

Effects of magnesium deficiency – More than skin deep

Navin Chandrakanth Chandrasekaran^{1,2}, Christopher Weir³, Sumaya Alfraji¹, Jeff Grice², Michael S Roberts² and Ross T Barnard¹

¹School of Chemistry and Molecular Biosciences, Australian Infectious Diseases Research Centre, The University of Queensland, Queensland 4072, Australia; ²School of Medicine, Translational Research Institute, The University of Queensland, Woolloongabba, Queensland 4102, Australia; ³Walter and Eliza Hall Institute of Medical Research and Department of Medical Biology, University of Melbourne, Parkville, Victoria 3052, Australia
Corresponding author: Ross T Barnard. Email: rossbarnard@uq.edu.au

Abstract

Dead Sea and magnesium salt therapy are two of the oldest forms of treatment for skin disease and several other disorders, supported by a body of largely anecdotal evidence. In this paper we review possible pathways for penetration of magnesium ions through the epidermis to reach the circulation, in turn replenishing cellular magnesium levels. We also discuss mechanisms for intercellular movement of magnesium ions and possible mechanisms for the interaction between magnesium ions and inflammatory mediators. Upon addition of magnesium ions *in vitro*, the expression of inflammatory mediators such as tumour necrosis factor α (TNF α) and nuclear factor κ B (NF κ B) is down regulated. Dysregulation of these and other inflammatory mediators has been linked to several inflammatory disorders, including asthma, arthritis, atherosclerosis and neuroinflammation.

Keywords: Magnesium, inflammation, skin

Experimental Biology and Medicine 2014; 239: 1280–1291. DOI: 10.1177/1535370214537745

Introduction

Dead Sea therapy is one of the oldest forms of treatment for skin disease and some chronic inflammatory diseases like arthritis and psoriasis.¹ Much of the research to date has attributed the clinical effects of Dead Sea therapy to its mineral composition; mostly to magnesium salts.^{2,3} Magnesium salts, such as magnesium sulphate (Epsom salts), have long been used as a spa product and as a therapeutic to manage clinical conditions.⁴

The central question addressed by this review is, “What are the underlying mechanisms by which magnesium ions could play a role in the regulation of inflammatory responses in the skin and systemically?” Several systematic studies have been conducted in humans, over the last two decades, in an effort to understand the effect of magnesium ions (Mg²⁺) in healing skin disorders. These will be reviewed. There has been renewed interest over the past decade in understanding the role of magnesium salts in clinical medicine, nutrition and physiology. This review will discuss the known clinical effects of magnesium deficiency, and both summarise and suggest molecular mechanisms that could mediate the inflammation induced by magnesium deficiency. We first discuss transdermal absorption as a possible route of administration for prevention and

treatment of magnesium deficiency and for controlling inflammation.

Transdermal absorption of magnesium

Transport of Mg²⁺ across skin is a critical precondition for the function of topical, therapeutic compounds in treating skin and inflammatory diseases. Transdermal absorption is a potentially important route of transport for components that are involved in biological processes.⁵ Even though much research has been carried out in the area of cutaneous permeation and transdermal absorption,^{5–7} mechanisms that lead to permeation of Mg²⁺ ions through the skin are not clearly understood and need further research.

Past studies on magnesium and other metal ion permeation through human skin demonstrated that it is not readily absorbed under normal physiological conditions, when the skin is intact and healthy.^{8–10} However, there is a considerable body of anecdotal and research data that attributes to magnesium a role in skin barrier and epidermal recovery after damage.^{3,11–13} In the case of compromised stratum corneum (SC), the viable epidermis and nerve endings (in atopic dermatitis [AD]) are exposed to incoming particles and chemicals.^{14,15} There is no effective barrier to restrict the movement of magnesium ions to epidermal cells or nerve

endings, thus permitting a role for Mg²⁺ in skin recovery and modulation of the immune or nervous systems.^{3,16} The permeability of the skin is modified in pathological conditions, with both macroscopic and microscopic lesions^{17,18} that would allow penetration of magnesium below the SC, and subsequent transport or diffusion by mechanisms to be discussed below. Thus it is necessary to consider both the normal skin as well as barrier compromised or diseased skin.

In normal skin, the SC forms the outermost layer, formed by continuous replacement from the newly differentiated daughter cells of keratinocyte stem cells, displacing outwards.¹⁹ It functions as a physical barrier hindering, but not completely preventing, transdermal penetration through its cellular structure.⁵ The radius of the hydrated magnesium ion relative to the radius of the dehydrated ion is greater compared to other ions such as calcium, potassium and sodium.^{10,20} Irrespective of oral or transdermal administration, this greater radius could sterically and energetically hinder transport across cellular membranes. A review article by Lansdown⁸ reports that magnesium in the form of hydrous polysilicate (talc) is not readily absorbed by normal skin, however, commonly used therapeutic formulations of magnesium utilize other salts such as chloride, or sulphates. The absorption kinetics such as solubility and permeation coefficients of chlorides and sulphates are different to those of polysilicates.^{4,8,21,22} Moreover, Table 7 in the same review article presents a positive score for percutaneous absorption of magnesium ions (although the temperature conditions are not specified). The same article cites the ability of Mg²⁺ to bind to hair.²³ This provides the possibility of magnesium permeation by shunt diffusion. Shunt diffusion is the mechanism by which diffusion occurs through hair follicles, pilosebaceous units and sweat glands,⁵ although these constitute a small proportion of skin surface area, with the density dependent on the location of the skin. In the case of bulk diffusion, water soluble molecules are able to enter through 10 Å pores created by protein subunits in the lipid of SC.^{5,6,24} These could provide an entrance for hydrated magnesium, the radius of which is 4.76 Å.^{25,26} Subsequently, the transport of Mg²⁺ into cells could be facilitated by transmembrane proteins such as SLC41A1 and transient receptor potential melastatin 7 (TRPM7).^{27,28} SLC41A2, a cell surface transmembrane protein with its N-terminus outside and C-terminus inside the cell membrane, is responsible for magnesium transport across the plasma membrane.²⁷ Immunohistochemistry on epidermal cells has demonstrated a plasma membrane localization of murine SLC41A2.²⁷ The N-terminus of this protein, accessible to extracellular components, is involved in transcellular movement of Mg²⁺, which is in turn required for homeostasis, cell growth and neuronal function.²⁷ Similarly, the human SLC41A1 functions as a Mg²⁺ transporter involved in magnesium homeostasis in epithelial cells.²⁷

Another important mechanism by which Mg²⁺ intracellular homeostasis in humans is facilitated, is *via* the protein TRPM7.²⁸ Knockout of TRPM7 in DT40 B cells (derived from an avian leucosis virus induced bursal lymphoma in

a white leghorn chicken) resulted in lowered intracellular Mg²⁺ and inhibition of cellular proliferation.²⁹ Under stress (apoptotic stimuli), the TRPM7-knockdown fibroblast cell line (3T3-M7shRNA6) was more resistant to apoptosis and had a lower intracellular concentration of reactive oxygen species (ROS) compared to control cells. This suggests a role for Mg²⁺, mediated by the magnesium transporter TRPM7, in cell survival and regulation of cellular ROS concentration.²⁹

Another factor influencing percutaneous absorption of magnesium ions through skin, is the negative charge carried on the surface of tissues.³⁰ Accordingly, it is likely that the positively charged magnesium ions can be absorbed on the negatively charged SC, enhancing the retention time and bioavailability on the skin surface.³¹ This coupled with bulk diffusion and the factors mentioned above could enhance magnesium ion penetration through normal human skin.

In normal human skin, factors such as temperature and humidity, osmolarity, dehydration and penetration enhancers, could lead to enhanced percutaneous absorption of magnesium. In certain therapies, increased temperature conditions can also provide improved skin permeability enabling penetration of mineral salts.^{5,32,33} A study conducted to measure the effect of heat on skin permeability showed a strong dependence of permeability on temperature. Short pulses of high temperature resulted in increases in calcein permeability in human cadaver skin.³⁴ In relation to Dead Sea therapy on normal human skin, the high salt concentration coupled with the hydrated state of the skin could together cause an osmotic effect,³⁵ leading to increased flux of ions through the skin due to a concentration gradient across the skin. However, in commercially available topical magnesium formulations it is likely that penetration enhancers would be necessary in order to enhance passage through the SC layer in normal skin. The role of these enhancers is to penetrate into the skin, reversibly decrease the barrier resistance of the SC and to create a water equilibrium between SC and viable epidermis.³⁶

Control of inflammation by magnesium: Possible mechanisms

It is well established that Mg²⁺ deficiency has a direct influence on inflammation.³⁷ However, the molecular mechanisms by which Mg²⁺ suppresses inflammation are unclear. A possible link could be activation by Mg²⁺ of the thiamine pyrophosphate (TPP)-dependent riboswitch, resulting in increased synthesis of thiazole from thiazole pyrophosphate³⁸ (see Figure. 1). The TPP-dependant riboswitch is the only known functional riboswitch mechanism in eukaryotes and it is known that TPP binding to *thiA* riboswitch is increased by Mg²⁺.^{38,39} The adenine rich sites on the riboswitch bind Mg²⁺ leading to structural changes favouring TPP binding.³⁸ The TPP-dependant *thiA* riboswitch up-regulates thiazole synthase, an enzyme required for catalysing the conversion of thiazole pyrophosphate to

