Clearly, these UCPs are proteins whose function we are still trying to determine.

A connection between mitochondrial uncoupling and ROS production has been proposed (84), and evidence linking this production and UCP2 expression in hepatocytes has been reported (85). In the hopes of understanding the physiologic role of the UCP2 gene, we, together with Ricquier and colleagues (86), recently generated mice with a targeted disruption of UCP2. These animals were not obese, did not gain weight differentially when challenged with a high-fat diet, and were not sensitive to cold exposure. However, they were completely resistant to certain infectious agents, and we found that UCP2 modulated production of cytokines and ROS in their macrophages (86). This result was not completely surprising given the relative abundance of transcripts for UCP2 in tissues such as lung, spleen, and intestine (62, 87). Although under certain circumstances hepatocytes appear to be able to express UCP2 (85, 88), expression of UCP2 in liver is largely confined to resident Kupffer cells (phagocytes that serve to clear antigen entering the body

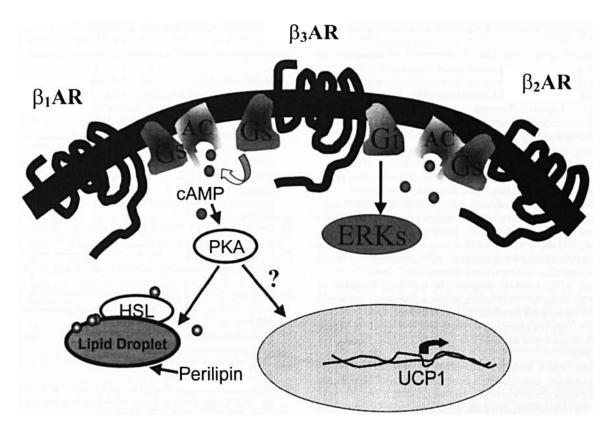


Figure 4. gkb-Adrenergic receptor subtypes in adipocytes and their signal transduction cascades. Two of the three receptor subtypes (β1 and β2) exert their effects via Gs while the β3 receptor interacts with both Gs- and Gi-linked signal transduction pathways.

from the gut), not to hepatocytes (89). This relationship between cytokines, ROS, and UCPs is intriguing, but considerably more work is required to understand this connection. We need to explore in greater detail the role of UCP2 in mechanisms of ROS sensing and signal transduction, to determine if changes in reducing equivalents from mitochondrial oxidation result from the lack of these UCPs, and to elucidate how the UCPs might contribute to modulated production of ROS. Finally, our *in vivo* data suggest the possibility that pharmacologic modulation of UCP2 might play a role in septic and parasitic challenges, immunosuppression, atherosclerosis, or apoptosis.

Summary

Studies of body weight regulation and the role of the sympathetic nervous system in adipose tissue metabolism continue to uncover new mechanisms that enhance our understanding of the intricate balance of fuel storage and utilization. Adrenergic stimulation of lipolysis and thermogenesis are impaired in most models of obesity, and the molecular bases of these alterations still need to be elucidated. Despite the extensive knowledge accumulated from pioneering work on brown fat thermogenesis, the discovery of homologs (UCP2 and UCP3) of the brown fat UCP have raised new questions concerning the possibility that UCP-mediated thermogenic mechanisms may exist in cells other than brown adipocytes. Although we do not yet understand the biochemical role(s) of UCP2 and UCP3, hints that they

participate in the generation of ROS and the modulation of ATP production continue to accumulate. There is still much work to be done, interesting hypotheses to explore, and undoubtedly surprises in store for us along the way as we continue to probe the molecular basis of fuel utilization, energy balance, and the control of body weight.

