

MINIREVIEW

Treatment of Viral and Neoplastic Diseases with Double-Stranded RNA Derivatives and Other New Agents

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Many attempts have been made to inhibit viral and neoplastic diseases by targeting the RNA system. The pathophysiologic significance of the microRNA system and the therapeutic potential of its manipulation are discussed. Studies of double-stranded RNA derivatives are reviewed. The therapeutic potential of one of these compounds, polyI:MPC, is emphasized. Studies of other related antiviral and antineoplastic agents are discussed, including 2'-deoxyoligocytidilates and telomerase inhibitors. *Exp Biol Med* 231:1283–1286, 2006

Key words: AIDS; double-stranded and single-stranded RNA; HIV infection; neoplastic diseases; thiolated polyI:polyC (pl:MPC); 2'-deoxyoligocytidilates; telomerase inhibitors; viral diseases

Introduction

With the rapidly developing drug resistance of some viruses and the difficulty in the timing of vaccine production and activity, there is great concern about viral disorders. Of particular concern are newly emerging viruses, hybrid viruses, and viruses bioengineered for use in bioterrorism.

Similar problems exist with certain types of neoplastic disease. For this reason, there is increasing interest in therapeutic manipulation of the RNA system. We will briefly summarize the related studies.

RNA Interference

Gene-silencing techniques (RNA interference) are of particular interest in cancer research. In a series of recent reports (1–5), the roles of interfering RNAs (microRNAs) in cancer development were discussed. It is anticipated that these types of studies may lead to new diagnostic and therapeutic approaches. MicroRNAs are 21- to 25-nucleotide-long regulatory molecules that affect normal growth and development in plants and animals (6). They inhibit the translation of selected mRNAs into proteins. We know little about the normal function of individual microRNAs or about the development and function of abnormal microRNAs in neoplastic cells (3, 7). Johnson *et al.* (8) reported that the let-7 family of microRNAs regulate the expression of the ras oncogene. Michael *et al.* (9) described two microRNAs that are present in reduced quantities in precancerous and cancerous colorectal tissue. O'Donnell *et al.* (1) found that some abnormal microRNAs are regulated by a protein encoded by the well-known c-myc oncogene. Preliminary research indicates that microRNAs controlled by c-myc may also inhibit the expression of the E2F gene family, known inducers of apoptosis that encode another protein involved in cellular proliferation (10). The two genes mutually promote a vicious regulatory cycle leading to increased proliferation of neoplastic cells. Years of research may be required before adequate therapy can be developed based on these initial findings. Research along

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these lines may be of importance in the therapy of certain viral disorders, as well as neoplastic diseases.

Studies of Double-Stranded RNA Derivatives

Double-stranded RNA derivatives have experimentally been found to have antiviral and antineoplastic effects, thought to be due in part to their capacity to induce interferons (IFNs). Haines *et al.* (11), Torrence and DeClereq (12), DeClereq (13), and Sen *et al.* (14) reported that double-stranded RNA has the capacity to induce IFNs in animal and human cells and to increase resistance to viral infections. A prototype of double-stranded RNA, polyI:polyC, has been reported to inhibit the growth of some tumor cell lines and the growth rate of human tumor xenografts in mice (15, 16). While polyI:polyC is the most potent among the double-stranded RNAs tested, it has proved to be too toxic for therapeutic application (17, 18). Selective thiolation at the 5 position of the cytosine bases in polyI:polyC resulted in partially thiolated double-stranded RNA, polyI:MPC (Fig. 1, Refs. 19, 20). PolyI:MPC was found to be an important inducer of IFN- α , IFN- β , and IFN- γ (20–29). It is of minimal toxicity *in vivo* in mice, rabbits, and guinea pigs and is subject to less degradation by plasma ribonucleases than its parent compound (22). PolyI:MPC has significant antitemplate activity against DNA and RNA polymerases, including reverse transcriptase (which is of great significance in the treatment of human immunodeficiency virus [HIV] infection) (23–27). This compound activates IFN-induced double-stranded RNA-dependent protein kinase (which inhibits protein synthesis) and activates 2',5'-oligoadenylate synthetase, which in turn activates a latent endoribonuclease and activates adenylate cyclase, thus increasing the concentration of cAMP. PolyI:MPC activates macrophages and augments the natural killer cell activity (20, 27). In primary human lymphocyte cultures, polyI:MPC was found to be a potent inhibitor of HIV replication *in vitro*, including multidrug-resistant HIV