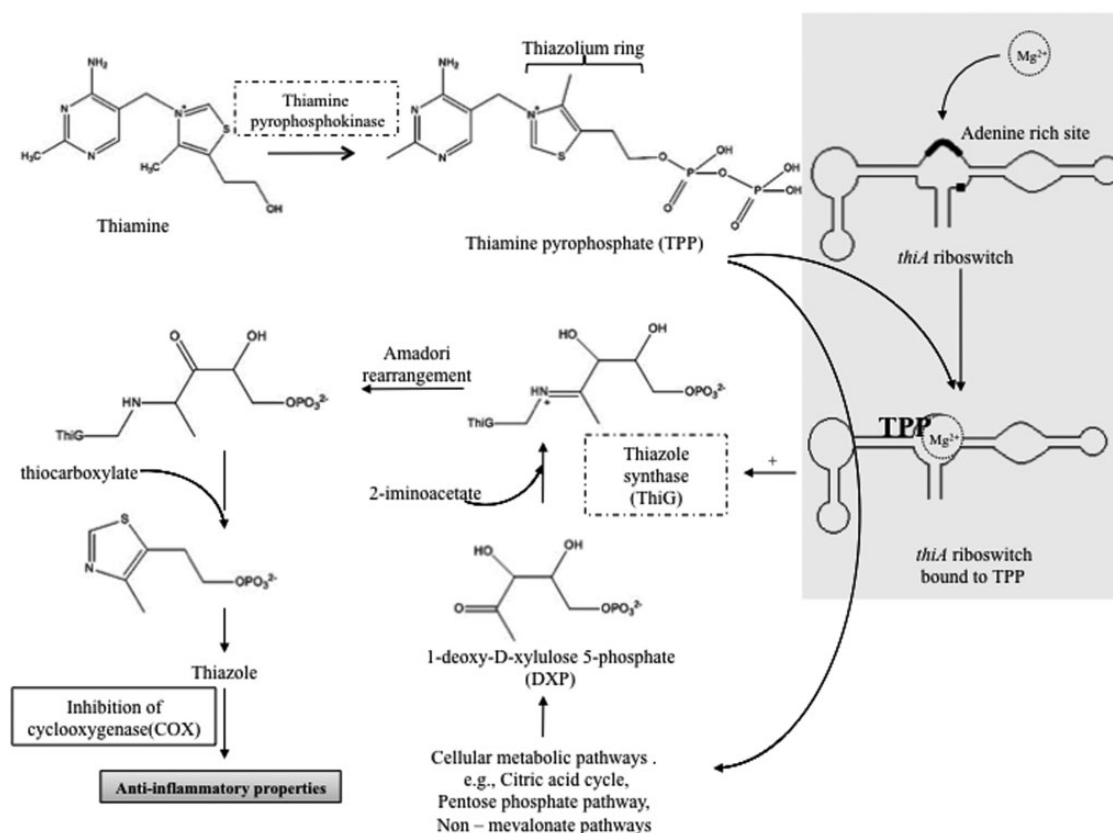


Figure 1 Possible influence of Mg^{2+} in thiazole synthesis in turn resulting in anti-inflammatory properties. Thiamine pyrophosphate (TPP), an activated form of thiamine binds to *thiA* riboswitch in the presence of magnesium ions (Mg^{2+}). An adenine rich site in the riboswitch binds Mg^{2+} increasing the affinity of TPP binding.¹³ The riboswitch undergoes structural change then expresses thiazole synthase,¹³ resulting in the formation of TPP intermediate and in turn forming thiazole derivatives. Thiazole derivatives are known to inhibit cyclooxygenase, hindering the formation of prostanoids, in turn producing anti-inflammatory effects.^{17–19} The *thiA* riboswitch is the only known eukaryotic riboswitch.¹⁴

thiazole.^{40,41} Thiazole derivatives have a spectrum of anti-inflammatory and neuroprotective activities.^{42–46}

Molecular mechanisms and inflammatory pathways

Illnesses related to inflammation following a state of chronic or acute hypomagnesaemia are well documented, both in humans and experimental rat models.^{37,47–49} However, there is a paucity of information in the literature regarding the cascade of molecular events culminating in inflammation during hypomagnesaemia.

Here we review known molecular mechanisms and propose additional ones that could be responsible for inflammation resulting from Mg^{2+} deficiency, additionally suggesting how this might relate to inflammation localized in the skin. Firstly an examination of how the hypomagnesaemic state results in tumour necrosis factor α (TNF α) and nuclear factor κB (NF κB) activation will be considered, in view of the importance as inducers of transcription of pro-inflammatory genes. These changes pave the way for pro-inflammatory cytokine responses, followed by alteration of macrophage and neutrophil activity, including their participation in a pro-inflammatory positive feedback loop (see Figure 2).

Magnesium in TNF α and NF κB signalling

It has been demonstrated that hypomagnesaemia in rodents³⁷ and in people with metabolic syndrome⁴⁷ result in elevated serum concentrations of TNF α compared to healthy counterparts (in the human studies), or controls (in rodent studies). An elevation in TNF α and magnesium deficiency was also observed in obese human subjects when compared to healthy and moderately overweight individuals.⁵⁰ A recent study revealed that TNF α levels declined following *in-vivo* administration of $MgSO_4$ to human subjects.⁵¹ Additional work showed that the magnesium ion component, not the sulphate, was responsible for the immunomodulatory effect.⁵¹ Other research has shown that magnesium deficiency in mice with a knockout of the gene encoding the TNF α receptor (TNF α R) caused less adverse effects on bone loss than in the wild-type controls fed the same diet.⁴⁸ These results suggest an inverse relationship between magnesium intake, TNF α concentration and TNF actions mediated by the TNF α R. The physiological significance of these observations becomes evident in the context of the known pro-inflammatory actions of TNF α .⁵² TNF α is a regulatory cytokine produced by various cell types including macrophages, T-helper cells (CD4+ T_H cells) and natural killer (NK) cells.⁵³ The predominant role

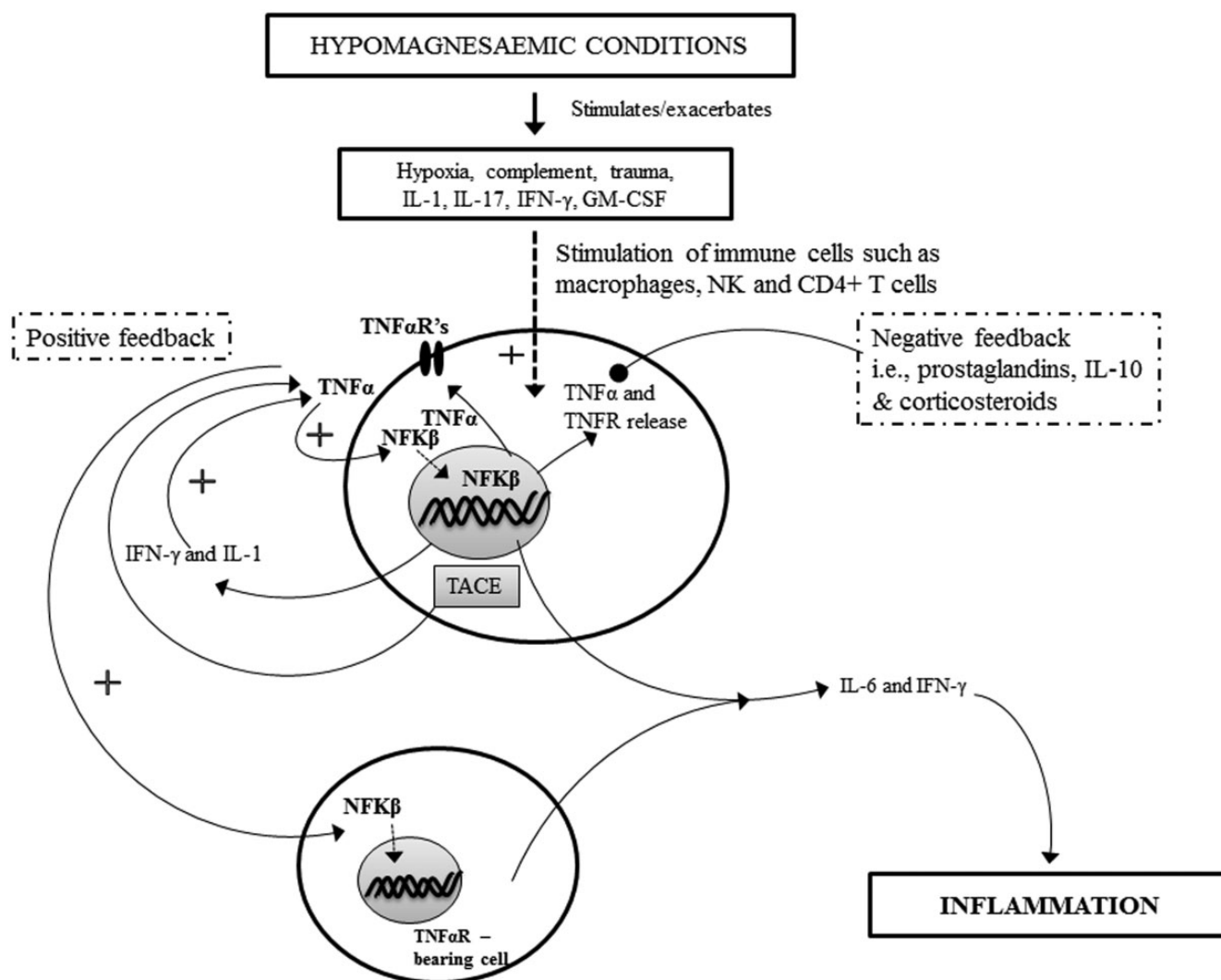


Figure 2 General summarized pathway of inflammation following hypomagnesaemia. Evidence suggests that hypomagnesaemic conditions can exacerbate trauma and hypoxia (leading to increased oxidative damage to cells and tissues from free radicals,^{112,113} and stimulates the production of interleukin 1 (IL-1), IL-17, interferon γ (IFN- γ) and granulocyte macrophage colony-stimulating factor (GM-CSF). The presence of these soluble proteins allows for the stimulation of various immune cells to produce tumour necrosis factor α (TNF α) and its reciprocal receptor (for cell surface presentation) following the nuclear translocation of nuclear factor κ B (NF κ B) acting as a transcription factor. The secretion of TNF α from cells (after TNF α converting enzyme (TACE) converts tm TNF α to its soluble form) allows for autocrine and paracrine effects that upon TNF α binding facilitates amplification of inflammatory responses such as transcription, translation and secretion of more TNF α , IL-1, IL6 and IFN- γ . A positive feedback loop is established and the latter two cytokines cause inflammation. Normally prostaglandins, IL-10 and corticosteroids have an inhibitory effect on TNF α transcription; however this is promoted by Mg²⁺ and may therefore be disrupted in hypomagnesaemic conditions.