- Mokdad AH, Serdula MK, Dietz WH, Bowman BA, Marks JS, Koplan JP. The continuing epidemic of obesity in the United States. J Am Med Assoc 284:1650-1651, 2000.
- Reitman ML, Arioglu E, Gavrilova O, Taylor SI. Lipoatrophy revisited. Trends Endocrinol Metab 11:410–416, 2000.
- Browne MW. Type of Body Fat That Fights Obesity. The New York Times 28 Dec: Section C, page 5, Column 1, 1993)
- Lindberg O. Brown Adipose Tissue. New York: American Elsevier Publishing, 1970.
- Smith RE. Thermogenic activity of the hibernating gland in the coldacclimated rat. Physiologist 4:113, 1961.
- Ball E, Jungas R. On the action of hormones which accelerate the rate of oxygen consumption and fatty acid release in rat adipose tissue in vitro. Proc Natl Acad Sci U S A 47:932-941, 1961.
- Bukowiecki L, Collet A, Follea N, Guay G, Jahjah L. Brown adipose tissue hyperplasia: a fundamental mechanism of adaptation to a cold and hyperphagia. Am J Physiol 242:E353-E359, 1982.
- Géloën A, Collet AJ, Bukowiecki LJ. Role of sympathetic innervation in brown adipocyte proliferation. Am J Physiol 263:R1176-R1181, 1902
- Girardier L. The regulation of the biological furnace of warm-blooded animals. Experientia 33:1121-1131, 1977.
- Cannon B, Hedin A, Nedergaard J. Exclusive occurrence of thermogenin antigen in brown adipose tissue. FEBS Lett 150:129-132, 1982.
- 11. Ricquier D, Bouillaud F. Mitochondrial uncoupling proteins: from

