Arachidonic Acid Metabolites Mediate the Radiation-Induced Increase in Glomerular Albumin Permeability

Mukut Sharma,*,1 Ellen T. McCarthy,* Ram Sharma,* Brian L. Fish,† Virginia J. Savin,* Eric P. Cohen,* and John E. Moulder†

*Division of Nephrology and the Kidney Disease Center, Department of Medicine, Medical College of Wisconsin, Milwaukee, Wisconsin 53226; and †Department of Radiation Oncology, Medical College of Wisconsin, Milwaukee, Wisconsin 53226

Radiation-induced renal injury is characterized by proteinuria, hypertension, and progressive decline in renal function. We have previously shown that in vivo or in vitro irradiation of glomeruli with a single dose of radiation (9.5 Gy) increases glomerular albumin permeability (Palb) within 1 hr. The current studies tested the hypothesis that this early radiation-induced increase in Palb is caused by the release of arachidonic acid and by the generation of specific arachidonic acid metabolites. Glomeruli obtained from WAG/Rij/MCW rats and cultured rat glomerular epithelial and mesangial cells were studied after irradiation (9.5 Gy, single dose). Arachidonic acid release and eicosanoid synthesis by glomeruli or cultured glomerular cells were measured after irradiation, and the effect of inhibitors of phospholipase A2 (PLA2) and cyclooxygenase (COX) on the irradiation-induced increase in Palb was assessed. Arachidonic acid release was demonstrated within 10 mins of irradiation of isolated glomeruli and monolayer cultures of glomerular epithelial and mesangial cells. Prostaglandin $F_{2\alpha}$ (PGF_{2\alpha}) and PGE₂ release was increased after irradiation of isolated glomeruli. Blocking arachidonic acid release or COX activity before irradiation completely prevented the increase in Palb. COX inhibition immediately after irradiation also diminished the radiation-induced increase in Palb. We conclude that arachidonic acid and its COX metabolites play an essential role in the early cellular changes that lead to the radiation-induced increase in Palb. Understanding of the early epigenetic effects of irradiation may lead to new intervention strategies against radiationinduced injury of normal tissues. Exp Biol Med 231:99-106, 2006

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Introduction

Radiation-induced renal injury (radiation nephropathy) is characterized by proteinuria, hypertension, and progressive decline in renal function (1). It is seen in the setting of total body irradiation (TBI) in preparation for bone marrow transplantation (2) but can also be observed after external beam therapeutic irradiation (3) and after therapeutic use of radiolabeled biologicals (4). Additionally, renal injury could occur as a result of radiation accidents or radiologic terrorism (5, 6). Radiation nephropathy develops months to years after irradiation, but the early cellular events that contribute to the appearance of clinical symptoms such as overt proteinuria are not clearly understood (1, 2, 7).

We have previously shown that radiation-induced renal failure in the rat can occur as early as 8 months after a single dose of 8.7 Gy and that renal dysfunction (e.g., elevated BUN) can be observed by 7 months after a single dose of 6.5 Gy (8). In the same strain of rats, in vivo irradiation with a single dose of 9.5 Gy increases glomerular albumin permeability (Palb) at 1 hr, even though rats irradiated in this protocol developed proteinuria only after 4 weeks (2). Palb is increased in a dose-dependent manner in glomeruli irradiated ex vivo at 5.5-9.5 Gy; this dose-dependent increase in P_{alb} is correlated (Kendal $\tau = 0.84$, P = 0.04) with the azotemia that develops later in rats irradiated in this protocol (7). This increase in Palb may serve as an indicator of initial changes in the glomerular filtration barrier and of the chronic renal injury that appears following renal or total body irradiation (1-7).

Ionizing radiation increases the release of arachidonic acid from membrane phospholipids and the generation of eicosanoids through up-regulation of phospholipase A₂ (PLA₂) (9) and cyclooxygenase (COX) (10). Eicosanoids

¹ To whom correspondence should be addressed at Rm. M4040, Nephrology/CVC/MEB, Medical College of Wisconsin, 8701 Watertown Plank Road, Milwaukee, WI 53226. E-mail: msharma@mcw.edu

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are known for their vascular effects and for their role in injury and inflammation (11–13). We hypothesized that radiation-induced release of arachidonic acid and/or radiation-induced generation of eicosanoids causes increased glomerular albumin permeability (P_{alb}). Results of our initial test of this hypothesis and its relevance to the prevention and mitigation of radiation-induced renal disease are presented in this communication.

Materials and Methods

Experimental Model. WAG/Rij/MCW rats were maintained on sterilized rat chow in a moderate-security barrier facility at the Biomedical Resource Center of the Medical College of Wisconsin (MCW). In all cases, kidneys were removed via abdominal incision after the animals were anesthetized using halothane (Halocarbon Laboratories, River Edge, NJ). Animal care was in accordance with NIH guidelines, and the Animal Care and Use Committee at the Medical College of Wisconsin approved of all protocols. All irradiations were carried out with a single dose of 9.5 Gy. This radiation dose induces a robust (and near maximal) effect on glomerular permeability (P_{alb}) (7) but is low enough that long-term *in vivo* studies can be carried out in a bone transplant model without causing acute radiation injury to other organ systems (8).