of this cytokine is the systemic regulation of immune cells with beneficial outcomes such as augmented recruitment of defence mechanisms during infection, including fever induction.⁵² However, in common with all endogenous immune mediators, balance is essential and a prolonged high serum concentration of TNF α results in prolonged inflammation and effective damage locally and systemically.⁵² Systemic diseases such as systemic lupus erythematosus and local organ diseases including psoriasis and osteoarthritis are all associated with dysregulation (increased concentrations) of this cytokine.^{49,54,55}

The biosynthesis of TNF α is increased by an array of stimuli including hypoxia, trauma, complement components⁵⁶ and various cytokines including interleukin 1 (IL-1), IL-17 interferon- γ (IFN- γ) and granulocyte macrophage

colony-stimulating factor (GM-CSF).⁵² It has been shown *in vitro*, using various cell culture models (including human and rat cell lines) that under hypomagnesaemic conditions in the culture medium, the concentrations of the aforementioned cytokines show an increase compared to concentrations in cell lines maintained under normomagnesaemic conditions.^{57–60} The increased concentration of IL-1 and IL-17⁶¹ can result in establishment of positive feedback loops (which can occur locally or systemically depending on the disease in question; see reviews referenced^{62,63} for more details beyond the scope of this review), with TNF α facilitating the generation of IL-1 and IFN- γ , in turn driving further TNF α effects.^{52,64–66} Transcription of TNF α is governed by NF κ B, the inflammation-related transcriptional factor,⁶⁷ discussed in more detail below.

Negative feedback loops also operate, these are known to regulate TNF α levels by inhibition of transcription of TNF α mRNA.⁵² This negative feedback arises when TNF α stimulates the production of molecules that inhibits TNF α transcription including prostaglandins, IL-10 and corticosteroids. Mg²⁺ is also known to promote prostaglandin synthesis, and so it can be speculated that in the hypomagnesaemic individual, the negative feedback loop is disrupted, with less feedback control over elevated TNF- α levels.^{52,64,68–70} It remains to be determined what levels of magnesium deficiency tip the balance towards uncontrolled positive feedback in humans, resulting in clinically observable effects. Nor is it known how prolonged such effects would be. There is a paucity of data regarding the effects of Mg²⁺ on IL-10 or corticosteroid levels, hence the interaction between Mg²⁺ and the endocrine system is ripe for investigation.

After translation, TNF α exists in a cell-surface bound precursor form, termed transmembrane TNF α (tmTNF α). This intermediate is converted to a soluble cytokine *via* TNF α -converting enzyme.^{71,72} Both tmTNF α and sTNF α are biologically active and capable of binding to their receptors, TNFR1 and TNFR2, triggering different downstream signalling events, which result in outcomes such as apoptosis, necroptosis, transcriptional factor AP-1 activation, or NF κ B activation and translocation to the nucleus.⁷³ Once binding has occurred between TNF α and its receptor (either TNFR1 or 2), a conformational change occurs in the latter, followed by the interaction between the intracellular domains of TNFR and other proteins including TRAF2, cIAP1 and cIAP2, forming a complex. This complex formation leads to the activation of the I κ B kinase (IKK) complex. The I κ B complex consists of two kinase subunits, which phosphorylate the NF κ B inhibitor protein I κ B α , tagging it for the ubiquitin-proteasome pathway of degradation. This allows NF κ B to freely translocate to the nucleus where it coordinates the transcription of genes such as cIAP1/2 and TRAF2, which are important in the regulation of NF κ B and apoptotic pathways.^{74,75}

Interplay and importance of Ca²⁺ and Mg²⁺ signalling

NF κ B activity is regulated by various secondary messengers including intracellular calcium ions (Ca²⁺_i). In rats fed a Mg²⁺ deficient diet, where plasma Mg²⁺ fell to 60% of control levels, a rise in Ca²⁺_i levels was observed.⁷⁶ This rise in Ca²⁺_i secondary to a decrease in Mg²⁺ is seen in a variety of systems including human patient studies, rodent models and cell culture (including immune cells), all of which are mentioned in the 2010 review by Rayssiguier *et al.*⁷⁷ The increased Ca²⁺_i can induce formation of reactive oxygen intermediates following an oxidative burst from cells such as neutrophils,⁷⁸ which in turn cause phosphorylation of I κ B (through an as yet unknown mechanism), which will release the active form of NF κ B for nuclear translocation.

The critical role of Ca²⁺ in the NF κ B pathway is supported by the observation that Ca²⁺ chelators prevent the induction of NF κ B activity *in vivo* in murine models.⁷⁹ Other research has shown that when Mg²⁺ deficient rats

were fed a Ca²⁺ deficient diet, the inflammatory effect was greatly reduced (as measured by reduced inflammation scores, prevention of leucocytosis and reduced splenomegaly) when compared to other hypomagnesaemic rats.⁸⁰ Once the active NF κ B crosses the nucleus it up-regulates the transcription of TNF α .^{81,82} It should be mentioned, however, that the evidence for NF κ B-mediated TNF α expression is mostly limited to murine models, and studies relating to humans are limited.^{83,84} However, in 2010 one study utilized mouse bone marrow-derived dendritic cells to demonstrate NF κ B-mediated positive expression of the TNF α gene.⁸¹ Additionally it would be of interest to study whether a high Ca²⁺, low Mg²⁺ state up-regulates NF κ B activity and TNF α expression permitting TNF α synergism with STAT6 to switch B cells to IgE production.⁸⁵ This is important to establish the role of IgE in atopic and inflammatory conditions such as AD.⁸⁵

The antagonism between Mg²⁺ and Ca²⁺, and competition for binding sites on receptors, enables Mg²⁺ to overcome the toxic effects produced by excessive Ca²⁺ concentrations in cells of the immune system that are located in the brain (human microglial cells in tissue culture).⁸⁶ In the case of neuroinflammation, *in vitro* experimentation has shown that an influx of Ca²⁺ into microglia (brain resident macrophages) and THP-1 cells activates their associated purinergic receptors and subsequently inflammation.⁸⁷ Mg²⁺ is effective in ameliorating the neurotoxic effect produced by over-activation of human microglial cells that occurs as a result of elevated levels of inflammatory cytokines in the cells such as TNF- α , IL-6 and nitrite ions.^{86,88} These agents are released as a result of intracellular inflammatory pathway activation, *via* P38 MAPK and NF κ B.⁸⁶

Pro-inflammatory cytokine responses and resident microflora

In addition to stimulating TNF α transcription, NF κ B activates the transcription of IL-6 and IFN- γ after binding to the cognate promoter regions.^{81,89,90} These potent pro-inflammatory cytokines have been shown to be present in high levels during periods of Mg²⁺ deficiency.^{91–93} Additionally, in one study of septic shock using *ex vivo* human whole-blood, it was shown that following addition of Mg²⁺, the baseline level of TNF α and IL-6 production fell.⁹⁴ This observation provides further evidence of the inverse relationship between these inflammatory mediators and magnesium concentration.⁹⁴ However, the concentration of Mg²⁺ was well above physiologically relevant levels.⁹⁴ More research in other conditions, using a similar experimental set up is needed to further elucidate links between Mg²⁺ levels in human blood and pro-inflammatory cytokine production. Further, a more recent study, using THP-1 cells isolated from human neonatal cord blood, found that treatment with Mg²⁺ (at levels known to be clinically effective *in vivo*) reduced the production of IL-1 β , TNF- α and IL-8 cytokines and IL-6 in cord blood monocytes.⁹⁵

IL-6 exerts effects on a variety of cells including T and B lymphocytes, hepatocytes, hematopoietic progenitor cells

and fibroblasts, with consequent systemic effects such as acute phase reactant protein production, immunoglobulin synthesis and naive CD4⁺ T cell differentiation into Th17 cells. IL-17 secreted from Th17 cells is responsible for autoimmune tissue injury.⁹⁶ In relation to skin pathology, it has been shown that the epidermis from psoriatic skin produces high levels of IL-6 in addition to over-expression of TNFR, due in turn to the higher levels of IFN- γ .⁹⁷ Many of the characteristic phenotypic features of keratinocytes from psoriatic skin, including growth activation and ICAM-1 up-regulation, are a result of the actions of IL-6, TNF α and IFN- γ .^{97,98} The histological features of psoriasis such as the presence and accumulation of inflammatory cells (including polymorphonuclear leukocytes) and epidermal hyperplasia have been attributed to elevated TNF α activity.^{99,100} Given the aforementioned studies⁹⁷ indicating a role for magnesium in modulating the production of pro-inflammatory cytokines such as IL-6 and TNF α , there may be a role for magnesium-containing compounds in the treatment of psoriasis. However, whether such treatment would be effective in either acute or chronic cases, differing levels of severity, or what the most effective dose and route of administration would be (i.e. topical or oral) remain to be shown.