- mitochondria to the regulation of energy balance. J Physiol 529:3-10, 2000.
- Bouillaud F, Ricquier D, Mory G, Thibault J. Increased level of mRNA for the uncoupling protein in brown adipose tissue of rats during thermogenesis induced by cold exposure or norepinephrine infusion. J Biol Chem 259:11583-11586, 1984.
- Kozak UC, Kopecky J, Teisinger J, Enerback S, Boyer B, Kozak LP. An upstream enhancer regulating brown fat-specific expression of the mitochondrial uncoupling protein gene. Mol Cell Biol 14:59-67, 1994.
- 14. Larose M, Cassard-Doulcier A-M, Fleury C, Serra F, Champigny O, Bouillaud F, Ricquier D. Essential cis-acting elements in rat uncoupling protein gene are in an enhancer containing a complex retinoic acid response domain. J Biol Chem 271:31533-31542, 1996.
- Kopecky J, Clarke G, Enerbäck S, Spiegelman B, Kozak LP. Expression of the mitochondrial uncoupling protein gene from the aP2 gene promoter prevents genetic obesity. J Clin Invest 96:2914–2923, 1995.
- Kopecky J, Hodny Z, Rossmeisl M, Syrovy I, Kozak LP. I: Reduction of dietary obesity in aP2-Ucp transgenic mice: physiology and adipose tissue distribution. Am J Physiol 270:E768-E775, 1996.
- Enerback S, Jacobsson A, Simpson EM, Guerra C, Yamashita H, Harper M, Kozak LP. Mice lacking mitochondrial uncoupling protein are cold-sensitive but not obese. Nature 387:90-94. 1997.
- Bray GA, York DA. Hypothalamic and genetic obesity in experimental animals: an autonomic and endocrine hypothesis. Physiol Rev 59:719– 809, 1979.
- Bray GA, Fisler J, York DA. Neuroendocrine control of the development of obesity: understanding gained from studies of experimental animal models. Front Neuroendocrinol 11:128-181, 1990.
- Burns TW, Langley PE, Terry BE, Bylund DB, Hoffman BB, Tharp MD, Lefkowitz RJ, Garcia-Sainz JA, Fain JN. Pharmacological characterizations of adrenergic receptors in human adipocytes. J Clin Invest 67:467-475, 1981.
- Fain J, Garcia-Sainz J. Adrenergic regulation of adipocyte metabolism.
 J Lipid Res 24:945–966, 1983.
- Bahouth SW, Malbon CC. Subclassification of β-adrenergic receptors of rat fat cells: a re-evaluation. Mol Pharmacol 34:318-326, 1988.
- Arch JR, Ainsworth AT, Cawthorne MA, Piercy V, Sennitt MV, Thody VE, Wilson C, Wilson S. Atypical β-adrenoceptor on brown adipocytes as target for anti-obesity drugs. Nature 309:163–165, 1984.
- van Liefde I, van Witzenberg A, Vauquelin G. Multiple β-adrenergic receptor subclasses mediate the isoproteronol-induced lipolytic response in rat adipocytes. J Pharmacol Exp Ther 262:552-558, 1992.
- Collins S, Daniel KW, Rohlfs EM, Ramkumar V, Taylor IL, Gettys TW. Impaired expression and functional activity of the β₃- and β-adrenergic receptors in adipose tissue of congenitally obese (C57BL/6J ob/ob) mice. Mol Endocrinol 8:518-527, 1994.
- Galitzky J, Carpéné C, Bousquet-Mélou A, Berlan M, Lafontan M.
 Differential activation of β₁-, β₂- and β₃- adrenoceptors by catecholamines in white and brown adipocytes. Fundam Clin Pharmacol 9:324-331, 1995.
- Rohlfs EM, Daniel KW, Premont RT, Kozak LP, Collins S. Regulation
 of the uncoupling protein gene (Ucp) by β₁, β₂, β₃-adrenergic receptor
 subtypes in immortalized brown adipose cell lines. J Biol Chem
 270:10723-10732, 1995.
- Fève B, Emorine LJ, Lasnier F, Blin N, Baude B, Nahmias C Strosberg AD, Pairault J. Atypical β-adrenergic receptor in 3T3-F442A adipocytes: pharmacological and molecular relationship with the human β₃-adrenergic receptor. J Biol Chem 266:20329-20336, 1991.
- Dixon TM, Daniel KW, Farmer SR, Collins S. CAATT/enhancer binding protein-α is required for transcription of the β₃AR gene during adipogenesis. J Biol Chem 276:722-728, 2001.
- Wu Z, Rosen ED, Brun R, Hauser S, Adelmant G, Troy AE, McKeon C, Darlington GJ, Spiegelman BM. Cross-Regulation of C/EBPα and PPARγ controls the transcriptional pathway of adipogenesis and insulin sensitivity. Mol Cell 3:151-158, 1999.
- 31. El-Jack AK, Hamm JK, Pilch PF, Farmer SR. Reconstitution of insu-