Arachidonic Acid Release from Isolated Glo**meruli.** Kidneys were removed from 8- to 9-week-old rats, and glomeruli were isolated in RPMI-1640 (Gibco, Invitrogen Corporation, Carlsbad, CA) by established sieving techniques (7). Glomeruli from both kidneys were pooled to obtain one sample from each rat. Isolated glomeruli were incubated for 60 mins with 16.7 kBq [3H]arachidonic acid (Amersham Pharmacia Biotech, Piscataway, NJ) in RPMI-1640 (Gibco) containing 0.1% bovine serum albumin (BSA, Sigma Chemical Company, St. Louis, MO). The incubator atmosphere was maintained at 37°C and 5% CO₂. Glomeruli were washed with fresh medium without radioactivity and incubated for 30 mins to allow distribution and equilibration of the labeled arachidonic acid. At the end of the equilibration period, glomeruli were washed with RPMI-1640.

The experimental group was irradiated with 9.5 Gy in a ¹³⁷Cs γ-ray generator (Model 137; JL Shepherd, San Fernando, CA). Sham-irradiated glomeruli were used as the control group. Glomeruli incubated with 5 μg/ml calcium ionophore A-23187 (Sigma Chemical Company) served as the positive control. Medium was collected at 10 or 60 mins posttreatment and mixed with 6 volumes of ethyl acetate acidified by 1 *M* formic acid (0.08% v/v). Radioactivity in the ethyl acetate extract was measured using a liquid scintillation counter (LKB RackBeta 1214, Pharmacia, Wallac Oy, Finland). Total protein in glomerular preparation and cells was determined by Lowry's method (14) using a reagent kit (Bio-Rad, Hercules, CA). The

measured radioactivity was expressed as counts per minute (cpm)/mg total protein.

Monolayers of Glomerular Epithelial Cells and Mesangial Cells. Rat glomerular epithelial cells (passage 10-14) were grown to confluence on 10-cm tissue culture petri dishes in K1 medium containing 2% Nu Serum (Collaborative Biomedical Products, Becton Dickinson, Bedford, MA). Rat mesangial cells (passage 10-14) were grown to confluence in RPMI-1640 containing 15% fetal bovine serum (FBS, Gibco) and maintained in RPMI-1640 containing 10% FBS. Confluent cultures were washed with serum-free RPMI-1640 and incubated with 10⁶ cpm [3H]arachidonic acid in RPMI-1640 containing 0.1% BSA for 60 min. Cells were washed with fresh radioactivity-free RPMI-1640 and incubated for 30 mins to allow equilibration of the radioactive arachidonic acid. At the end of the equilibration period, cells were washed with fresh RPMI-1640, divided into three groups, and irradiated with 9.5 Gy (137Cs γ-ray), sham treated, or incubated with calcium ionophore (5 µg/ml). Each petri dish was used as one sample. Aliquots of the media were collected at 10 mins after irradiation. Released radioactivity was extracted with acidified ethyl acetate and measured as described in the previous section.

Competitive Enzyme Immunoassay (EIA) of Prostaglandin E_2 (PGE₂) and $F_{2\alpha}$ (PGF_{2\alpha}). We have previously shown that PGE₂ or PGF_{2\alpha} induced an increase in P_{alb} (14) and therefore selected these two COX products of arachidonic acid for analysis in the present studies. Isolated glomeruli were irradiated or sham-irradiated as described above. Aliquots of media at 5 or 15 mins after irradiation were processed for EIA of PGE₂ or PGF_{2\alpha} according to the manufacturer's instructions (Cayman Chemical Company, Ann Arbor, MI). Total protein was determined by Lowry's method (14), and the concentration of eicosanoids expressed as picograms per milligram protein.

Treatment of Isolated Glomeruli with Inhibitors. Isolated glomeruli were incubated with the PLA₂ inhibitor quinacrine (10 µM, Sigma Chemical Company) or a COX inhibitor (see below) and irradiated with 9.5 Gy (137 Cs γ -rays), and P_{alb} was determined at 60 mins postirradiation. The COX inhibitors assessed were indomethacin (1 µM, Sigma Chemical Company), acetylsalicylic acid (aspirin, 1.25 mM, Sigma Chemical Company), and ibuprofen ($[\pm]$ -2-[4-isobutylphenyl]-propionic acid, 20 μM , Cayman Chemical) as they are the most commonly used experimental and clinical COX inhibitors. We have previously shown that indomethacin can effectively prevent an increase in Palb generated by other agents (15). Inhibitortreated glomeruli without irradiation were used as control. Sham-irradiated control glomeruli were preincubated with 10 µL of ethanol, the vehicle. Irradiated glomeruli were used as positive control. All incubations were carried out at 37°C for 10 mins in 1 ml final volume made with a physiological buffer containing 5% BSA.

In a separate series of experiments, isolated glomeruli were incubated with ibuprofen (20 μM) for 30 mins before or immediately after (within 2 mins) irradiation (9.5 Gy ^{137}Cs γ -rays), and P_{alb} was measured after 60 mins. Control groups consisted of glomeruli treated with or without ibuprofen alone. This experimental design was used to allow us to assess whether the COX inhibitors were working via a "mitigation" mechanism as opposed to a "protection" mechanism. We selected ibuprofen instead of indomethacin and acetylsalicylic acid because ibuprofen inhibits COX-1 and COX-2 at about equimolar concentrations (16), and it is the most commonly used nonsteroidal antiinflammatory agent. Such distinction is not critical for potential applications in therapeutic irradiation but is critical for application to radiation accidents or radiologic terrorism that may affect a very large number of people in a short time (5, 6).