Another interesting finding is that mice fed on a Mg²⁺ deficient diet for four days showed higher levels of IL-6 and TNF α mRNA in the liver and intestine, a drop in the levels of the mRNA of zonula occludens-1, occludin and proglucagon in the ileum (three factors controlling gut barrier integrity and function) and possessed reduced gut bifidobacteria levels when compared to controls.¹⁰¹ *Bifidobacterium* strains have been shown to repress inflammation in a variety of situations including studies of ulcerative colitis and skin inflammation (i.e. acne).^{102–104} With respect to skin inflammation, *Bifidobacterium* strains appear to facilitate reduction in substance P, a molecule that increases TNF α expression. One study in human female volunteers demonstrated a reduction in sensitive skin and heightened resistance to physical and chemical insults to the skin (in contrast to a negative control cream) following topical application of *B. longum* spp. products. Additionally, in the same paper, the authors report a statistically significant reduction in markers of inflammation (including oedema, mast cell degranulation and TNF α release) following the application of a *B. longum* preparation on *ex-vivo* human skin explants.¹⁰⁴

However, in the above mentioned¹⁰¹ mouse study, the animals on a Mg²⁺-deficient diet for 21 days demonstrated a potentially compensatory increase in caecal *Bifidobacteria* levels, restoration of intestinal barrier function and a waning of inflammation when compared to control mice. This is in contrast to mice on the same diet for four days only; in the latter case decreased levels of *Bifidobacteria*, and an increase in IL-6, TNF α and other markers of pathology were present. Drawing generalizable conclusions from this study is difficult given that it was a mouse-model study (not human) and measurements of bacteria and relevant mRNA levels were not continued after 21 days. If there was adaptation, were there any other long-term adverse effects on physiology? In the future it will also be important

to work out the complex relationship between the gut bacteria and cytokine responses; is it bidirectional or unidirectional? What feedback loops exist? Moreover, the regulation is unlikely to be the same for all cytokines. These are all questions that need to be investigated.

Changes in macrophages and neutrophils

In rodent studies, a state of hypomagnesaemia has been associated with the activation of macrophages and neutrophils.^{105,106} It has been suggested that this is due to the increase in circulating pro-inflammatory mediators, including substance P, IL-6, TNF α and IFN- γ ; however, given that macrophages and neutrophils are amongst the cells that produce these cytokines, it remains difficult to say whether the aforementioned cells are activated by these substances or if they are activated directly by the low levels of Mg²⁺.¹⁰⁷ Malpuech-Brugère and colleagues suggest that it is due to the latter, a drop in circulating Mg²⁺ concentration, which would lead to a significant increase in Ca²⁺ levels,^{37,108} in turn stimulating cellular proliferation. Whilst macrophages are also able to be activated independently of Ca²⁺_i (for example, *via* the lipopolysaccharide pathway), the Ca²⁺-dependent activation pathway results in a more rapid expression of IL-6.¹⁰⁸

Aside from enhanced protein expression of IL-6 and TNF α , activated macrophages also demonstrate increased expression of IL-1 proteins in Mg²⁺-deficient rats when compared to a control group. In the same study it was postulated that this contributed to cardiac lesions (IL-1 promotes expression of endothelin from heart endothelial cells which causes vasospasm).¹⁰⁶ Additionally, IL-1 is antagonistic to endothelial proliferation, suggesting another mechanism behind the lesions seen.¹⁰⁹ Furthermore, in the skin, IL-1 is implicated in wound repair and skin pathology, with IL-1 receptor (IL-1R) knockout mice demonstrating reduced cutaneous and deep tissue fibrosis and scarring and restoration of skin architecture.¹¹⁰

Neutrophils are capable of releasing superoxide anions *via* their NADPH oxidase system and can contribute to tissue damage during Mg²⁺ deficiency, as demonstrated in a study which also demonstrated phagocytic activity of neutrophils in rats on a magnesium deficient diet.¹⁰⁵ The study also showed that the free radical production from neutrophils is inhibited when high Mg²⁺ levels are present in the extracellular space.¹⁰⁵ Mg²⁺ was also found to inhibit superoxide in cultured human neutrophils.¹¹⁰ The latter experiment also revealed synergistic inhibition of superoxides with the addition of zinc ions to the magnesium solutions.¹¹⁰ Neutrophil migration to the skin, a process that promotes keratinocyte apoptosis is TNF α dependant. These phenomena are key features of hyper-proliferative skin diseases such as psoriasis.¹¹¹

Clinical implications of magnesium deficiency

Magnesium is a micronutrient required for normal growth and development. Numerous clinical disorders have been associated with magnesium deficiency. Inflammation is a primary reaction brought about by magnesium deficiency,

creating oxidative stress and subsequent immune stress. The clinical disorders could be a consequence of this stress response.^{91,112,113} Magnesium ions bind to macromolecules and cell membranes. Mg^{2+} is known to affect cellular functions, including the transport of potassium and Ca^{2+} , modulation of signal transduction, cell proliferation and energy metabolism.¹¹⁴ Early stages of Mg^{2+} deficiency can be characterized by a wide range of symptoms such as anorexia, vomiting, weakness, paraesthesia, muscular cramps, irritability and impaired cognitive functioning reflected by a decreased attention span. Mg^{2+} deficiency is related to poor dietary Mg^{2+} intake, often as a result of lifestyle changes, leading to the aforementioned health disorders.^{114–116} Mg^{2+} deficiency triggers inflammatory responses, including abnormal calcium homeostasis, activation of *N*-methyl-D-aspartate (NMDA) receptors, release of neurotransmitters, membrane oxidation and activation of NF κ B (see previous section, 'Magnesium in TNF α and NF κ B signalling').^{76,91} Some of the important inflammation-related clinical disorders, known to be caused by or exacerbated by, magnesium deficiency, are outlined below.

The pathogenesis of asthma, a chronic inflammatory disorder involves activation of NF κ B and, expression of pro-inflammatory cytokines, chemokines and inflammatory mediators (IFN- γ and ROS).¹¹⁷ In this pathological condition, NF κ B activation leads to dysregulation of cytokines and infiltration of inflammatory cells such as mononuclear cells and fibroblasts in the lung.^{117,118} Indeed, increased NF κ B activity has been observed in the airways of asthmatic patients.¹¹⁸ In acute asthma, the therapeutic effect of magnesium is well established.¹² Studies in acute asthma have shown that intravenous and inhaled magnesium sulphate ($MgSO_4$) improved lung function and reduced hospitalization frequency, particularly in patients with the lowest levels of forced expired volume.¹¹⁶ In chronic asthma patients with persistent airflow limitation short-term treatment with magnesium inhalations had no statistically significant, direct bronchodilating effect, however, clinical observations suggested heterogeneity in the response, probably related to treatment intensity, and supported further exploration of magnesium administration in those patients.¹¹⁶

A characteristic property of Mg^{2+} is its antagonism of Ca^{2+} (see prior section 'Interplay and importance of Ca^{2+} and Mg^{2+} signalling'). It competes with Ca^{2+} for entry into cells through voltage-gated channels and receptors and inhibits Ca^{2+} release from the sarcoplasmic reticulum.^{119,120}

The synovial fluid from patients with rheumatoid arthritis contains elevated levels of TNF α (an activator of NF κ B), which is important in the pathogenesis.^{118,121} Mg^{2+} deficiency can lead to lipid peroxidation and membrane oxidation, which in turn activates the NF κ B pathway.^{91,122} Activation of inflammatory responses due to Mg^{2+} deficiency causes chronic inflammation leading to different types of arthritis, depending on the site of NF κ B activation.

Studies in humans indicate that low Mg^{2+} intake and blood plasma concentration are linked with enhanced risk of atherosclerotic disease.¹²³ Atherosclerosis is currently classified as an inflammatory disease, having interactions

between modified lipoproteins, macrophages, T lymphocytes and the components of arterial walls,⁹¹ leading to the development of atherosclerotic lesions. Experimental results suggest regression of such lesions and suppression of atherogenesis in low-density lipid receptor deficient mice fed with Mg^{2+} supplement.^{124,125} Infusion of Mg^{2+} at supra-physiological concentrations causes vasodilation of coronary arteries and systemic vasculature, antiarrhythmic effects and platelet inhibition.¹²⁶ Studies also show dietary administration of Mg^{2+} attenuates atherosclerotic lesions by lowering serum cholesterol and triglycerides in cholesterol fed animals.⁹¹ On the basis of these studies, it appears that Mg^{2+} concentration regulates lipid metabolism and reduces atherosclerosis in animal models.⁹¹

Even though the immune system and, in particular, the inflammatory response operates systemically, the inter-relationship between inflammation occurring in the nervous system and systemic inflammation needs to be better understood. It is widely accepted that several neurological disorders are characterised by an inflammatory component.^{127,128} There are several drugs undergoing test that are posited to act by reducing neurodegeneration, at least in part through inhibition of the inflammatory response of glial cells.¹²⁹ However, these drugs exert their effect throughout the body, resulting in global immunosuppression.¹³⁰ It would be ideal for such drugs to specifically target the glial cells and control inflammation in the brain without producing systemic immunosuppression.^{129,131} Studies conducted in mice using the compound 4,6-diphenyl-3-(4-(pyrimidin-2-yl)piperazin-1-yl)pyridazine (MW01-5-188WH), aimed at selective suppression of neuroinflammation, has yielded some positive outcomes without producing extra-neural inflammation.¹²⁹

Mg^{2+} administration could potentially be an effective treatment of neurodegenerative diseases *via* its antagonism of Ca^{2+} channels. This selectively suppresses neuroinflammation.^{86,91} If treatment for neurodegenerative diseases involved administration of Mg^{2+} locally to the brain, it could conceivably avoid the generalised stress on the immune system that is caused by non-targeted anti-inflammatory drugs. Experiments conducted on rat ischaemic and excitotoxic brain injury models shows the activity of Mg^{2+} as a neuroprotective agent.^{114,132,133} This is achieved by Mg^{2+} blockade of NMDA receptors and enhancement of regional cerebral blood flow to ischaemic areas of the brain. Mg^{2+} also inhibits entry of Ca^{2+} into cells through voltage-operated and receptor-operated channels.^{114,126}