- lin-sensitive glucose transport in fibroblasts requires expression of both PPARy and C/EBPa. J Biol Chem **274**:7946-7951, 1999.
- Laudat MH, Pairault J. An impaired response of adenylate cyclase to stimulation by epinephrine in adipocyte plasma membranes from genetically obese mice (ob/ob). Eur J Biochem 56:583-589, 1975.
- Dehaye JP, Winand J, Christophe J. Lipolysis and cyclic AMP levels in epididymal adipose tissue of obese hyperglycemic mice. Diabetologia 13:553-561, 1977.
- Shepherd RE, Malbon CC, Smith CJ, Fain JN. Lipolysis and adenosine 3[prime]:5[prime]-monophosphate metabolism in isolated white fat cells from genetically obese-hyperglycemic mice (ob/ob). J Biol Chem 252:7243-7248, 1977.
- Trayhurn P, James WPT. Thermoregulation and non-shivering thermogenesis in the genetically obese (ob/ob) mouse. Pfluegers Arch Eur J Physiol 373:189–193, 1978.
- Cox JE, Powley TL. Development of obesity in diabetic mice pair-fed with lean siblings. J Comp Physiol Psychol 91:347–358, 1977.
- Levin BE, Triscari J, Sullivan AC. Altered sympathetic activity during development of diet-induced obesity in rats. Am J Physiol 244:R347– R371 1083
- Levin BE, Triscari J, Sullivan AC. Relationship between sympathetic activity and diet-induced obesity in two rat strains. Am J Physiol 245:R364–R371, 1983.
- Takeuchi H, Matsuo T, Tokuyama K, Shimomura Y, Suzuki M. Dietinduced thermogenesis is lower in rats fed a lard diet than in those fed a high oleic acid safflower oil diet, a safflower oil diet or a linseed oil diet. J Nutr 125:920-925, 1994.
- Bégin-Heick N, Heick HMC. Increased response of adipose tissue of the *ob/ob* mouse to the action of adrenaline after treatment with thyroxin. Can J Physiol Pharmacol 55:1320-1329, 1977.
- Begin-Heick N. Adenylate cyclase in lean and obese (ob/ob) mouse epididymal white adipocytes. Can J Biochem Cell Biol 58:1033–1038, 1980.
- Muzzin P, Revelli JP, Kuhne F, Gocayne JD, McCombie WR, Venter JC, Giacobino JP, Fraser CM. An adipose tissue-specific β-adrenergic receptor: molecular cloning and down-regulation in obesity. J Biol Chem 266:24053-24058, 1991.
- Collins S, Daniel KW, Rohlfs EM. Depressed expression of adipocyte β-adrenergic receptors is a common feature of congenital and dietary obesity in rodents. Int J Obesity 23:669-677, 1999.
- Champigny O, Ricquier D, Blondel O, Mayers RM, Briscoe MG, Holloway BR. β3-Adrenergic receptor stimulation restores message and expression of brown-fat mitochondrial uncoupling protein in adult dogs. Proc Natl Acad Sci U S A 88:10774-10777, 1991.
- 45. Himms-Hagen J, Cui J, Danforth E Jr., Taatjes DJ, Lang SS, Waters BL, Claus TH. Effect of CL-316,243, a thermogenic β₃-agonist, on energy balance and brown and white adipose tissues in rats. Am J Physiol 266:R1371–R1382, 1994.
- Collins S, Daniel KW, Petro AE, Surwit RS. Strain-specific response to β₃-adrenergic receptor agonist treatment of diet-induced obesity in mice. Endocrinology 138:405–413, 1997.
- 47. Fisher MH, Amend AM, Bach TJ, Barker JM, Brady EJ, Candelore MR, Carroll D, Cascieri MA, Chiu SH, Deng L, Forrest MJ, Hegarty-Friscino B, Guan XM, Hom GJ, Hutchins JE, Kelly LJ, Mathvink RJ, Metzger JM, Miller RR, Ok HO, Parmee ER, Saperstein R, Strader CD, Stearns RA, MacIntyre DE, Thompson GM, Tota L, Vicario PP, Weber AE, Woods KW, Wyvratt MJ, Zafian PT, MacIntyre DE. A selective human β3 adrenergic receptor agonist increases metabolic rate in rhesus monkeys. J Clin Invest 101:2387–2393, 1998.
- 48. Sasaki N, Uchida E, Niiyama M, Yoshida T, Saito M. Anti-obesity effects of selective agonists to the β3-adrenergic receptor in dogs. II. Recruitment of thermogenic brown adipocytes and reduction of adiposity after chronic treatment with a β3-adrenergic agonist. J Vet Med Sci 60:465-469, 1998.
- Guerra C, Koza RA, Yamashita H, Walsh K, Kozak LP. Emergence of brown adipocytes in white fat in mice is under genetic control: effects on body weight and adiposity. J Clin Invest 102:412-420, 1998.

