Mini Review

Trends in polymeric electrospun fibers and their use as oral biomaterials

Agnes B Meireles¹, Daniella K Corrêa², João VW da Silveira³, Ana LG Millás⁴, Edison Bittencourt⁴, Gustavo EA de Brito-Melo¹ and Libardo A González-Torres³

¹Pharmacy Department, Laboratory of Immunology, UFVJM and PPGCF-UFVJM, Diamantina, MG 39100-000, Brazil; ²Institute of Science and Technology – UFVJM, Diamantina, MG 39100-000, Brazil; ³Institute of Science and Technology – UFVJM, Diamantina, MG 39100-000, Brazil; ⁴Chemical Engineering Department, UNICAMP, Campinas, SP 13083-852, Brazil Corresponding author: Libardo A González-Torres. Email: I.gonzales@ict.ufvjm.edu.br

Impact statement

Nanotechnology is a challenge for many researchers that look for obtaining different materials behaviors by modifying characteristics at a very low scale. Thus, the production of nanostructured materials represents a very important field in bioengineering, in which the electrospinning technique appears as a suitable alternative. This review discusses and provides further explanation on this versatile technique to produce novel polymeric biomaterials for oral applications. The use of electrospun fibers is incipient in oral areas. mainly because of the unfamiliarity with the technique. Provided disclosure, possibilities and state of the art are aimed at supporting interested researchers to better choose proper materials, understand, and design new experiments. This work seeks to encourage many other researchers-Dentists, Biologists, Engineers, Pharmacists-to develop innovative materials from different polymers. We highlight synthetic and natural polymers as trends in treatments to motivate an advance in the worldwide discussion and exploration of this interdisciplinary field.

Abstract

Electrospinning is one of the techniques to produce structured polymeric fibers in the micro or nano scale and to generate novel materials for biomedical proposes. Electrospinning versatility provides fibers that could support different surgical and rehabilitation treatments. However, its diversity in equipment assembly, polymeric materials, and functional molecules to be incorporated in fibers result in profusion of recent biomaterials that are not fully explored, even though the recognized relevance of the technique. The present article describes the main electrospun polymeric materials used in oral applications, and the main aspects and parameters of the technique. Natural and synthetic polymers, blends, and composites were identified from the available literature and recent developments. Main applications of electrospun fibers were focused on drug delivery systems, tissue regeneration, and material reinforcement or modification, although studies require further investigation in order to enable direct use in human. Current and potential usages as biomaterials for oral applications must motivate the development in the use of electrospinning as an efficient method to produce highly innovative biomaterials, over the next few years.

Keywords: Nano and microfibers, biopolymers, nanotechnology, dental treatments, dentistry

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Introduction

The development of new materials and oral treatments led to shorter healing periods and rehabilitation procedures with minor pain. Furthermore, materials to be used in oral conditions are currently challenging concerning durability, resistance, degradability, and biocompatibility. Thus, researches that support studies on production of novel materials are very important to improve current treatments and to provide more efficient procedures.

The physical and mechanical properties of structured biomaterials in micro or nano scale could generate superior structures with potential for therapeutic use. Electrospinning is one

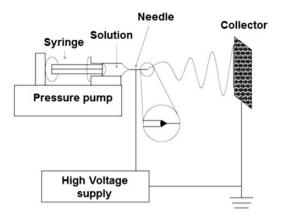


Figure 1. Schematic representation of a typical electrospinning apparatus. A polymeric solution is ejected from a needle controlled by a syringe pump. The needle is connected to a high voltage DC supply that provides charge into the polymer solution. A fiber jet is produced by the solvent evaporation and then deposited on the collector.