The nervous and immune systems interact bi-directionally. Mg^{2+} deficiency is known to induce a systemic stress response by activating neuroendocrine pathways, modifying production and activity of neuromediators such as acetylcholine, catecholamines and substance P. These have well established roles in the progression of both local and systemic inflammatory responses.¹³⁴ Administration of Mg^{2+} has been shown to block Ca^{2+} traffic through cell surface channels, acting as a broad inhibitor of neuroinflammation.^{86,135} Elevated systemic levels of Mg^{2+} have been shown to reduce damaging consequences of Ca^{2+} induced neuroinflammation in Parkinson's disease and Alzheimer's disease.⁸⁶

AD is a skin disease that is a result of interactions between skin, nervous system and immune cells. Nerve growth factor (NGF) is a neurotrophin, mainly produced in the basal keratinocytes and are present in elevated levels in plasma of AD patients.¹³⁶ In normal skin, less expression of NGF was found.^{137,138} These NGFs are known to be involved in the extension of C-fibres and promote increased density of nerve endings. In AD this results in exposure of a high density of unprotected nerves to external conditions, leading to the itch and scratch cycle that characterizes the disease.^{14,139} Semaphorin3A (Sema3A) is another factor involved in the development of AD. It is an axon guidance molecule that inhibits outgrowth of sensory neurons. It does so by binding to plexin-A1-4 and its co-receptor neuropilin-1 (NRP-1), subduing the effects caused by NGFs.^{140,141} Sema3A acts by suppressing nerve extension, and inhibition of histamine release from mast cells (existing treatments for AD rely on blocking the histamine receptors (H1-R) with anti-histamines or topical steroids). Further, Sema3A also binds to NRP-1.¹⁴² NRP-1 is known to activate the NFκβ pathway and to initiate keratinocyte proliferation.¹⁴³

One study has shown that increased calcium ion concentration (0.45–0.75 mmol/L) in normal human epidermal keratinocytes augments the expression of Sema3A.¹⁴⁴ While calcium and magnesium have antagonist effects at the cellular level, it is possible that the skin barrier recovery after treatment with Dead Sea minerals (comprising calcium and magnesium salts) is due to the combined role of calcium in upregulating Sema3A, and the prevention by magnesium of mast cell degranulation through other mechanisms (potentially *via* effects on TNFα or STAT-6^{145,146}). However, a direct role for magnesium ions in regulation of Sema3A and NGFs remains to be established, and is an important area for further investigation, given the role of these factors in AD.

In relation to skin disease, a clinical study was conducted on 30 AD candidates, in which subjects were tested over six weeks for transepidermal water loss (TEWL), skin hydration, skin redness and skin roughness.³ Upon treating one of their arms with a 5% Dead Sea salt solution at 38–42°C and the other arm with tap water (38–42°C) as control, an improvement in TEWL, with reduction in AD symptoms in the Dead Sea salt treated group was shown.³ Further work is needed, to confirm these studies and to measure the intracellular and molecular correlates of the structural changes in the skin.

Conclusion

Absorption of Mg²⁺ ions across the normal SC could occur under conditions of elevated temperature or changed hydration conditions (for example high salt concentrations). Absorption of magnesium will take place in cases of skin pathology or injury, where there is physical disruption of the SC. Subsequently, transmembrane proteins, such as SLC41A2 could assist intercellular transport of magnesium ions, leading to further penetration through the organ systems.²⁷ The action of magnesium ion as an anti-inflammatory agent could be *via* several pathways, such as activation

of the TPP-dependant riboswitch.³⁸ Magnesium deficiency results in activation of TNFα and NFκβ, which can further facilitate pro-inflammatory cytokines.^{76,106} It would also be of interest to study the effect of Mg²⁺ on the synergism between TNF-α and STAT6, a mediator of IgE receptor mediated mast cell responses in late phase allergic responses and AD.^{145,146} Experimental data from humans and mice suggest an inverse relationship between magnesium intake and TNFα concentration, in addition to several other markers of inflammation.^{147–149} The hypomagnesaemic condition increases the influx of calcium into cells, resulting in elevated NFκβ activity.^{150,151} Evidence for activation of neutrophils and macrophages by calcium ions in mice has also been found.¹⁵⁰ Collectively, the inflammatory responses triggered by magnesium deficiency can result in clinical disorders. The interaction between Mg²⁺ and inflammatory mediators is ripe for investigation. For example, there is a paucity of data relating to the effects of Mg²⁺ on IL-10 or corticosteroid concentrations. More research is needed to further elucidate links between Mg²⁺ levels in human blood and pro-inflammatory cytokines. The effect of treatment with magnesium containing compounds in acute or chronic diseases with differing levels of severity, and the most effective doses and routes of administration in these cases, remain to be systematically determined. Although a role of magnesium deficiency in neurodegenerative disease is established, and may be mediated by interaction of magnesium ions with glial cells,¹³⁴ the inter-relationship between inflammation in the nervous system and systemic inflammation needs to be better understood. Thus, there exist plausible mechanisms by which several metabolic and inflammatory conditions might potentially be alleviated through magnesium administration, either systemically or locally. These mechanisms are ripe for further investigation.

Author contributions: All authors participated in the writing, review and editing of this manuscript. NCC and CW contributed equally to the writing of this manuscript.

ACKNOWLEDGMENTS

This work was funded by grants from the University of Queensland, The National Health and Medical Research council of Australia, and Cancer Council Queensland.

REFERENCES

1. Sukenik S, Abu-Shakra M, Kudish S, Flusser D. Dead Sea and Tiberias as health resort areas for patients suffering from different types of arthritis. *Harefuah* 2006;**145**:117–22
2. Shani J, Eevn-paz Z, Avrach WW, Rubinstein N, Livshin R, Justesen NPB, Harkmark W. Topical replacement therapy of psoriasis by Dead Sea salts. *Dermatosen* 1991;**39**:49–53
3. Proksch E, Nissen HP, Bremgartner M UC. Bathing in magnesium rich Dead Sea salt improves skin barrier function, enhances skin hydration, and reduces inflammation in atopic dry skin. *Int J Dermatol* 2005;**44**:151–7
4. Durlach J, Guiet-Bara a, Pagès N, Bac P, Bara M. Magnesium chloride or magnesium sulfate: a genuine question. *Magnes Res* 2005;**18**:187–92
5. Brisson P. Percutaneous absorption. *Can Med Assoc J* 1974;**110**:1182–5
6. Kligman AM. A biological brief on percutaneous absorption. *Drug Dev Ind Pharm* 1983;**9**:521–60