- Liggett S, Freedman NJ, Schwinn DA, Lefkowitz RJ. Structural basis for receptor subtype-specific regulation revealed by a chimeric β₃-/ β₂-adrenergic receptor. Proc Natl Acad Sci U S A 90:3665–3669, 1993.
- Cao W, Luttrell LM, Medvedev AV, Pierce KL, Daniel KW, Dixon TM, Lefkowitz RJ, Collins S. Direct binding of activated c-Src to the β₃-adrenergic receptor is required for MAP kinase activation. J Biol Chem 275:38131–38134, 2000.
- Lefkowitz RJ. G protein-coupled receptors. III. New roles for receptor kinases and β-arrestins in receptor signaling and desensitization. J Biol Chem 273:18677–18680, 1998.
- Largis EE, Burns MG, Muenkel HA, Dolan JA, Claus TH. Antidiabetic and antiobesity effects of a highly selective β3-adrenoceptor agonist (CL 316,243). Drug Dev Res 32:69-76, 1994.
- 54. Collins S, Surwit RS. The β-adrenergic receptors and the control of adipose tissue metabolism and thermogenesis. In: Means AR, Ed. Recent Progress in Hormone Research. Bethesda, MD: The Endocrine Society, pp309–328, 2001.
- 55. Rodbell M. On the mechanism of activation of fat cell adenylate cyclase by guanine nucleotides: an explanation for the biphasic inhibitory and stimulatory effects of the nucleotides and the role of hormones. J Biol Chem 250:5826-5834, 1975.
- Cooper D, Schlegel W, Lin M, Rodbell M. The fat cell adenylate cyclase system. J Biol Chem 254:8927–8931, 1979.
- Murayama T, Ui M. Loss of the inhibitory function of the guanine nucleotide regulatory component of adenylate cyclase due to its ADP ribosylation by islet-activating protein, pertussins toxin, in adipocyte membranes. J Biol Chem 258:3319–3326, 1983.
- 58. Bloom JD, Dutia MD, Johnson BD, Wissner A, Burns MG, Largis EE, Dolan JA, Claus TH. Disodium (R,R)-5-[2[[2-(3-chlorophenyl)-2-hydroxyethyl]-amino]propyl]-1,3-benzodioxole-2,2-dicarboxylate (CL 316,243): a potent β-adrenergic agonist virtually specific for β₃ receptors: a promising antidiabetic and antiobesity agent. J Med Chem 35:3081-3084, 1992.
- Begin-Heick N. β₃-adrenergic activation of adenylyl cyclase in mouse white adipocytes: modulation by GTP and effect of obesity. J Cell Biochem 58:464–473, 1995.
- Soeder KJ, Snedden SK, Cao W, Della Rocca GJ, Daniel KW, Luttrell LM, Collins, S. The β₃-adrenergic receptor activates mitogenactivated protein kinase in adipocytes through a Gi-dependent mechanism. J Biol Chem 274:12017–12022, 1999.
- Cao W, Medvedev AV, Daniel KW, Collins S. β-Adrenergic activation of p38 MAP kinase in adipocytes: cAMP induction of the uncoupling protein-1 (UCP1) gene requires p38 MAP kinase. J Biol Chem 276:27077-27082, 2001.
- 62. Fleury C, Neverova M, Collins S, Raimbault S, Champigny O, Levi-Meyrueis C, Bouillaud F, Seldin MF, Surwit RS, Ricquier D, Warden CH. Uncoupling protein-2: a novel gene linked to obesity and hyperinsulinemia. Nat Genet 15:269–272, 1997.
- 63. Gimeno RE, Dembski M, Weng X, Deng N, Shyjan AW, Gimeno CJ, Iris F, Ellis SJ, Woolf EA, Tartaglia LA. Cloning and characterization of an uncoupling protein homolog: a potential molecular mediator of human thermogenesis. Diabetes 46:900–906, 1997.
- 64. Boss O, Samec S, Paoloni-Giacobino A, Rossier C, Dulloo A, Seydoux J, Muzzin P, Giacobino JP. Uncoupling protein-3: a new member of the mitochondrial carrier family with tissue-specific expression. FEBS Lett 408:39-42, 1997.
- Vidal-Puig A, Solanes G, Grujic D, Flier JS, Lowell BB. UCP3: an uncoupling protein homologue expressed preferentially and abundantly in skeletal muscle and brown adipose tissue. Biochem Biophys Res Commun 235:79