lines in preliminary results (23). A recent study indicated that 7.5% thiolation is optimal for this compound from the point of view of IFN- α , IFN- β , and IFN- γ production and antiviral and antiproliferative activity (25). The effect of polyI:MPC is being explored alone and in combination with other anti-HIV agents, including those reported in the retroviral MAIDS model in mice (30). This model was found to be a useful, inexpensive, and quick early screening approach to acquired immunodeficiency syndrome (AIDS) and AIDS-related lymphoma (31). Investigations will proceed to nonhuman primate models and eventually to clinical studies. We anticipate that these agents and derivatives may contribute to the treatment of viral and neoplastic diseases, overcome disease-related IFN resistance (32), and stimulate other factors of the immune system. Figure 1 shows the chemical structure and partial 5-thiolation of cytosine residues of polynucleotides.

2'-Deoxyoligocytidilates

Investigators have produced and evaluated a series of polyI:MPC compounds that are closely and distantly related, as well as others in viral and neoplastic pathology. The most effective included oligonucleotides converted to 4 thio-deoxyuridylic acid (s^4d UMP)₃₅ (Fig. 2) containing oligomers of the 4 amino group of the cytosine bases, which were converted to the corresponding 4-thio group in 2'-deoxyoligocytidilates. 2'-Deoxyoligocytidilates inhibited a number of established tumor cell lines *in vitro* and was not cytotoxic to human granulocyte-macrophage progenitor cells. It was effective *in vitro* against multidrug-resistant HIV lines (23). 2'-Deoxyoligocytidilates also appeared to inhibit attachment of HIV to target cell receptors and viral fusion to cell membranes. In a series of publications, the chemistry and the antiviral effects (29, 33, 34) of this agent

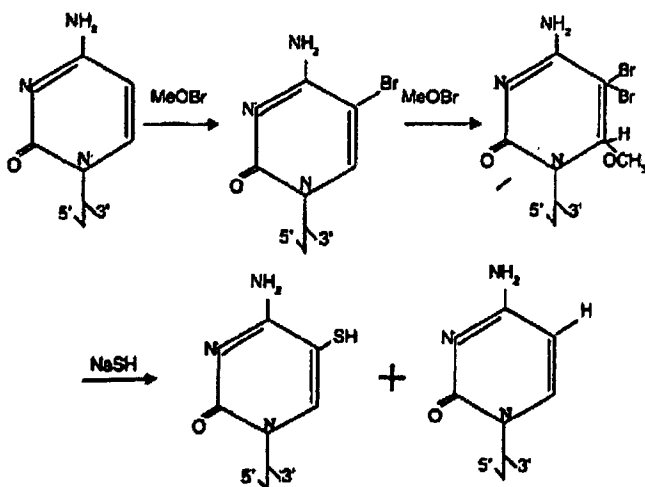


Figure 1. Chemical structure and partial 5-thiolation of cytosine residues of polynucleotides. MeOBr, methyl hypobromide; NaSH, sodium sulphhydride.

35-meric oligo (4-thio-2'-deoxyuridylate)

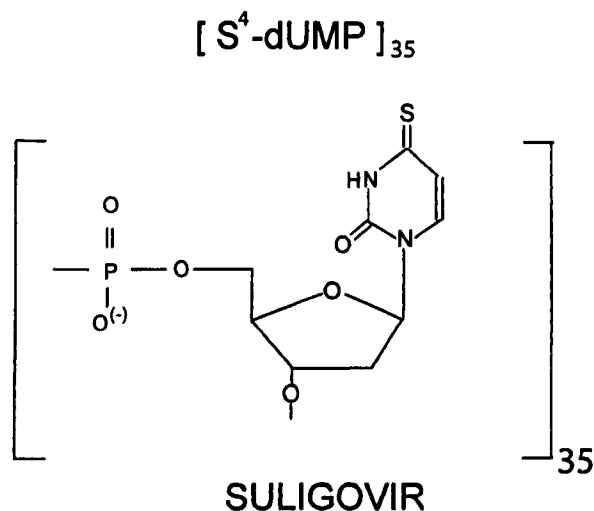


Figure 2. Chemical structure of (s^4d UMP)₃₅ (Suligovir).

were discussed. Further *in vivo* studies of the possible role of 2'-deoxyoligocytidilates in oncology are under way.