7. Winkelmann RK. The relationship of the structure of the epidermis to percutaneous absorption. *Br J Dermatol* 1969;**81**:11–22
8. Lansdown AB. Physiological and toxicological changes in the skin resulting from the action and interaction of metal ions. *Crit Rev Toxicol* 1995;**25**:397–462
9. Hostýnek JJ, Hinz RS, Lorence CR, Price M, Guy RH. Metals and the skin. *Crit Rev Toxicol* 1993;**23**:171–235
10. Jahnhen-Dechent W, Ketteler M. Magnesium basics. *Clin Kidney J* 2012;**5**:i3–i14
11. Abels DJ, Even-Paz Z, Efron D. Bioclimatology at the Dead Sea in Israel. *Clin Dermatol* 2014;**14**:653–8
12. Boaz M, Shtendik L, Oron M, Portugal-Cohen M, Kohen R, Biro A, Cernes R, Barnea Z, Maor Z, Katzir Z. A randomized controlled clinical trial comparing the efficacy of Dead Sea mineral-enriched body lotion versus two types of placebo in the treatment of cutaneous dryness, itching, peeling and tightness in hemodialysis patients. *Nephron Clin Pract* 2009;**113**:169–76
13. Halevy S, Giryes H, Friger M, Sukenik S. Dead sea bath salt for the treatment of psoriasis vulgaris: a double-blind controlled study. *J Eur Acad Dermatol Venereol* 1997;**9**:237–42
14. Takano N, Sakurai T, Kurachi M. Effects of anti-nerve growth factor antibody on symptoms in the NC/Nga mouse, an atopic dermatitis model. *J Pharmacol Sci* 2005;**99**:277–86
15. Washington N, Washington C, Wilson CG. Structure of the skin; passage of drug through the skin. *Physiological pharmaceuticals: barriers to drug absorption*. London: Taylor and Francis Group, 2001, pp. 182–7
16. Denda M, Katagiri C, Hirao T, Maruyama N, Takahashi M. Some magnesium salts and a mixture of magnesium and calcium salts accelerate skin barrier recovery. *Arch Dermatol Res* 1999;**291**:560–3
17. Ainsworth C. Skin: into the breach. *Nature* 2011;**479**:S12–13
18. Brown P. Atopy: marching with allergies. *Nature* 2011;**479**:S14–15
19. Denda M. Skin barrier function as a self-organizing system. *Cell* 2000;**15**:227–32
20. Maguire ME, Cowan JA. Magnesium chemistry and biochemistry. *Biometals* 2002;**15**:203–10
21. McCallum DI, Hall GF. Umbilical granulomata—with particular reference to talc granuloma. *Br J Dermatol* 1970;**83**:151–6
22. Tye MJ, Hashimoto K, Fox F. Talc granulomas of the skin. *JAMA* 1966;**198**:1370–2
23. Kopito L, Elian E, Shwachman H. Sodium, potassium, calcium, and magnesium in hair from neonates with cystic fibrosis and in amniotic fluid from mothers of such children. *Pediatrics* 1972;**49**:620–4
24. Elias PM, Tsai J, Menon GK, Holleran WM, Feingold KR. The potential of metabolic interventions to enhance transdermal drug delivery. *J Invest Dermatol Symp Proc* 2002;**7**:79–85
25. Diebler H, Eigen M, Ilgenfritz G, Maass G, Winkler R. Kinetics and mechanism of reactions of main group metal ions with biological carriers. *Pure Appl Chem Int Union Pure Appl Chem* 1969;**20**:93–116
26. Eigen M. Fast elementary steps in chemical reaction mechanisms. *Pure Appl Chem Int Union Pure Appl Chem* 1963;**6**:97–116
27. Sahni J, Nelson B, Scharenberg AM. SLC41A2 encodes a plasma-membrane Mg^{2+} transporter. *Biochem J* 2007;**401**:505–13
28. Chen H-C, Su L-T, González-Pagán O, Overton JD, Runnels LW. A key role for $Mg(2+)$ in TRPM7's control of ROS levels during cell stress. *Biochem J* 2012;**445**:441–8
29. Sahni J, Scharenberg AM. TRPM7 ion channels are required for sustained phosphoinositide 3-kinase signaling in lymphocytes. *Cell Metab* 2008;**8**:84–93
30. Rojanasakul Y, Wang LY, Bhat M, Glover DD, Malanga CJ, Ma JK. The transport barrier of epithelia: a comparative study on membrane permeability and charge selectivity in the rabbit. *Pharm Res* 1992;**9**:1029–34
31. Piemi MP, Korner D, Benita S, Marty JP. Positively and negatively charged submicron emulsions for enhanced topical delivery of anti-fungal drugs. *J Control Release* 1999;**58**:177–87
32. Singh I, Morris AP. Performance of transdermal therapeutic systems: effects of biological factors. *Int J Pharm Invest* 2011;**1**:4–9
33. Roberts MS, Cross SE, Pellet MA. *Skin transport. Dermatological and transdermal formulations*. New York: Markel Dekker, 2002, pp. 1–30
34. Park J-H, Lee J-W, Kim Y-C, Prausnitz MR. The effect of heat on skin permeability. *Int J Pharm* 2008;**359**:94–103
35. Hirvonen J, Murtomäki L, Kontturi K. Effect of diffusion potential, osmosis and ion-exchange on transdermal drug delivery: theory and experiments. *J Control Release* 1998;**56**:33–9
36. Williams AC, Barry BW. Penetration enhancers. *Adv Drug Deliv Rev* 2012;**64**:128–37
37. Nowacki W, Daveau M, Malpuech-Bruge C. Inflammatory response following acute magnesium deficiency in the rat. *Biochim Biophys Acta* 2000;**1501**:91–8
38. Yamauchi T, Miyoshi D, Kubodera T, Nishimura A, Nakai S, Sugimoto N. Roles of Mg^{2+} in TPP-dependent riboswitch. *FEBS Lett* 2005;**579**:2583–8
39. Kubodera T, Watanabe M, Yoshiuchi K, Yamashita N, Nishimura A, Nakai S, Gomi K, Hanamoto H. Thiamine-regulated gene expression of *Aspergillus oryzae* thiA requires splicing of the intron containing a riboswitch-like domain in the 5'-UTR. *FEBS Lett* 2003;**555**:516–20
40. Lai EC. RNA sensors and riboswitches: self-regulating messages. *Curr Biol* 2003;**13**:R285–91
41. Winkler W, Nahvi A, Breaker RR. Thiamine derivatives bind messenger RNAs directly to regulate bacterial gene expression. *Nature* 2002;**419**:952–6
42. Kumar A, Rajput CS, Bhati SK. Synthesis of 3-[4'-(p-chlorophenyl)-thiazol-2'-yl]-2-[(substituted azetidinone/thiazolidinone)-amino-methyl]-6-bromoquinazolin-4-ones as anti-inflammatory agent. *Bioorg Med Chem* 2007;**15**:3089–96
43. Holla BS, Malini KV, Rao BS, Sarojini BK, Kumari NS. Synthesis of some new 2,4-disubstituted thiazoles as possible antibacterial and anti-inflammatory agents. *Eur J Med Chem* 2003;**38**:313–8
44. Kalkhambkar RG, Kulkarni GM, Shivkumar H, Rao RN. Synthesis of novel triheterocyclic thiazoles as anti-inflammatory and analgesic agents. *Eur J Med Chem* 2007;**42**:1272–6
45. Rostom SAF, el-Ashmawy IM, Abd el Razik HA, Badr MH, Ashour HMA. Design and synthesis of some thiazolyl and thiadiazolyl derivatives of antipyrine as potential non-acidic anti-inflammatory, analgesic and antimicrobial agents. *Bioorg Med Chem* 2009;**17**:882–95
46. Koufaki M, Kiziridi C, Nikoloudaki F, Alexis MN. Design and synthesis of 1,2-dithiolane derivatives and evaluation of their neuroprotective activity. *Bioorg Med Chem Lett* 2007;**17**:4223–7
47. Guerrero-Romero F, Rodríguez-Morán M. Hypomagnesemia, oxidative stress, inflammation, and metabolic syndrome. *Diabetes Metab Res Rev* 2006;**22**:471–6
48. Rude RK, Wei L, Norton HJ, Lu SS, Dempster DW, Gruber HE. TNF α receptor knockout in mice reduces adverse effects of magnesium deficiency on bone. *Growth Factors* 2009;**27**:370–6
49. Aringer M, Feierl E, Steiner G, Stummvoll G, Höfler E, Steiner C, Radda I, Smolen J, Graninger W. Increased bioactive TNF in human systemic lupus erythematosus: associations with cell death. *Lupus* 2002;**11**:102–8
50. Rodríguez-Morán M, Guerrero-Romero F. Elevated concentrations of TNF- α are related to low serum magnesium levels in obese subjects. *Magnes Res* 2004;**17**:189–96
51. Sugimoto J, Romani AM, Valentin-Torres AM, Luciano AA, Ramirez Kitchen CM, Funderburg N, Mesiano S, Bernstein HB. Magnesium decreases inflammatory cytokine production: a novel innate immunomodulatory mechanism. *J Immunol* 2012;**188**:6338–46
52. Tracey D, Klareskog L, Sasso EH, Salfeld JG, Tak PP. Tumor necrosis factor antagonist mechanisms of action: a comprehensive review. *Pharmacol Ther* 2008;**117**:244–79
53. Silswal N, Singh AK, Aruna B, Mukhopadhyay S, Ghosh S, Ehtesham NZ. Human resistin stimulates the pro-inflammatory cytokines TNF- α and IL-12 in macrophages by NF- κ B-dependent pathway. *Biochem Biophys Res Commun* 2005;**334**:1092–101
54. Bowcock AM, Krueger JG. Getting under the skin: the immunogenetics of psoriasis. *Nat Rev Immunol* 2005;**5**:699–711
55. Stannus O, Jones G, Cicuttini F, Parameswaran V, Quinn S, Burgess J, Ding C. Circulating levels of IL-6 and TNF- α are associated with knee radiographic osteoarthritis and knee cartilage loss in older adults. *Osteoarthritis Cartilage* 2010;**18**:1441–7