 –82, 1997.
- 66. Gong D, He Y, Karas M, Reitman M. Uncoupling protein-3 is a mediator of thermogenesis regulated by thyroid hormone, β3adrenergic agonists, and leptin. J Biol Chem 272:24129-24132, 1997.
- 67. Fleury C, Sanchis D. The mitochondrial uncoupling protein-2: current status. Int J Biochem Cell Biol 31:1261-1278, 1999.

- 68. Boss O, Muzzin P, Giacobino JP. The uncoupling proteins, a review. Eur J Endocrinol 139:1-9, 1998.
- Ricquier D, Fleury C, Larose M, Sanchis D, Pecqueur C, Raimbault S, Gelly C, Vacher D, Cassard-Doulcier AM, Levi-Meyrueis C, Champigny O, Miroux B, Bouillaud F. Contributions of studies on uncoupling proteins to research on metabolic diseases. J Intern Med 245:637-642, 1999.
- Surwit RS, Wang S, Petro AE, Sanchis D, Raimbault S, Ricquier D, Collins S. Diet-induced changes in uncoupling proteins in obesityprone and obesity-resistant strains of mice. Proc Natl Acad Sci U S A 95:4061–4065, 1998.
- Boss O, Samec S, Dulloo A, Seydoux J, Muzzin P, Giacobino J. Tissue-dependent upregulation of rat uncoupling protein-2 expression in response to fasting or cold. FEBS Lett 142:111-114, 1997.
- Weigle D, Selfridge L, Schwartz M, Seeley R, Cummings D, Havel P, Kuijper JL, BeltrandelRio H. Elevated free fatty acids induce uncoupling protein 3 expression in muscle. Diabetes 47:298-302, 1998.
- Samec S, Seydoux J, Dulloo AG. Role of UCP homologues in skeletal muscles and brown adipose tissue: mediators of thermogenesis or regulators of lipids as fuel substrate? FASEB J 12:715-24, 1998.
- Samec S, Seydoux J, Dulloo AG. Skeletal muscle UCP3 and UCP2 gene expression in response to inhibition of free fatty acid flux through mitochondrial β-oxidation. Pflugers Arch 438:452-457, 1999.
- Cadenas S, Buckingham JA, Samec S, Seydoux J, Din N, Dulloo AG, Brand MD. UCP2 and UCP3 rise in starved rat skeletal muscle but mitochondrial proton conductance is unchanged. FEBS Lett 462:257– 260, 1999.
- Aubert J, Champigny O, Saint-Marc P, Negrel R, Collins S, Ricquier D, Ailhaud G. Up-regulation of UCP-2 gene expression by PPAR agonists in preadipose and adipose cells. Biochem Biophys Res Commun 238:606-611, 1997.
- Camirand A, Marie V, Rabelo R, Silva JE. Thiazolidinediones stimulate uncoupling protein-2 expression in cell lines representing white and brown adipose tissues and skeletal muscle. Endocrinology 139:428-431, 1998.
- Viguerie-Bascands N, Saulnier-Blache JS, Dandine M, Dauzats M, Daviaud D, Langin D. Increase in uncoupling protein-2 mRNA expression by BRL49653 and bromopalmitate in human adipocytes. Biochem Biophys Res Commun 256:138–141 1999.
- Medvedev AV, Snedden SK, Raimbault S, Ricquier D, Collins S. Transcriptional regulation of the mouse uncoupling protein-2 gene: double E-box motif is required for peroxisome proliferator-activated receptor-γ-dependent activation. J Biol Chem 276:10817-10823, 2001.
- Gong DW, Monemdjou S, Gavrilova O, Leon LR, Marcus-Samuels B, Chou CJ, Everett C, Kozak LP, Li C, Deng C, Harper ME, Reitman ML. Lack of obesity and normal response to fasting and thyroid hormone in mice lacking uncoupling protein-3. J Biol Chem 275:16251– 16257, 2000.
- Vidal-Puig AJ, Grujic D, Zhang CY, Hagen T, Boss O, Ido Y, Szczepanik A, Wade J, Mootha V, Cortright R, Muoio DM, Lowell BB. Energy metabolism in uncoupling protein 3 gene knockout mice. J Biol Chem 275:16258-16266, 2000.
- 82. Clapham JC, Arch JR, Chapman H, Haynes A, Lister C, Moore GB, Piercy V, Carter SA, Lehner I, Smith SA, Beeley LJ, Godden RJ, Herrity N, Skehel M, Changani KK, Hockings PD, Reid DG, Squires SM, Hatcher J, Trail B, Latcham J, Rastan S, Harper AJ, Cadenas S, Buckingham JA, Brand MD, Abuin A. Mice overexpressing human uncoupling protein-3 in skeletal muscle are hyperphagic and lean. Nature 406:415-418, 2000.
- Stuart JA, Harper JA, Brindle KM, Jekabsons MB, Brand MD. Physiological levels of mammalian uncoupling protein 2 do not uncouple yeast mitochondria. J Biol Chem 276:18633-18639, 2001.
- 84. Negre-Salvayre A, Hirtz C, Carrera G, Cazenave R, Troly M, Salvayre R, Penicaud L, Casteilla L. A role for uncoupling protein-2 as a regulator of mitochondrial hydrogen peroxide generation. FASEB J 11:809-815, 1997.