Measurement of Glomerular Convectional Permeability to Albumin. The in vitro assay of albumin permeability indicates changes in the characteristics of the glomerular filtration barrier that restricts macromolecules (colloids) to plasma. The capillary plasma pressure attributed to macromolecules is called oncotic pressure or colloidal osmotic pressure. Lowering the extracapillary oncotic pressure can generate an oncotic gradient that would result in a movement of fluid into the capillary. The volume response of glomerular capillaries to an oncotic gradient was measured as previously described (7). In brief, following the experimental manipulations as described above glomeruli were transferred to a glass cover slip coated with poly-L-lysine (1 mg/ml, Sigma Chemical Company) and observed using videomicroscopy before and about 1 min after the medium containing 5% BSA was replaced by medium containing 1% BSA. This exchange of medium produces an oncotic gradient across the glomerular capillary wall and results in a net influx of fluid and an increase in glomerular volume. The increase in volume (ΔV) of each glomerulus in response to an oncotic gradient was expressed as: $\Delta V = (V_{\text{final}} - V_{\text{initial}})/V_{\text{initial}} \times 100\%$.

There is a direct relationship between the increase in glomerular volume (ΔV) and the oncotic gradient ($\Delta \Pi$) applied across the capillary wall. We used this principle to calculate σ_{alb} , using the ratio of ΔV of experimental to ΔV of control glomeruli in response to identical oncotic gradients: $\sigma_{alb} = \Delta V_{experimental}/\Delta V_{control}$.

Convectional albumin permeability (P_{alb}) was defined as $(1 - \sigma_{alb})$ to describe the movement of albumin consequent to water flow. When σ_{alb} is zero, albumin moves at the same rate as water, and P_{alb} is 1.0. Alternatively, when σ_{alb} is 1.0, albumin cannot cross the membrane with water, and P_{alb} is zero.

Statistical Analysis. P_{alb} values are expressed as mean \pm SEM. Values of groups were compared using t test. Significance of difference between groups was expressed as P value. Selected data were rechecked and reconfirmed by nonparametric analysis using the Mann-Whitney test.

Results

Release of Arachidonic Acid from Rat Glomeruli. Irradiation of glomeruli with a 9.5-Gy single dose resulted in increased release of arachidonic acid and its metabolites at 10 mins (both P < 0.001; Fig. 1). Radioactivity in the medium from the calcium ionophoretreated positive control group at 10 mins and 60 mins was comparable to that observed in the irradiated group. The released arachidonic acid in the irradiated and ionophoretreated groups at 60 mins was 57% and 66% of that at 10 mins. The sham-irradiated control groups at 10 and 60 mins were not different from each other (P > 0.1; Fig. 1).

Release of Arachidonic Acid from Irradiated Monolayers of Glomerular Epithelial Cells and Mesangial Cells. Irradiation of cultured glomerular epithelial cells (Fig. 2A) and mesangial cells (Fig. 2B) with a 9.5-Gy single dose resulted in increased release of arachidonic acid and its metabolites at 10 mins (both P < 0.001). Irradiation caused a 2.2-fold increase in released radioactivity from glomerular epithelial cells and mesangial cells, whereas the ionophore-induced increase in the released radioactivity was 5.2- and 7.4-fold from glomerular epithelial cells and mesangial cells, respectively (Fig. 2).

Release of PGE₂ and PGF_{2 α} from Irradiated Glomeruli. PGE₂ synthesized by irradiated glomeruli was 45% higher than that by control glomeruli at 15 mins (P < 0.001), but the difference was not significant (P > 0.10) at 5 mins (Fig. 3A). PGF_{2 α} synthesized by irradiated glomeruli at 15 mins showed a 145% increase over the control group (P < 0.001), but the difference was not significant (P > 0.10) at 5 mins (Fig. 3B).

Effect of Inhibitors of Arachidonic Acid Metabolism on the Irradiation-Induced Increase in P_{alb} . The P_{alb} of groups treated with quinacrine (Q) alone, indomethacin (I) alone, or acetylsalicylic acid (A) alone were not different from that of the sham-irradiated control (all P > 0.10; Fig. 4). Pretreatment with quinacrine, indomethacin, or acetylsalicylic acid blocked the radiation-induced increase in P_{alb} (all P < 0.001 vs. the Rad alone group; Fig. 4).

Ibuprofen also protected the glomerular permeability barrier from radiation injury (Fig. 5). Addition of ibuprofen before as well as immediately after irradiation alleviated the radiation-induced increase in $P_{\rm alb}$ (both P < 0.001; Fig. 5), but treatment before irradiation was more effective than treatment after irradiation.