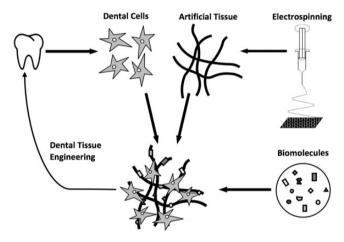


Figure 2. Dental cells regeneration using polymeric electrospun fibers. The fibers can be loaded with different growth factors that can improve cell proliferation, or with the presence of antimicrobial agents to inhibit bacterial contamination.

of the technologies for producing such structured materials. It has been reported as an efficient technique for polymeric fibers fabrication with diameter range from 3 to 5000 nm, 1,2 from which several polymers have been successfully electrospun into micro and nanofibers in recent years.³ Electrospinning devices are generally formed by a syringe, a needle, a source of high voltage, a collector field, and a precision controlled pump. The technique consists in creating micro or nanoscale fibers by high voltage application to a syringe loaded with a polymer solution. The fibers are deposited on a metallic collector charged with a voltage, as schematized in Figure 1.4 The process of electrospinning is relatively simple for implementation and presents low cost when compared to other fiber fabrication processes, such as melt fibrillation,⁵ island-in-sea,⁶ gas jet, ⁷ nanolithography, ⁸ and self-assembly. ⁹⁻¹² Fibers assemblies from synthetic and natural polymers could be achieved and applications range from sensors, filtration, protective material, electrical or optical areas to biomedical field.¹³ Tissue engineering (TE), ^{14,15} drug delivery, ¹⁶ wound dressing, and nanofiber reinforced composites 17,18 are potential uses in

dentistry. Figure 2 shows a simplified representation how electrospun fibers are employed in dental TE.

This review summarizes most frequently used materials in biomedical areas, including natural and artificial polymers exploring possibilities or recent use in researches on oral biomaterials. It is also shortly described the electrospinning process, including important production parameters and introducing recent areas of development.

Electrospinning process and parameters

The fibers morphology (Figure 3) and the process of the electrospinning are affected by different factors: the molecular weight of the polymer, source of voltage, concentration of the polymer solution, needle gauge type, and the distance between the needle and the pitch collector.¹⁹ Spinnability, fiber diameters, fiber uniformity, fiber alignment, defects control (e.g. beads, junctions, and pores), and other properties are tunable by adjusting these factors and also including substrate-related parameters (polymer concentration, viscosity, molecular weight, surface tension) and apparatus-related parameters (flow rate and electric field). All these parameters are important to obtain flawless fibers and the choice of a polymer and its production must consider these factors and their modulation.

Spinning voltage is related to some defects as bead formation in fibers (Figure 3(a)).²⁰ However, a low voltage can increase the time of solvent evaporation and high voltages cause the rapid evaporation of solvent, producing a fiber with less pores and defects. ¹⁷ Spinning voltage influences also the shape of electrospun fibers in a macroscopical level and in the homogeneity of the generated structures (Figure 3(b) and (c)).17

The distance between the needle and the collector field should be carefully adjusted. The fiber morphology is influenced by the distance, which in turn influences in deposition time and evaporation rate of the solvent.²¹ The fiber may break before reaching the collector field and create thick fibers with greater distances between them; consequently, the solvent cannot fully evaporate resulting in inaccurate fiber.²² The diameter of the needle directly affects the diameter of the fiber, and when it is reduced, pore formation decreases.

Environmental conditions such as: humidity, atmospheric pressure, and temperature differently influence on all process parameters. The humidity directly affects the polymer solution resulting in changes in the morphology, like small holes, increased roughness, etc. The effects of relative humidity on electrospun fiber morphology are dependent on polymer hydrophobicity, solvent miscibility with water, and solvent volatility.²³Too low atmospheric pressure does not allow the process to occur. On the other hand, when working temperature increases, surface tension and viscosity of the fluid decrease and solvent evaporation could be accelerated, terminating the process prematurely.²⁴ In other cases, when the environment is very cold, the spinning process is hindered.