- Lee FY, Li Y, Zhu H, Yang S, Lin HZ, Trush M, Diehl AM. Tumor necrosis factor increases mitochondrial oxidant production and induces expression of uncoupling protein-2 in the regenerating mice liver. Hepatology 29:677-687, 1999.
- 86. Arsenijevic D, Onuma H, Pecqueur C, Raimbault S, Manning BS, Miroux B, Couplan E, Alves-Guerra MC, Goubern M, Surwit R, Bouillaud F, Richard D, Collins S, Ricquier D. Disruption of the uncoupling protein-2 gene in mice reveals a role in immunity and reactive oxygen species production. Nat Genet 26:435–439, 2000.
- Ricquier D, Bouillaud F. The uncoupling protein homologues: UCP1, UCP2, UCP3, StUCP and AtUCP. Biochem J 345:161-179, 2000.
- 88. Cortez-Pinto H, Yang SQ, Lin HZ, Costa S, Hwang CS, Lane MD, Bagby G, Diehl AM. Bacterial lipopolysaccharide induces uncoupling protein-2 expression in hepatocytes by a tumor necrosis factor-α-

- dependent mechanism. Biochem Biophys Res Commun 251:313-319, 1998.
- 89. Larrouy D, Laharrague P, Carrera G, Viguerie-Bascands N, Levi-Meyrueis C, Fleury C, Pecqueur C, Nibbelink M, Andre M, Casteilla L, Ricquier D. Kupffer cells are a dominant site of uncoupling protein 2 expression in rat liver. Biochem Biophys Res Commun 235:760–764, 1997.
- Rothwell NJ, Stock MJ. A role for brown adipose tissue in dietinduced thermogenesis. Nature 281:31–35. 1979.
- Lehmann JM, Moore LB, Smith-Oliver TA, Wilkison WO, Willson TM, Kliewer SA. An antidiabetic thiazolidinedione is a high affinity ligand for the nuclear receptor PPARy. J Biol Chem 270:12953– 12956. 1995.
- 92. Collins S, Surwit RS. Pharmacologic manipulation of *ob* expression in a dietary model of obesity. J Biol Chem **271**:9437–9440. 1996.