Discussion

We demonstrate for the first time that a radiation-induced increase in glomerular albumin permeability can be prevented by inhibiting the cyclooxygenase pathway of arachidonic acid metabolism. Irradiation causes rapid release of arachidonic acid from both isolated intact glomerula and cultured glomerular cells. Furthermore, glomerular synthesis of PGE_2 and $PGF_{2\alpha}$ was increased

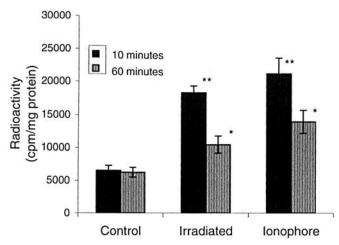
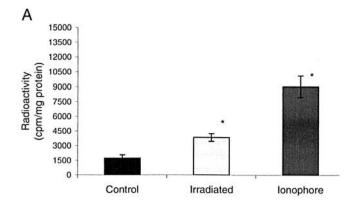


Figure 1. Release of arachidonic acid from irradiated rat glomeruli. Isolated glomeruli were preincubated with [3 H]arachidonic acid for 1 hr. After a 30-min equilibration, glomeruli were irradiated (9.5 Gy, single dose), incubated with calcium ionophore (5 μ g/ml), or shamirradiated. Arachidonic acid and metabolites were extracted from media at 10 or 60 mins posttreatment, and total radioactivity was measured. Glomeruli from two kidneys of the same animal were pooled to obtain one sample (8 samples in each group). Data are shown as the means \pm the standard error. Significant differences from pooled sham-irradiated controls are marked (*P < 0.01 and **P < 0.001).

immediately following irradiation. Inhibition of the release and metabolism of arachidonic acid blocked the increase in P_{alb} , supporting our hypothesis that one or more eicosanoids mediate the early glomerular changes that precedes proteinuria in a model of radiation nephropathy.

Proteinuria is associated with several cardiovascular diseases and is one of the major problems in modern medicine. It reflects a fairly advanced stage of renal damage that is compounded by the impact of the protein in the glomerular filtrate on tubular function and by the effects of progressive increase in urinary protein (17–19). Clinical radiation nephropathy is also characterized by proteinuria, azotemia, and hypertension that develop months to years after irradiation (3, 20). In a rat model of radiation nephropathy, overt proteinuria develops only after about 4 weeks (1). The latency before the development of proteinuria in radiation nephropathy offers an opportunity to explore the initial changes that lead to pathophysiologic symptoms and organ failure so as to intervene in disease progression.

We have reported that although P_{alb} increases within 1 hr after irradiation, receptor-mediated tubular binding of albumin deteriorates only several weeks later (21), indicating a difference in the effect of radiation on glomeruli and tubules. We have used *in vitro* determination of P_{alb} as a marker of preproteinuric glomerular changes in the Zucker obese rats (22), puromycin aminonucleoside (PAN)-induced nephrosis (23), dietary salt-induced proteinuria with hypertension (24), and recurrent focal segmental glomerulosclerosis (FSGS) (25). In each of these models, glomerular injury (as indicated by increased P_{alb}) precedes the onset of



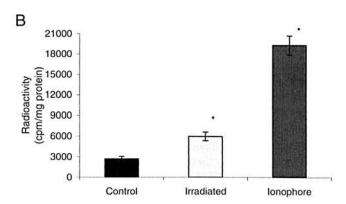
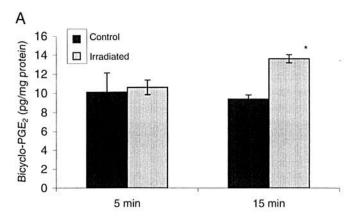


Figure 2. Release of arachidonic acid from irradiated monolayers of glomerular epithelial cells and mesangial cells. Confluent cultures of glomerular epithelial cells (A) or mesangial cells (B) were preincubated with [3 H]arachidonic acid for 1 hr. After equilibration for 30 mins, cells were irradiated (9.5 Gy, single dose), incubated with calcium ionophore (5 µg/ml), or sham-treated. Arachidonic acid and metabolites were extracted from the medium at 10 mins posttreatment. (A) Total radioactivity released from control (n = 8), irradiated (n = 7), and ionophore-treated (n = 4) experiments with glomerular epithelial cells. (B) Total radioactivity released from control (n = 8), irradiated (n = 8), and ionophore-treated (n = 5) mesangial cells. Data are shown as the means \pm the standard error. Significant differences from sham-irradiated controls are marked ($^*P < 0.001$).

proteinuria, suggesting that *in vitro* glomerular permeability is a sensitive and reliable marker of the initial injury to the glomerular filtration barrier. Identification of early markers of glomerular and tubular injury is essential for effective management and treatment of radiation-induced renal dysfunction (6).

The immediate increase in glomerular permeability following irradiation reflects rapid metabolic events affecting the structure and function of the glomerular protein permeability barrier and is less likely to reflect altered gene expression at this time point. Such epigenetic processes may trigger molecular pathways that alter biological characteristics in a tissue/cell-specific manner and result in gene activation (26). We postulate that immediate responses to injurious agents or environmental changes may involve membrane- and cytoskeleton-associated molecules and



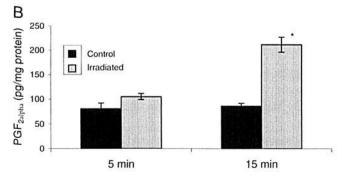


Figure 3. PGE₂ and PGF_{2 α} release from isolated irradiated glomeruli. Isolated glomeruli were irradiated (9.5 Gy, single dose) or sham-irradiated. Released eicosanoids were extracted from media at 5 or 15 mins after irradiation, and PGE₂ (A) and PGF_{2 α} (B) were determined by EIA. Glomeruli from two kidneys of the same animal were pooled to obtain one sample (n = 3 samples in each group). Data are shown as the means \pm standard error. Significant differences from concurrent sham-irradiated controls (*P< 0.001).

signaling pathways. Arachidonic acid, a polyunsaturated essential fatty acid in membrane phospholipids, and its metabolites may play key roles in the epigenetic events that lead to increased permeability.