The viscosity and conductivity of the polymer solution are the main characteristics that influence electrospinning process. The viscosity is related to the polymer molecular weight and the amount of dissolved solids in the solution. Solution

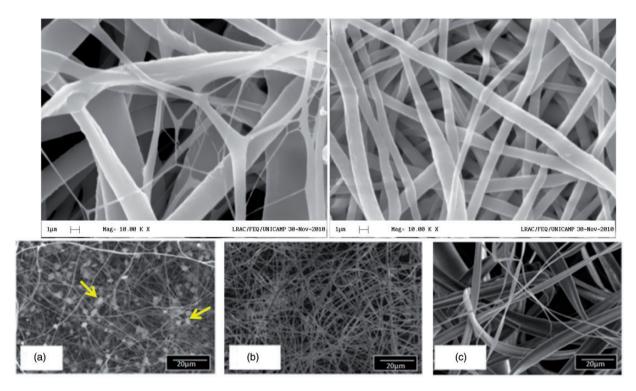


Figure 3. SEM images from electrospun PCL fibers. In (a) is presented beads formation (arrows). In (b) and (c) are presented nanofibers with regular aspect. The upper left image presents a wider diameter size distribution compared with the image at the upper right, due to the lack of control of the process.

concentration is reported to strongly affect fiber size, with fiber diameter increasing with the solution concentration in a power law relationship.²⁰ In addition, electrospinning from solutions of high concentration is reported to produce fiber sizes with bimodal distribution, reminiscent of distributions observed in the similar droplet generation process of electrospraying. High viscosity can difficult the ejection of the solution from the needle and can also dry before it reaches the metallic collector. The conductivity of the polymer solution influences the diameter of the fiber with direct relation: as the conductivity increases, it produces denser fibers.¹⁷

Polymers used in oral applications

Examples and most used polymers are presented in Table 1, indicating the main oral applications and some common composites.

The poly(ε-caprolactone) (PCL), poly(lactic acid) (PLA), poly(L-lactide) (PLLA), and poly(lactic-co-glycolic acid) (PLGA) are polymers commonly used in biomedical and dental applications. Natural polymers such as chitosan, collagen, and cellulose were cited and are also currently used. ^{27,45,52,53,56,60} Other polymers such as poly(ethylene oxide) (PEO), folyological polydioxanone (PDS), folyological polydioxanone (PDS), folyological polydioxanone (PU), fo

PCL

PLC is a biodegradable thermoplastic polymer that present relatively low cost. Its physicochemical properties enable it for applications in the nanometric scale processing and prototyping. 45

It has been widely used in drug delivery systems for longterm dental implants area. PCL was used in association with alginate for drug release during dental treatments to prevent bacterial accumulation at the site of implantation.²² PCL has been also used in drug delivery for creation of membranes incorporating nano-hydroxyapatite and amoxicillin,²⁷ obtaining good biocompatibility and reducing bacterial contamination of periodontal defects. Further applications were described for TE. PCL scaffolds mineralized with apatite improve cell proliferation in dental pulp. PCL when combined with bioactive glass gradually improves cell proliferation and the mechanical properties of scaffolds obtained by materials blending.³⁴ In periodontal regeneration, PCL has been effectively used in combination with gelatin, with engineered nanotopology of structures and mechanical stimulation.³⁰

PLA

PLA is a versatile polymer that can be adapted to different morphologies. ⁴⁰ It is degradable in the human tissues; however, its degradation produces acids that difficult the regeneration process requiring blending with other materials. ³⁹

PLA is used mostly for controlled drug delivery and tissue regeneration. To reduce tissue damage caused for delivered acids, PLA nanofibers were loaded with beta-tricalcium phosphate (β -TCP) which proved to be a viable option in PCL membranes.³⁹ PLA nanofibers associated with metronidazole (MNZ) were used to control microbiological proliferation during periodontitis treatment, inhibiting bacteria growth during the treatment.⁴⁰ Ampicilin and MNZ were incorporated in PLA matrix for drug delivery,

Table 1. Relation of electrospun polymers for oral applications.