56. Bussi re FI, Tridon A, Zimowska W, Mazur A, Rayssiguier Y. Increase in complement component C3 is an early response to experimental magnesium deficiency in rats. *Life Sci* 2003;**73**:499–507
57. Weglicki WB. Hypomagnesemia and inflammation: clinical and basic aspects. *Annu Rev Nutr* 2012;**32**:55–71
58. Barbagallo M, Dominguez LJ. Magnesium and aging. *Curr Pharm Des* 2010;**16**:832–9
59. Ferr  S, Baldoli E, Leidi M, Maier JAM. Magnesium deficiency promotes a pro-atherogenic phenotype in cultured human endothelial cells via activation of NF B. *Biochim Biophys Acta* 2010;**1802**:952–8
60. Maier JAM, Malpuech-bruge C, Rayssiguier Y, Mazur A. Low magnesium promotes endothelial cell dysfunction: implications for atherosclerosis, inflammation and thrombosis. *Biochim Biophys Acta* 2004;**1689**:13–21
61. Ogura H, Murakami M, Okuyama Y, Tsuruoka M, Kitabayashi C, Kanamoto M, Nishihara M, Iwakura Y, Hirano T. Interleukin-17 promotes autoimmunity by triggering a positive-feedback loop via interleukin-6 induction. *Immunity* 2008;**29**:628–36
62. Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 cells. *Annu Rev Immunol* 2009;**27**:485–517
63. Weber A, Wasiliew P, Kracht M. Interleukin-1 (IL-1) pathway. *Sci Signal* 2010;**3**:1
64. Bachwich PR, Chensue SW, Larrick JW, Kunkel SL. Tumor necrosis factor stimulates interleukin-1 and prostaglandin E2 production in resting macrophages. *Biochem Biophys Res Commun* 1986;**136**:94–101
65. Dinarello CA, Cannon JG, Wolff SM, Bernheim HA, Beutler B, Cerami A, Figari IS, Palladino MA, O'Connor JV. Tumor necrosis factor (cachectin) is an endogenous pyrogen and induces production of interleukin 1. *J Exp Med* 1986;**163**:1433–50
66. Beutler B, Cerami A. Cachectin and tumour necrosis factor as two sides of the same biological coin. *Nature* 1986;**320**:584–8
67. Collart MA, Baeuerle P, Vassalli P. Regulation of tumor necrosis factor alpha transcription in macrophages: involvement of four kappa B-like motifs and of constitutive and inducible forms of NF-kappa B. *Mol Cell Biol* 1990;**10**:1498–506
68. Foey AD, Parry SL, Williams LM, Feldmann M, Foxwell BM, Brennan FM. Regulation of monocyte IL-10 synthesis by endogenous IL-1 and TNF-alpha: role of the p38 and p42/44 mitogen-activated protein kinases. *J Immunol* 1998;**160**:920–8
69. De Jong EC, Vieira PL, Kalinski P, Kapsenberg ML. Corticosteroids inhibit the production of inflammatory mediators in immature monocyte-derived DC and induce the development of tolerogenic DC3. *J Leukoc Biol* 1999;**66**:201–4
70. Abbott L, Nadler J, Rude RK. Magnesium deficiency in alcoholism: possible contribution to osteoporosis and cardiovascular disease in alcoholics. *Alcohol Clin Exp Res* 1994;**18**:1076–82
71. Black RA. Tumor necrosis factor-  converting enzyme. *Int J Biochem Cell Biol* 2002;**34**:1–5
72. Kriegler M, Perez C, DeFay K, Albert I, Lu SD. A novel form of TNF/cachectin is a cell surface cytotoxic transmembrane protein: ramifications for the complex physiology of TNF. *Cell* 1988;**53**:45–53
73. Cabal-Hierro L, Lazo PS. Signal transduction by tumor necrosis factor receptors. *Cell Signal* 2012;**24**:1297–305
74. Vince JE, Pantaki D, Feltham R, Mace PD, Cordier SM, Schmukle AC, Davidson AJ, Callus BA, Wong WW-L, Gentle IE, Carter H, Lee EF, Walczak H, Day CL, Vaux DL, Silke J. TRAF2 must bind to cellular inhibitors of apoptosis for tumor necrosis factor (tnf) to efficiently activate nf- B and to prevent tnf-induced apoptosis. *J Biol Chem* 2009;**284**:35906–15
75. Wang Y, Zhang P, Liu Y, Cheng G. TRAF-mediated regulation of immune and inflammatory responses. *Sci China Life Sci* 2010;**53**:159–68
76. Malpuech-Brug re C, Rock E, Astier C, Nowacki W, Mazur A, Rayssiguier Y. Exacerbated immune stress response during experimental magnesium deficiency results from abnormal cell calcium homeostasis. *Life Sci* 1998;**63**:1815–22
77. Rayssiguier Y, Libako P, Nowacki W, Rock E. Magnesium deficiency and metabolic syndrome: stress and inflammation may reflect calcium activation. *Magnes Res* 2010;**23**:73–80
78. Waddell TK, Fialkow L, Chan CK, Kishimoto TK, Downey GP. Potentiation of the oxidative burst of human neutrophils. A signaling role for L-selectin. *J Biol Chem* 1994;**269**:18485–91
79. Pahl HL, Baeuerle PA. Activation of NF-kappa B by ER stress requires both Ca²⁺ and reactive oxygen intermediates as messengers. *FEBS Lett* 1996;**392**:129–36
80. Bussi re FI, Gueux E, Rock E, Mazur A, Rayssiguier Y. Protective effect of calcium deficiency on the inflammatory response in magnesium-deficient rats. *Eur J Nutr* 2002;**41**:197–202
81. Yamauchi S, Ito H, Miyajima A. I B , a nuclear I B protein, positively regulates the NF- B-mediated expression of proinflammatory cytokines. *Proc Natl Acad Sci* 2010;**107**:11924–9
82. Drouet C. Enhancers and transcription factors controlling the inducibility of the tumor necrosis factor-  promoter. *J Immunol* 1991;**147**:1694–700
83. Baeuerle PA, Henkel T. Function and activation of NF-kappa B in the immune system. *Annu Rev Immunol* 1994;**12**:141–79
84. Shea LM, Beehler C, Schwartz M, Shenkar R, Tuder R. Hyperoxia activates NF-KB and increases TNF- . *J Immunol* 1996;**157**:3902–8
85. Geha RS, Jabara HH, Brodeur SR. The regulation of immunoglobulin E class-switch recombination. *Nat Rev Immunol* 2003;**3**:721–32
86. Lee M, Jantarotnotai N, McGeer E, McLarnon JG, McGeer PL. Mg²⁺ ions reduce microglial and THP-1 cell neurotoxicity by inhibiting Ca²⁺ entry through purinergic channels. *Brain Res* 2011;**1369**:21–35
87. Ac  a-Castillo C, Coddou C, Bull P, Brito J, Huidobro-Toro JP. Differential role of extracellular histidines in copper, zinc, magnesium and proton modulation of the P2X7 purinergic receptor. *J Neurochem* 2007;**101**:17–26
88. Zhao ML, Liu JS, He D, Dickson DW, Lee SC. Inducible nitric oxide synthase expression is selectively induced in astrocytes isolated from adult human brain. *Brain Res* 1998;**813**:402–5
89. Libermann TA, Baltimore D. Activation of interleukin-6 gene expression through the NF-KB transcription factor. *Mol Cell Biol* 1990;**10**:2327–34
90. Young HA, Hardy KJ. Role of interferon-gamma in immune cell regulation. *J Leukoc Biol* 1995;**58**:373–81
91. Mazur A, Maier JAM, Rock E, Gueux E, Nowacki W, Rayssiguier Y. Magnesium and the inflammatory response: potential physiopathological implications. *Arch Biochem Biophys* 2007;**458**:48–56
92. Weglicki WB, Chmielinska JJ, Kramer JH, Mak IT. Cardiovascular and intestinal responses to oxidative and nitrosative stress during prolonged magnesium deficiency. *Am J Med Sci* 2011;**342**:125–8
93. Weglicki WB, Dickens BF, Wagner TL, Chmielinska JJ, Phillips TM. Immunoregulation by neuropeptides in magnesium deficiency: ex vivo effect of enhanced substance P production on circulating T lymphocytes from magnesium-deficient mice. *Magnes Res* 1996;**9**:3–11
94. Nowacki W, Malpuech-Brug re C, Rock E, Rayssiguier Y. High-magnesium concentration and cytokine production in human whole blood model. *Magnes Res* 2009;**22**:93–6
95. Suzuki-Kakisaka H, Sugimoto J, Tatarbe M, Romani AM, Ramirez Kitchen CM, Bernstein HB. Magnesium sulfate increases intracellular magnesium reducing inflammatory cytokine release in neonates. *Am J Reprod Immunol* 2013;**70**:213–20
96. Tanaka T, Narazaki M, Kishimoto T. Therapeutic targeting of the interleukin-6 receptor. *Annu Rev Pharmacol Toxicol* 2012;**52**:199–219
97. Krueger JG, Krane JF, Carter DM, Gottlieb AB. Role of growth factors, cytokines, and their receptors in the pathogenesis of psoriasis. *J Invest Dermatol* 1990;**94**:135S–40S
98. Groves RW, Allen MH, Ross EL, Barker JN, MacDonald DM. Tumour necrosis factor alpha is pro-inflammatory in normal human skin and modulates cutaneous adhesion molecule expression. *Br J Dermatol* 1995;**132**:345–52
99. Biedermann T, Kneilling M, Mailhammer R, Maier K, Sander CA, Kollias G, Kunkel SL, H ltner L, R cken M. Mast cells control neutrophil recruitment during T cell-mediated delayed-type hypersensitivity reactions through tumor necrosis factor and macrophage inflammatory protein. *J Exp Med* 2000;**192**:1441–52
100. Chan JR, Blumenschein W, Murphy E, Diveu C, Wiekowski M, Abbondanzo S, Lucian L, Geissler R, Brodie S, Kimball AB,