In the present study a single dose of ionizing radiation increased the release of arachidonic acid from glomeruli (Fig. 1) as well as from glomerular cells (Fig. 2). The finding that arachidonic acid in the medium was lower at 60 mins after irradiation than at 10 mins suggests a dynamic exchange of arachidonic acid between the medium and glomerular cells in the intervening period. The immediate release of arachidonic acid by glomeruli and glomerular cells after irradiation is in marked contrast to the response of cultured HL-60 cells (a leukemia cell line) in which arachidonic acid release occurred only at about 30 mins after irradiation (27). The difference between the radioactivity released from whole glomeruli and cultured glomerular cells indicates differences between cells within the glomerulus. Thus, the release of arachidonic acid in response to irradiation appears to occur in a tissue- or cellspecific manner that depends on the composition of the

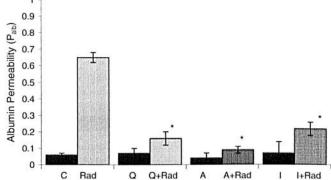


Figure 4. Effect of inhibitors of arachidonic acid metabolism on the radiation-induced increase in P_{alb} . Isolated glomeruli preincubated with quinacrine (Q), indomethacin (I), or acetylsalicylic acid (A) were either used as control (Q, I, and A, 15 samples in each group) or after irradiation (Q+Rad, I+Rad, and A+Rad, 15 samples in each group). Sham-treated glomeruli were used as baseline control (C, 30 samples), and irradiated glomeruli without drug treatment were used as positive control (Rad, 20 samples). P_{alb} was determined at 1 hr postirradiation. Results are shown as the means \pm the standard error. Significant differencest from animals treated with radiation alone are marked (*P < 0.001).

membrane. The characteristic features of cellular membrane such as conductance, fluidity, and permeability are determined by its protein and lipids, which, in turn, are determined by the chain length and the degree of unsaturation of fatty acids (29). Thus, the radiation-induced changes in the membrane composition and the increased levels of free arachidonic acid are likely to affect several cellular events. For example, arachidonic acid activates protein kinase C (PKC) in vitro (30–32) and thus influences the signaling events mediated by PKC. These changes may occur without invoking DNA-dependent synthesis of response molecules. Our findings suggest a role for arachidonic acid and its metabolites in the early cellular signaling changes that lead to increased P_{alb}.

Nonesterified free arachidonic acid is metabolized through the COX, lipoxygenase, and cytochrome P-450 pathways into several eicosanoids that are important in vascular regulation and tissue response to injury (Fig. 6). Earlier reports indicate that total body irradiation (TBI) of mice or rats increases urinary prostaglandins and thromboxane B_2 , indicating a role for these metabolites in the immediate response to radiation (33–37). Changes in urinary PGE₂ after TBI were found to follow a biphasic pattern (38). Some of eicosanoids may be of systemic origin because thoracic shielding of rats prevented urinary excretion of thromboxane B_2 (39) following TBI. Our results show that irradiation induces increased levels of PGE₂ and PGF_{2 α} in glomeruli (Fig. 4).

The renal prostaglandins and thromboxanes act as autacoids to modulate renal hemodynamics and may have direct effects on the glomerular filtration barrier. We have recently shown that PGE_2 and $PGF_{2\alpha}$ cause increased glomerular albumin permeability of isolated glomeruli in the absence of neurohumoral or hemodynamic factors (15).

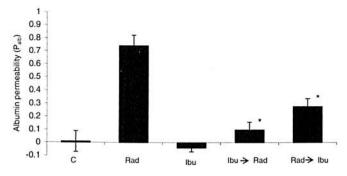


Figure 5. Effect of ibuprofen on the radiation-induced increase in P_{alb} . Glomeruli were sham-irradiated (C), given irradiation only (Rad), given ibuprofen only (Ibu), preincubated with ibuprofen for 30 mins followed by irradiation (Ibu \rightarrow Rad), or irradiated and then incubated with ibuprofen for 60 mins (Rad \rightarrow Ibu) (15 samples in each group). Data are shown as the means \pm standard error. Significant differences from animals treated with irradiation only are marked (*P < 0.001).

Further, PGE_2 increases depolymerization of actin filaments in podocytes, indicating cytoskeletal changes that may alter glomerular filtration barrier function (40). Cellular effects of prostaglandins are receptor mediated. For example, PGE_2 binds to four E-prostanoid receptors (EP1–EP4), each of which preferentially couples to a different signal transduction pathway, including stimulation of cAMP generation by the EP2 and EP4 receptors; inhibition of cAMP generation via G_i by EP3 receptors, and activation of phosphatidylinositol hydrolysis by EP1 receptors. $PGF_{2\alpha}$ has been shown to bind to the FP receptor and cause mesangial cell contraction and proliferation through the phosphoinositide–phopholipase C and protein kinase C signaling mechanisms (41, 42).