Electrospun polymer	Application	Year	References
PCL	Drug delivery	2018	25
PCL	Tissue regeneration	2017	26
PCL	Drug delivery	2016	27
PCL	Tissue regeneration	2016	28
PCL	Tissue regeneration	2016	29
PCL-GEL	Tissue regeneration	2016	30
PCL, PCL-GEL	Wound dressing	2015	31
PCL	Tissue regeneration	2015	32
PCL	Modification of materials	2014	33
PCL	Drug delivery	2013	22
PCL	Tissue regeneration	2013	34
PCL	Drug delivery	2013	35
PLA, PCL	Tissue regeneration	2018	36
PLA	Drug delivery	2016	37
PLA, PCL-GEL	Tissue regeneration	2016	38
PLA	Tissue regeneration	2013	39
PLA	Drug delivery	2012	40
PLA	Tissue regeneration	2011	41
PLLA	Drug delivery	2017	42
PLLA, PCL	Tissue regeneration	2016	43
PLLA, PCL	Tissue regeneration	2014	44
PLLA, PLGA, PGA, PCL, Chitosan, PLA	Tissue regeneration	2011	45
PLLA, PLGA	Tissue regeneration	2011	4
PLLA	Modification of materials	2008	46
PLGA, PCL	Tissue regeneration	2017	47
PDS	Drug delivery	2018	48
PDS	Drug delivery	2016	49
PDS	Tissue regeneration	2015	50
PDS	Drug delivery	2013	51
Chitosan	Tissue regeneration	2015	52
Chitosan	Membrane dressing	2015	53
Nylon-6/chitosan	Modification of materials	2015	54
Nylon-6	Modification of materials	2015	55
Collagen	Tissue regeneration	2012	56
PLGA	Tissue regeneration	2016	57
PVA	Modification of materials	2016	58
PU + GEL	Wound dressing	2016	59
PAN	Modification of materials	2015	60

PAN: poliacrylonitrile; PCL: poly(e-caprolactone); PLLA: poly(L-lactide); PDS: polydioxanone; PLGA: poly(lactic-co-glycolic acid); PVA: polyvinyl alcohol.

finding potential applications in periodontal and endodontic infections.³⁷

PLA in combination with PCL and gelatine was produced incorporating tetracycline exhibiting good characteristics to be used as antibacterial dental implant coating.³⁸ PLA nanofibers together with poly(DL-lactide-co-e-caprolactone), nano-hydroxyapatite, n-HAp, have been also produced.⁴¹ Resulting nanofibers formed layers that improved mechanical and biological properties resulting in the raise of periodontal membrane regeneration.

PLLA

PLLA is a biodegradable and easily applicable polymer, with a porous morphology when polymerized with other polymers. PLLA has good physical and mechanical properties and it is commonly used in association with other materials. 45

PLLA presents advantages for oral applications, because of its degradation rate and its potential for cell adhesion and proliferation. The disadvantages observed are the possibility to release of acids during degradation and its high

hydrophobicity. 4,45 These aspects can be reduced with the combination of PLLA with other materials. A recent study shows that HA-loaded PLLA/PCL can lead to osteoblast/ odontoblast differentiation. 43 Scaffolds of PLLA have been also point out as an excellent environment for proliferation and adhesion of cells of different types.⁶² The mechanical properties such as compressive strength, diametral tensile strength, flexural strength, and linear shrinkage of PLLA fibers have been improved by mixing other materials as bisphenyl glycidylmethacrylate (Bis-GMA) and triethylenglycoldimethacrylate (TEGDMA).41

Polyethylene oxide and polyvinyl alcohol

PEO is described as a relatively high molecular weight, biocompatible material that effectively improves the mechanical and biological properties of composite hydrogels. PEO has presented a significant impact on the mechanical and rheological properties associated with a glucose-sensitive antibacterial chitosan.⁶¹ It has been verified that a chitosan fraction contributed to an antimicrobial effect, decreased tendency to

spherulitic crystallization of PEO, and enhanced puncture and tensile strength. PEO is used as an additive in tooth paste because it reduces biofilm formation.⁶³ Inhibition of bacterial adhesion on enamel was observed suggesting that the PEO could be utilized in caries prevention.⁶⁴