- Gorman DM, Smith K, de Waal Malefyt R, Kastelein R a, McClanahan TK, Bowan EP. IL-23 stimulates epidermal hyperplasia via TNF and IL-20R2-dependent mechanisms with implications for psoriasis pathogenesis. *J Exp Med* 2006;**203**:2577–87
101. Pachikian BD, Neyrinck AM, Deldicque L, Backer FC De, Catry E, Dewulf EM, Sohet FM, Bindels LB, Everard A, Francaux M, Guiot Y, Cani PD, Delzenne NM. Changes in intestinal bifidobacteria levels are associated with the inflammatory response in magnesium-deficient mice. *J Nutr* 2010;**140**:509–14
 102. Setoyama H, Imaoka A, Ishikawa H, Umesaki Y. Prevention of gut inflammation by Bifidobacterium in dextran sulfate-treated gnotobiotic mice associated with bacteroides strains isolated from ulcerative colitis patients. *Microbes Infect* 2003;**5**:115–22
 103. Bowe WP, Logan AC. Acne vulgaris, probiotics and the gut-brain-skin axis - back to the future? *Gut Pathog* 2011;**3**:1
 104. Guéniche A, Bastien P, Ovine JM, Kermici M, Courchay G, Chevalier V, Breton L, Castiel-Higounenc I. Bifidobacterium longum lysate, a new ingredient for reactive skin. *Exp Dermatol* 2010;**19**:1–8
 105. Bussière FI, Gueux E, Rock E, Girardeau JP, Tridon A, Mazur A, Rayssiguier Y, Girardeau J. Increased phagocytosis and production of reactive oxygen species by neutrophils during magnesium deficiency in rats and inhibition by high magnesium concentration. *Br J Nutr* 2002;**87**:107–13
 106. Weglicki WB, Phillips TM, Freedman a M, Cassidy MM, Dickens BF. Magnesium-deficiency elevates circulating levels of inflammatory cytokines and endothelin. *Mol Cell Biochem* 1992;**110**:169–73
 107. Mak IT, Dickens BF, Komarov AM, Wagner TL, Phillips TM, Weglicki WB. Activation of the neutrophil and loss of plasma glutathione during Mg-deficiency—modulation by nitric oxide synthase inhibition. *Mol Cell Biochem* 1997;**176**:35–9
 108. Marriott I, Mason MJ, Elhofy A, Bost KL. Substance P activates NF- κ B independent of elevations in intracellular calcium in murine macrophages and dendritic cells. *J Neuroimmunol* 2000;**102**:163–71
 109. Ferré S, Mazur A, Maier JAM. Low-magnesium induces senescent features in cultured human endothelial cells. *Magnes Res* 2007;**20**:66–71
 110. Uchida T, Itoh H, Nakamura Y, Kobayashi Y, Hirai K, Suzuki K, Sugihara K, Kanayama N, Hiramatsu M. Zinc and magnesium ions synergistically inhibit superoxide generation by cultured human neutrophils—a promising candidate formulation for amnioinfusion fluid. *J Reprod Immunol* 2010;**85**:209–13
 111. Uchi H, Terao H, Koga T, Furue M. Cytokines and chemokines in the epidermis. *J Dermatol Sci* 2000;**24**:S29–38
 112. Petraut I, Zimowska W, Mathieu J, Bayle D, Rock E, Favier A, Rayssiguier Y, Mazur A. Changes in gene expression in rat thymocytes identified by cDNA array support the occurrence of oxidative stress in early magnesium deficiency. *Biochim Biophys Acta* 2002;**1586**:92–8
 113. Libby P. Inflammatory mechanisms: the molecular basis of inflammation and disease. *Nutr Rev* 2007;**65**:140–6
 114. Saris NE, Mervaala E, Karppanen H, Khawaja JA, Lewenstam A, Magnesium. An update on physiological, clinical and analytical aspects. *Clin Chim Acta* 2000;**294**:1–26
 115. Altura BM, Altura BT. Cardiovascular risk factors and magnesium: relationships to atherosclerosis, ischemic heart disease and hypertension. *Magnes Trace Elem* 2011;**10**:182–92
 116. Zandsteeg AMG, Hirmann P, Pasma HR, Yska J-P, ten Brinke A. Effect of MgSO₄ on FEV₁ in stable severe asthma patients with chronic air-flow limitation. *Magnes Res* 2009;**22**:256–61
 117. Bochner BS. Immunological aspects of allergic asthma. *Annu Rev Immunol* 1994;**12**:295–335
 118. Yamamoto Y, Gaynor RB. Therapeutic potential of inhibition of the NF- κ B pathway in the treatment of inflammation and cancer. *J Clin Invest* 2001;**107**:135–42
 119. Dunnett J, Nayler WG. Calcium efflux from cardiac sarcoplasmic reticulum: effects of calcium and magnesium. *J Mol Cell Cardiol* 1978;**10**:487–98
 120. D'Angelo EK, Singer HA, Rembold CM. Magnesium relaxes arterial smooth muscle by decreasing intracellular Ca²⁺ without changing intracellular Mg²⁺. *J Clin Invest* 1992;**89**:1988–94
 121. Feldmann M, Maini SRN. Role of cytokines in rheumatoid arthritis: an education in pathophysiology and therapeutics. *Immunol Rev* 2008;**223**:7–19
 122. Altura BM, Gebrewold A, Zhang A, Altura BT. Low extracellular magnesium ions induce lipid peroxidation and activation of nuclear factor- κ B in canine cerebral vascular smooth muscle: possible relation to traumatic brain injury and strokes. *Neurosci Lett* 2003;**341**:189–92
 123. Ma J, Folsom AR, Melnick SL, Eckfeldt JH, Sharrett AR, Nabulsi AA, Hutchinson RG, Metcalf PA. Associations of serum and dietary magnesium with cardiovascular disease, hypertension, diabetes, insulin, and carotid arterial wall thickness: the ARIC study. Atherosclerosis Risk in Communities Study. *J Clin Epidemiol* 1995;**48**:927–40
 124. Hellerstein EE, Vitale JJ, White PL, Hegsted DM, Zamcheck N, Nakamura M. Influence of dietary magnesium on cardiac and renal lesions of young rats fed an atherogenic diet. *J Exp Med* 1957;**106**:767–76
 125. Whelton PK, Klag MJ. Magnesium and blood pressure: review of the epidemiologic and clinical trial experience. *Am J Cardiol* 1989;**63**:26G–30G
 126. Woods KL. Possible pharmacological actions of magnesium in acute myocardial infarction. *Br J Clin Pharmacol* 1991;**32**:3–10
 127. Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelenboom P, Emmerling M, Fiebich BL, Finch CE, Frautschy S, Griffin WS, Hampel H, Hull M, Landreth G, Lue L, Mrak R, Mackenzie IR, McGeer PL, O'Banion MK, Pachter J, Pasinetti G, Plata-Salaman C, Rogers J, Rydel R, Shen Y, Streit W, Strohmeyer R, Tooyoma I, Van Muiswinkel FL, Veerhuis R, Walker D, Webster S, Wegrzyniak B, Wenk G, Wyss-Coray T. Inflammation and Alzheimer's disease. *Neurobiol Aging* 2000;**21**:383–421
 128. McGeer PL, McGeer EG. Inflammation and the degenerative diseases of aging. *Ann N Y Acad Sci* 2004;**1035**:104–16
 129. Ralay Ranaivo H, Craft JM, Hu W, Guo L, Wing LK, Van Eldik LJ, Watterson DM. Glia as a therapeutic target: selective suppression of human amyloid-beta-induced upregulation of brain proinflammatory cytokine production attenuates neurodegeneration. *J Neurosci* 2006;**26**:662–70
 130. Giulian D. Microglia and the immune pathology of Alzheimer disease. *Am J Hum Genet* 1999;**65**:13–8
 131. Craft JM, Watterson DM, Van Eldik LJ. Neuroinflammation: a potential therapeutic target. *Expert Opin Ther Targets* 2005;**9**:887–900
 132. Marinov MB, Harbaugh KS, Hoopes PJ, Pikus HJ, Harbaugh RE. Neuroprotective effects of preischemia intraarterial magnesium sulfate in reversible focal cerebral ischemia. *J Neurosurg* 1996;**85**:117–24
 133. McDonald JW, Silverstein FS, Johnston MV. Magnesium reduces (NMDA)-mediated brain injury in perinatal rats. *Neurosci Lett* 1990;**109**:234–8
 134. Weglicki WB, Phillips TM, Mak IT, Cassidy MM, Dickens BF, Stafford R, Kramer JH. Cytokines, neuropeptides, and reperfusion injury during magnesium deficiency. *Ann N Y Acad Sci* 1994;**723**:246–57
 135. Klegeris A, Bissonnette CJ, McGeer PL. Modulation of human microglia and THP-1 cell toxicity by cytokines endogenous to the nervous system. *Neurobiol Aging* 2005;**26**:673–82
 136. Pincelli C, Sevigani C, Manfredini R, Grande A, Fantini F, Bracci-Laudiero L, Aloe L, Ferrari S, Cossarizza A, Giannetti A. Expression and function of nerve growth factor and nerve growth factor receptor on cultured keratinocytes. *J Invest Dermatol* 1994;**103**:13–8
 137. Dou Y-C, Hagströmer L, Emtestam L, Johansson O. Increased nerve growth factor and its receptors in atopic dermatitis: an immunohistochemical study. *Arch Dermatol Res* 2006;**298**:31–7
 138. Wolfgang A Nockher, Sanchaita Sonar, Harald Renz. Neurotrophins. *Allergy and allergic diseases*. New York, NY: Wiley-Blackwell, 2008:507
 139. Tominaga M, Takamori K. An update on peripheral mechanisms and treatments of itch. *Biol Pharm Bull* 2013;**36**:1241–7
 140. Kruger RP, Aurandt J, Guan K-L. Semaphorins command cells to move. *Nat Rev Mol Cell Biol* 2005;**6**:789–800
 141. Neufeld G, Kessler O. The semaphorins: versatile regulators of tumour progression and tumour angiogenesis. *Nat Rev Cancer* 2008;**8**:632–45

142. Ikezawa Z, Komori J, Ikezawa Y, Inoue Y, Kirino M, Katsuyama M, Aihara M. A role of *staphyococcus aureus*, interleukin-18, nerve growth factor and semaphorin 3A, an axon guidance molecule, in pathogenesis and treatment of atopic dermatitis. *Allergy Asthma Immunol Res* 2010;**2**:235–46
143. Nikolett N, Katalin F, Sarolta B, Istvan B, Zsuzsanna BC, Lajos K, Marta S. NRP1 activates NF-κB signaling pathway and initiates proliferation in keratinocytes. *Int J Genomic Med* 2013;**1**:102
144. Fukamachi S, Bito T, Shiraishi N, Kobayashi M, Kabashima K, Nakamura M, Tokura Y. Modulation of semaphorin 3A expression by calcium concentration and histamine in human keratinocytes and fibroblasts. *J Dermatol Sci* 2011;**61**:118–23
145. Malaviya R, Uckun FM. Role of STAT6 in IgE receptor/FcεRI-mediated late phase allergic responses of mast cells. *J Immunol* 2002;**168**:421–6
146. Ichiro Katayama, Hiroyuki Murota, Ken Igawa, Takahiro Satoh, Kiyoshi Nishioka, Hiroo Yokozeki A. Targeting STAT6 in atopic eczema/dermatitis. In: Ruby Pawankar, Stephen T. Holgate, Lanny J. Rosenwasser (ed.) *Allergy Frontiers: Future Perspectives*. Tokyo: Springer, 2010, pp.167–78
147. Song Y, Li TY, van Dam RM, Manson JE, Hu FB. Magnesium intake and plasma concentrations of markers of systemic inflammation and endothelial dysfunction in women. *Am J Clin Nutr* 2007;**85**:1068–74
148. Sara AC, Yiqing S, Lauren N, Lesley T, Ian H dB, Fran T, Robert W, Simin L. Relations of dietary magnesium intake to biomarkers of inflammation and endothelial dysfunction in an ethnically diverse cohort of postmenopausal women. *Diab Care* 2010;**33**:304–10
149. Rayssiguier Y, Gueux E, Nowacki W, Rock E, Mazur A. High fructose consumption combined with low dietary magnesium intake may increase the incidence of the metabolic syndrome by inducing inflammation. *Magnes Res* 2006;**19**:237–43
150. Brown DM, Donaldson K, Borm PJ, Schins RP, Dehnhardt M, Gilmour P, Jimenez LA, Stone V. Calcium and ROS-mediated activation of transcription factors and TNF-α cytokine gene expression in macrophages exposed to ultrafine particles. *Am J Physiol Lung Cell Mol Physiol* 2004;**286**:L344–53
151. Kelepouris E, Agus ZS. Hypomagnesemia: renal magnesium handling. *Semin Nephrol* 1998;**18**:58–73

(Received February 9, 2014, Accepted April 14, 2014)