Release of arachidonic acid from membrane phospholipids by PLA₂ and its metabolism into PGE₂ and PGF_{2 α} by COX-1 and -2 increases following injury (28, 37, 43). Present studies also indicate the role of these enzymes in the radiation-induced increase in glomerular albumin permeability. Radiation-induced increase in PLA₂ and COX-2 has been demonstrated in cells such as astrocyte and microglial cultures (9, 44-46). Further evidence for the role of PLA₂ and COX in the cellular response to radiation injury is provided by pharmacologic inhibition or gene deletion. Treatment with NS-398, a COX-2 specific inhibitor, lowers the increased levels of PGE₂ in irradiated brain (46). In addition, the radiation-induced increase in PGE₂ in intestinal cells is attenuated by indomethacin (47), Tepoxalin (an inhibitor of COX-1, COX-2, and 5-lipoxygenase) (48), or by COX-1 gene deletion (10).

The present studies show that inhibitors of the COX pathway of arachidonic acid metabolism block the radiation-induced increase in glomerular P_{alb}. Because of the current interest in developing agents that will be effective for the mitigation and treatment of radiation injuries (5, 6), we further tested the efficacy of COX inhibitors given after irradiation (a mitigation protocol). We observed a significant protective effect of ibuprofen on radiation-induced increase

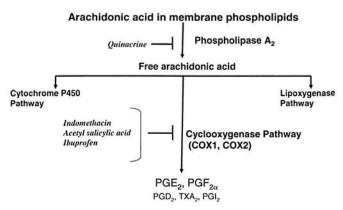


Figure 6. A simplified version of arachidonic acid metabolism. Cellular arachidonic acid is largely present in its esterified form with membrane phospholipids. Nonesterified arachidonic acid is generated by the action of phospholipase A_2 on phospholipids. The nonesterified form of arachidonic acid is metabolized *via* three main metabolic pathways involving cyclooxygenases, lipoxygenases, and cytochrome P450 enzymes. We have previously shown that cyclooxygenase products PGE₂ and PGF_{2 α} cause increased glomerular albumin permeability (14).

in P_{alb} under both conditions, although pretreatment appears to be more effective than postirradiation, indicating the extremely rapid nature of changes induced by irradiation (Fig. 5).

Indomethacin, acetylsalicylic acid, and ibuprofen are clinically used NSAIDs that inhibit both COX-1 and -2 to varying degrees. The easily available S-(+) enantiomer of (±)-ibuprofen reversibly inhibits both isoforms of COX at about equimolar concentrations (ID₅₀ 8.9–14.0 μ M and 7.2–8.2 μ M for COX-1 and COX-2, respectively) (16). Our findings strongly suggest that one or more COX metabolites of arachidonic acid participate in the early events that lead to increase in P_{alb} and may be important in the processes that lead to clinical disease weeks to months later in this model of radiation nephropathy.

Proteinuria is managed and treated using strategies that include cytotoxic agents, angiotensin-converting enzyme inhibitors (ACE), and nonsteroidal antiinflammatory drugs (NSAIDs). These regimens are aimed at retardation of the progress of proteinuria and the effects of increased protein in the glomerular filtrate. NSAIDs lower proteinuria in nephrotic syndrome resistant to corticosteroid and cytotoxic therapy (49–53). However, the mechanism by which this treatment limits glomerular filtration of proteins is not understood. In this regard, we have shown that indomethacin protects the glomerular filtration barrier from injury caused by some eicosanoids of the COX pathway (15). The present data also indicate that eicosanoids such as PGE₂ are involved in a radiation-induced increase in glomerular protein permeability.

In summary, our results suggest that metabolites of arachidonic acid contribute to the increase in glomerular permeability that is observed after a single dose of radiation. This finding points to a role for epigenetic phenomena in cellular responses to radiation. Traditionally, DNA damage

was believed to be responsible for all radiation-induced biological changes, including the immediate release of arachidonic acid (54). The importance of radiation-induced extranuclear signaling in gene expression is being increasingly acknowledged (26). We hypothesize that irradiation initiates cellular processes before the onset of genomic changes and that increased glomerular permeability is the functional manifestation of the early cellular responses to injury. Because clinical symptoms of renal injury appear weeks to months after irradiation and represent culmination of progressive changes in the entire kidney, detection of increased glomerular albumin permeability may be useful in developing strategies for early interventions. These preliminary findings also suggest that certain inhibitors of the arachidonic acid metabolism may be useful for both preventing and mitigating radiation injuries.