Telomerase Inhibitors

Other synthesized compounds are telomerase inhibitors (35, 36). Telomerase, a ribonucleoprotein complex, includes an RNA template and a catalytic subunit with reverse transcriptase activity and is responsible for the maintenance of normal telomere structure (37). In contrast to healthy adult somatic cells (38–40), most human neoplastic cells exhibit high telomerase activity, with some exceptions, including stem cells of renewable tissues and activated lymphocytes (41). Telomerases thus protect cells against apoptosis (40). Telomerase inhibitors have been described previously (41). Matthes and Lehmann (42) were the first to propose the evaluation of antitelomerase chimeric oligonucleotides, which contain a moiety targeting the primer binding sites and a sequence targeting the RNA as well. One compound that is fairly effective in inhibiting telomerases is a chimeric oligonucleotide composed of an antisense sequence directed against RNA template regions and a base modified moiety. The chemical synthesis has been described (29, 33, 35). Evaluation in several biological model studies is under way, including studies of various therapeutic combinations. In a recent study, retinoid and arsenic combination therapy was effective in patients with active promyelocytic leukemia, including those who were resistant to retinoic acid alone (43). This appears to be due in part to partial telomerase inhibition. Future studies will investigate specific telomerase inhibitors relative to their effect in combination therapy of promyelocytic leukemia and related diseases.

Summary

This brief review demonstrates the potent antiviral activity (such as agents to treat multidrug-resistant HIV) and antineoplastic effects of new therapies that are based on double-stranded RNA derivatives (including polyI:polyC), deoxyoligocytidilate compounds, and telomerase inhibitors (such as those containing methylated deoxyuridine moiety). Further *in vitro* studies are planned to investigate the effects of these agents in animal and human models.

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1. O'Donnell KA, Wentzel EA, Zeller KI, Dans CV, Mendell JT. c-Myc-regulated microRNAs modulate E2F1 expression. *Nature* 435:839–843, 2005.
2. Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Grider A, Ebert BL, Mak RH, Ferrando AA, Downing JR, Jacks T, Horvitz HR, Golub TR. MicroRNA expression profiles classify human cancers. *Nature* 435:834–838, 2005.
3. He L, Thompson JM, Hemann MT, Hermendo-Mange E, Mu D,