- Cohen EP, Fish BL, Moulder JE. The renin-angiotensin system in experimental radiation nephropathy. J Lab Clin Med 139:251-257, 2002.
- Cohen EP. Renal failure after bone marrow transplantation. Lancet 357: 6–7, 2001.
- Cohen EP, Robbins MEC. Radiation nephropathy. Semin Nephrol 23: 486–499, 2003.
- Lambert B, Cybulla M, Weiner SM, Van de Wiele C, Ham H, Dierckx, RA, Otte A. Renal toxicity after radionuclide therapy. Rad Res 161: 607-611, 2004.
- Moulder JE. Post-irradiation approaches to treatment of radiation injuries in the context of radiological terrorism and radiation accidents: a review. Int J Radiat Biol 80:3-10, 2004.
- Coleman CN, Blakely WF, Fike JR, Macvittie TJ, Metting NF, Mitchell JB, Moulder JE, Preston RJ, Seed TM, Stone HB, Tofilon PJ, Wong RSL. Molecular and cellular biology of moderate-dose (1-10 Gy) radiation and potential mechanisms of radiation protection: Report of a workshop at Bethesda, Maryland, December 17-18, 2001. Rad Res 159:812-834, 2003.
- Sharma M, Sharma R, Ge XL, Fish BL, McCarthy ET, Savin VJ, Cohen EP, Moulder JE. Early detection of radiation-induced glomerular injury by albumin permeability assay. Rad Res 155:474-480, 2001.
- Moulder JE, Fish BL. Late toxicity of total body irradiation with bone marrow transplantation in a rat model. Int J Rad Oncol Biol Phys 16: 1501–1509, 1989.
- Steinauer KK, Gibbs I, Ning S, French JN, Armstrong J, Dnox SJ. Radiation induces upregulation of cyclooxygenase-2 (COX-2) protein in PC-3 cells. Int J Rad Oncol Biol Phys 48:325-328, 2000.
- Houchen CW, Stenson WF, Cohn SM. Disruption of cyclooxygenase-1 gene results in an impaired response to radiation injury. Am J Physiol 279:G858–G865, 2000.
- 11. Foegh ML, Ramwell PW. Physiological implications of products in the arachidonic acid cascade. In: Pace-Asciak P, Granstrom E, Eds. Prostaglandins and Related Substances, Vol 5 of Neuberger A, van Deenen LLM, Series Eds. New Comprehensive Biochemistry. Amsterdam: Elsevier, pp1-34, 1983.
- Marks F. Arachidonic acid and companions: an abundant source of biological signals. In: Marks F, Furstenberger G, Eds. Prostaglandins, Leukotrienes and Other Eicosanoids: From biogenesis to clinical application. Weinheim: Wiley-VCH, pp1-40, 1999.
- Khanpure SP, Letts LG. Perspectives and clinical significance of the biochemical and molecular pharmacology of eicosanoids. In: Curtis-Prior P, Ed. Eicosanoids. New York: John Wiley & Sons, pp131-162, 2004.

- 14. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. J Biol Chem 193:265–275, 1951.
- McCarthy ET, Sharma M, Ge XL, Sharma R. Indomethacin protects glomeruli from increase in albumin permeability caused by sera from patients with focal segmental glomerulosclerosis. Kidney Int 61:534– 541, 2002.
- Meade EA, Smith WL, DeWitt DL. Differential inhibition of prostaglandin endoperoxide synthase (cyclooxygenase) isozymes by aspirin and other non-steroidal anti-inflammatory drugs. J Biol Chem 268:6610–6614, 1993.
- Williams MV. The cellular basis or renal injury by radiation. Br J Cancer 53(Suppl VII):257-264, 1986.
- Withers HR, Mason KA, Thames HDJ. Late radiation response of kidney assayed by tubule cell survival. Br J Radiol 59:587–595, 1986.
- Stewart FA. Radiation nephropathy after abdominal irradiation or totalbody irradiation. Rad Res 143:235–237, 1995.
- Cohen EP, Lawton EP. Pathogenesis, prevention, and management of radiation nephropathy. In: Tobias JS, Thomas PRM, Eds. Current Radiation Oncology, Vol 3. London: Edward Arnold Ltd, pp94-109, 1997
- Yammani RR, Sharma M, Seetharam S. Moulder JE, Dahms NM, Seetharam B. Loss of albumin and megalin binding to renal cubilin in rats results in albuminuria after total-body irradiation. Am J Physiol 283:R339–R346, 2002.
- McCarthy ET, Sharma M, Ge XL, Reddy RS, Ahmad F, Sharma R. Increased albumin permeability (P_{alb}) in glomeruli isolated from obese Zucker rats. J Am Soc Nephrol 12:842A(A4409), 2001.
- McCarthy ET, Sharma R, Sharma M. Protective effect of 20-hydroxyeicosatetraenoic acid (20-HETE) on glomerular protein permeability barrier. Kidney Int 67:152–156, 2005.
- Dahly-Vernon AJ, Sharma M, McCarthy ET, Savin VJ, Roman RJ. Transforming growth factor beta and glomerular protective actions of 20-hydroxyeicosatetraenoic acid agonist. Hypertension 45:1-6, 2005.
- Sharma M, Sharma R, McCarthy ET, Savin VJ. The focal segmental glomerulosclerosis permeability factor: Biochemical characteristics and biological effects. Exp Biol Med 229:85–98, 2004.
- Mothersill C, Seymour C. Radiation-induced bystander effects, carcinogenesis and models. Oncogene 22:7028-7033, 2003.
- Hallahan DE, Virudachalam S, Kuchibhotla J, Kufe DW. Membranederived second messenger regulates x-ray-mediated tumor necrosis factor—a gene induction. Proc Natl Acad Sci U S A 91:4897–4901, 1994
- Yatvin M, Gipp J, Dennis W. Influence of unsaturated fatty acids, membrane fluidity and oxygenation on the survival of an E. coli fatty acid auxotroph following gamma-irradiation. Int J Radiat Biol 25:539– 548, 1979.
- Samuni AM, Barenholz Y. Stable nitroxide radicals protect lipid acyl chains from radiation damage. Free Radical Biol Med 22:1165-1174, 1997.
- McPhail L, Clayton C, Synderman R. A potential second messenger role for unsaturated fatty acids: activation of Ca²⁺-dependent protein kinase. Science 224:622-625, 1984.
- Murakami K, Chan S, Routtenberg A. Protein kinase C activation by cis-fatty acid in the absence of Ca²⁺ and phospholipids. J Biol Chem 261:15424–15429, 1986.
- Peters-Golden M, McNish RW, Sporn PH, Balazovich K. Basal activation of protein kinase C in rat alveolar macrophages: implications for arachidonate metabolism. Am J Physiol 261:L462–L471, 1991.
- Donlon M, Steel L, Helgeson EA, Shipp A, Catravas GN. Radiationinduced alterations in prostaglandin excretion in the rat. Life Sci 32: 2631–2639, 1983.
- Hahn GL, Menconi MJ, Cahill M, Polgar P. The influence of gamma radiation on arachidonic acid release and prostacyclin synthesis. Prostaglandins 25:783-791, 1983.
- 35. Lognonne JL, Ducousso R, Rocquet G, Kergonou JF. Influence of