- Goodson S, Powers S. A microRNA polycistron as potential human oncogene. *Nature* 435:828–833, 2005.
4. Meltzer PS. Cancer genomics: small RNAs with big impacts [news]. *Nature* 435:745–746, 2005.
5. Hampton T. MicroRNAs move into cancer research [news]. *JAMA* 294:411–412, 2005.
6. Bartel DP. MicroRNAs: genetics, biogenesis, mechanism, and function. *Cell* 116:281–297, 2004.
7. Ota A, Tagawa H, Kaman S, Tsuzuki S, Karpas A, Kira S, Yoshida Y, Seto M. Identification and characterization of a novel gene, *C13orf25*, as a target for 13q31-q32 amplification in malignant lymphoma. *Cancer Res* 64:3087–3095, 2004.
8. Johnson SM, Grosshans H, Shingara J, Byrom M, Jarvis R, Cheng A, Labouvier E, Reinert KL, Brown D, Slack FJ. RAS is regulated by the let-7 microRNA family. *Cell* 120:635–647, 2005.
9. Michael MZ, O'Connor SM, van Holst Pellekaan NG, Young GP, James RJ. Reduced accumulation of specific microRNAs in colorectal neoplasia. *Mol Cancer Res* 1:882–891, 2003.
10. Ginsberg D. E2F1 pathways to apoptosis. *FEBS Lett* 529:122–125, 2002.
11. Haines DS, Strauss KI, Gillespie DH. Cellular response to double-stranded RNA. *J Cell Biochem* 46:9–20, 1991.
12. Torrence PF, DeClereq E. Interferon inducers: general surgery and classification. *Methods Enzymol* 78:291–299, 1981.
13. DeClereq E. Synthetic interferon inducers. *Top Curr Chem* 52:173–203, 1974.
14. Sen GC, Taira H, Lengyel P. Interferon, double-stranded RNA, and protein phosphorylation: characteristics of a double-stranded RNA-activated protein kinase system partially purified from interferon treated Ehrlich ascites tumor cells. *J Biol Chem* 253:5915–5921, 1978.
15. Levy HB. Interferon and interferon inducers, I: the treatment of malignancies. *Arch Intern Med* 126:78–83, 1970.
16. Levy HB, Law LW, Rabson AS. Inhibition of tumor growth by polyinosinic-polycytidylic acid. *Proc Natl Acad Sci U S A* 62:357–361, 1969.
17. Bucur N, Mizuno M, Wakabayashi T, Yoshida J. Growth inhibition of experimental glioma by human interferon- β superinduced by cationic liposomes entrapping polyinosilic:polycytidylic acid. *Neurol Med Chir (Tokyo)* 38:469–474, 1998.
18. Stebbing N, Grantham, CA. Anti-viral activity against encephalomyocarditis virus and Semliki Forest virus and acute toxicity of poly I and poly C administered sequentially to mice. *Arch Virol* 51:199–215, 1976.
19. Bardos TJ, Aradi J, Ho YK, Halman TI. Biochemical properties of 5-sulfur-substituted pyrimidine nucleosides and nucleotides. *Ann N Y Acad Sci* 255:522–531, 1975.
20. Bardos TJ, Novak P, Chakrabati P, Ho YK. Partially thiolated polycytidylic acid. In: Townsend LB, Tipson RS, Eds. *Nucleic Acid Chemistry*. New York: J Wiley & Sons Inc, pp881–884, 1978.
21. O'Malley JA, Hu YK, Chakrabarti P, DiBerardino L, Chandra P, Orinda DA, Byrd DM, Bardos TJ, Carter WA. Antiviral activity of partially thiolated polynucleotides. *Mol Pharmacol* 11:61–69, 1975.
22. Vastola KA, Ho YK, Bardos TJ, Grossmayer BJ, Fruck-Diviak L, O'Malley JA. Poly I-mercapto poly C: antiviral, anticellular, and pharmacologic effects. *Res Commun Chem Pathol Pharmacol* 45:407–419, 1984.
23. Bardos TJ, Schinazi, RF, Ling KH, Helder AR. Structure-activity relationships and mode of action of 5-mercapto-substituted oligo- and polynucleotides as antitemplates inhibiting replication of human immunodeficiency virus type 1. *Antimicrob Agents Chemother* 36:108–114, 1992.
24. Cavanaugh PF Jr, Ho YK, Hughes RG Jr, Bardos TJ. Selectivity of antitemplates as inhibitors of deoxyribonucleic acid polymerases. *Biochem Pharmacol* 31:4055–4060, 1982.