- whole-body gamma irradiation upon arachidonic acid metabolism in rat platelets, Biochimie 67:1015–1021, 1985.
- Eldor A, Vlodavsky I, Fuks Z, Matzner Y, Rubin DB. Arachidonic metabolism and radiation toxicity in cultures of vascular endothelial cells. Prostaglandins Leukot Essent Fatty Acids 36:251-258, 1989.
- Michalowski AS. On radiation damage to normal tissues and its treatment. II. Anti-inflammatory drugs. Acta Oncol 33:139–157, 1994.
- 38. Steel LK, Rafferty MA, Wolfe WW, Egan JE, Kennedy DA, Catravas GN, Jackson WE 3rd, Dooley MA. Urinary excretion of cyclic nucleotides, creatinine prostaglandin E₂ and thromboxane B₂ from mice exposed to whole-body irradiation from an enhanced neutron field. Int J Radiat Biol 50:695-715, 1986.
- Schneidkraut MJ, Kot PA, Ramwell PW, Rose JC. Regional release of cyclooxygenase products after radiation exposure of the rat. J Appl Physiol 61:1264–1269, 1986.
- Martineau LC, McVeigh LI, Jasmin BJ, Kennedy CR. p38 MAP kinase mediates mechanically induced COX-2 and PG EP4 receptor expression in podocytes: implications for the actin cytoskeleton. Am J Physiol 286:F693-F701, 2004.
- Breyer MD. Prostaglandin receptors in the kidney: a new route for intervention? Exp Neph 6:180–188, 1998.
- Bresnahan BA, Kelefiotis D, Stratidakis I, Lianos EA. PGF_{2α}-induced signaling events in glomerular mesangial cells. Proc Soc Exp Biol Med 212:165–173, 1996.
- Lianos EA. Biosynthesis and role of arachidonic acid metabolites in glomerulonephritis, Nephron. 37:73-77, 1984.
- 44. Chen X, Gresham A, Morrison A, Pentland AP. Oxidative stress mediates synthesis of cytosolic phospholipase A₂ after UVB injury. Biochim Biophys Acta 1299:23–33, 1996.
- 45. Isoherranen K, Punnonen K, Jansen C, Uotila P. Ultraviolet irradiation

- induces cyclooxygenase-2 expression in keratinocytes. Br J Dermatol 140:1017–1022, 1999.
- 46. Kyrkanides S, Moore AH, Olschowka JA, Daeschner JC, Williams JP, Hansen JT, Kerry O'Banion M. Cyclooxygenase-2 modulates brain inflammation-related gene expression in central nervous system radiation injury. Brain Res (Mol Brain Res) 104:159–169, 2002.
- Cohn SM, Schloemann S, Tessner T, Seibert K, Stenson WF. Crypt stem cell survival in the mouse intestinal epithelium is regulated by prostaglandins synthesized through cyclooxygenase-1. J Clin Invest 99: 1367-1379, 1997.
- Panes J, Molla M, Casadevall M, Salas A, Sans M, Conill C, Anderson DC, Rosello-Catafau J, Granger DN, Pique JM. Tepoxalin inhibits inflammation and microvascular dysfunction induced by abdominal irradiation in rats. Aliment Pharmacol Ther 14:841-850, 2000.
- Donker AJM, Brentjens JRH, van der Hem GK, Arisz L. Treatment of the nephrotic syndrome with indomethacin. Nephron 22:373-381, 1978
- Glassock RJ, Focus on proteinuria. Am J Nephrol 10(Suppl 1):88–93, 1990.
- Bergstein JM. Prostaglandin inhibitors in the treatment of nephrotic syndrome. Pediatr Nephrol 5:335–338, 1991.
- Hutchison FN. Hormonal modulation of proteinuria in the nephrotic syndrome. Am J Nephrol 13:337–346, 1993.
- Remuzzi A, Remuzzi G. The effects of nonsteroidal anti-inflammatory drugs on glomerular filtration of proteins and their therapeutic utility. Semin Nephrol 15:236-243, 1995.
- Kaleta EW, Applegate LA, Ley RD. Photoreactivation of ultraviolet radiation-induced release of arachidonic acid from marsupial cells. Photochem Photobiol 54:747-752, 1991.