25. Chadha KC, Dembinski WE, Dunn CB, Aradi J, Bardos TJ, Dunn JA, Ambrus JL Sr. Effect of increasing thiolation of the polycytidylic acid strand of poly I:poly C on the α , β and γ interferon-inducing properties, antiviral and antiproliferative activities. *Antiviral Res* 64:171–177, 2004.
26. Kung MP, Ho YK, Bardos TJ. Action of partially thiolated polynucleotides on the DNA polymerase alpha from regenerating rat liver. *Cancer Res* 36:4537–4542, 1976.
27. Heider AR, Bardos TJ. Oligonucleotides and polynucleotides as potential cancer chemotherapeutic agents. In: Foye WO, Ed. *Cancer Chemotherapeutic Agents*. Washington: American Chemical Society, pp526–575, 1996.
28. Milkulski AJ, Bardos TJ, Chakrabarti P, Kalman TI, Zsindely A. Inhibition of DNA-dependent RNA polymerase with partially thiolated polynucleotides. *Biochem Biophys Acta* 319:294–303, 1973.
29. Horvath A, Aradi J. Advantages of sodium perchlorate solution as mobile phase for purification of synthetic oligonucleotides by anion exchange chromatography. *Anal Biochem* 338:341–343, 2005.
30. Chadha KC, Stadler I, Ambrus JL, Nair MPN. Effect of alcohol and MAIDS virus infection upon immunological status in C57BL/6 mice. *Recent Res Dev Immunol* 2:41–51, 2000.
31. Stadler I, Chadha KC, Nakeeb S, Toumbis C, Butsch J, Mathur N, Munschauer F, Vladutiu A, Satchidanand SK, Ambrus JL. Pentoxifylline and meclofenamic acid treatment reduces clinical manifestations in a murine model of AIDS. *J Pharmacol Exp Ther* 268:10–13, 1994.
32. Chadha KC, Ambrus JL Jr, Dembinski W, Ambrus JL Sr. Interferons and interferon inhibitory activity in disease and therapy. *Exp Biol Med* 229:285–290, 2004.
33. Tokes S, Aradi J. (s^4dU)₃₅: A novel, highly potent oligonucleotide inhibitor of the human immunodeficiency virus type 1 reverse transcriptase. *FEBS Lett* 396:43–46, 1996.
34. Horvath A, Tokes S, Hartman T, Watson K, Turpin JA, Buckheit RW Jr, Sebestyen Z, Szollosi J, Benko I, Bardos TJ, Dunn JA, Fesus L, Toth FD, Aradi J. Potent inhibition of HIV-1 entry by (s^4dU)₃₅. *Virology* 334:214–223, 2005.
35. Szatmari I, Tokes S, Dunn C, Bardos TJ, Aradi J. Modified telomeric repeat amplification protocol: a quantitative radioactive assay for telomerase without using electrophoresis. *Anal Biochem* 282:80–88, 2000.
36. Tarkanyi I, Horvath A, Szatmari I, Eizert H, Vamosi G, Damjanovich S, Segal-Bendirdjian E, Aradi J. Inhibition of human telomerase by oligonucleotide chimeras, composed of an antisense moiety and a chemically modified homo-oligonucleotide. *FEBS Lett* 579:1411–1416, 2005.
37. Morin GB. The human telomere terminal transferase enzyme is a ribonucleoprotein that synthesizes TTAGGG repeats. *Cell* 59:521–529, 1989.
38. Kelleher C, Teixeira MT, Forstemann K, Lingner J. Telomerase: biochemical considerations for enzyme and substrate. *Trends Biochem Sci* 27:572–579, 2002.
39. Kim NW, Piatyszek MA, Prowse KR, Harley CB, West MD, Ho PLC, Coviello GM, Wright WE, Weinrich SL, Shay JW. Specific association of human telomerase activity with immortal cells and cancer. *Science* 266:2011–2015, 1994.
40. Stewart SA, Hahn WC, O'Connor BF, Banner EN, Lundberg AS, Modha P, Mizuno H, Brooks MW, Fleming M, Zimonjic DB, Popescu NC, Weinberg RA. Telomerase contributes to tumorigenesis by a telomere length-independent mechanism. *Proc Natl Acad Sci U S A* 99:12606–12611, 2002.
41. Mergny JL, Riou JF, Mailliet P, Teulade-Fichou MP, Gilson E. Natural and pharmacological regulation of telomerase. *Nucleic Acids Res* 30:839–865, 2002.
42. Matthes E, Lehmann C. Telomerase protein rather than its RNA is the target of phosphorothioate-modified oligonucleotides. *Nucleic Acids Res* 27:1152–1158, 1999.
43. Tarkanyi I, Dudognon C, Hillion J, Pendino F, Lanotte M, Aradi J, Segal-Bendirdjian E. Retinoid/arsenic combination therapy of promyelocytic leukemia: induction of telomerase-dependent cell death. *Leukemia* 19:1806–1811, 2005.