Polyvinylalcohol (PVA) presents biocompatibility, being prepared easily and having non-toxic solvent. It is a water-soluble synthetic polymer and the back bone chains are highly interconnected by hydrogen bonding due the presence of abundant hydroxyl groups. These characteristics contribute to chemical resistance and mechanical properties, including high tensile and impact strength. The hydroxyl groups can be used to incorporate molecules of biological origin, such as collagens, hyaluron and deoxyribonucleic acid. PVA is biodegradable, decomposing non toxically into water and carbon dioxide. Electrospinning is one of the micro and nano-size production methods for PVA for the reinforcement of dental materials.

Poly DL-lactide-co-glycolide and PDS

Poly DL-lactide-co-glycolic (PLGA) is a biodegradable and biocompatible polymer that closely resembles natural proteins that may be metabolically hydrolyzed to monomers of lactic acid and glycolic acid. It is used for reinforcement of other materials without changing characteristics such as cell adhesion. Furthermore, the PLGA can be used in the controlled release of drugs during bone growth^{66,67} and in membranes that simultaneously release drugs, support bone growth, and possess barrier functions.⁵⁷

The PDS is a polymer frequently used as suture material. In recent studies, investigation for tissue regeneration and drug delivery system was performed. Bioactive scaffolds were produced using PDS for regenerative endodontics and their results demonstrated efficiency of the material for adhesion and proliferation of fibroblasts exhibiting good biocompatibility. It also evidenced its potential for encapsulation of distinct bioactive molecules.⁶⁸

PVDF and nylon

Polyvinylidene fluoride (PVDF) is a biocompatible, flexible material with high mechanical strength. It has been electrospun with drugs to form membranes, exhibiting potential for antibiotics release in wound healing. In addition, membranes produced with PVDF presented good anti-bacterial properties.⁶⁹

Nylon-6 (polyamide) is also a biocompatible material with excellent mechanical, chemical, and thermal properties. Nylon composites have been also used for reinforcement of dental materials. Nylon has been produced by electrospinning in dental resins with aligned or random oriented fibers. Best mechanical properties were observed for aligned fibers. Silica-nylon reinforcement reported to increase the flexural and fracture strength of interim partial fixed dental protheses.

Natural polymers

Natural polymers are adequate as biomaterials because they are structurally similar to biological tissues. They sometimes

possess similar or identical monomers, as those found in organic matrices, reducing the problems associated to toxicity or inflammatory reactions.⁷¹ Nevertheless, there exist some disadvantages as the possible rejection of the material by the organism, strong physiological induced activity, and low mechanical properties.⁷² Some of the main natural polymers with possible oral applications are presented: chitosan, collagen, and cellulose.

Chitosan

Chitosan is extracted from the crustacean shells and is the second largest natural polysaccharide after cellulose.⁷³ Chitosan possesses good properties, like biodegradability, biocompatibility, it has fungicidal, antimicrobial, and antitumor activity and also has strong healing properties, as it offers an excellent environment for cell adhesion and proliferation. Applications are focus of a range of studies reporting its medical potential as nerve regeneration, bone, cartilage, epidermal healing, gene therapies, drug delivery, semipermeable membranes, and development of biomaterials.⁷⁴ Another major advantage of chitosan is that its production process is economically and environmentally viable since it comes from by-products of the fishing activity of parts that were discarded in addition to being a renewable source of raw material.⁷⁵ In oral medicine, chitosan has been produced as membrane for bone regeneration⁵² and caries prevention.⁵³

Collagen

Collagen is the main organic component of the dentin matrix and represents an alternative to produce scaffolds for dental regeneration. Collagen-hydroxyapatite composite is used due to the enhanced biological and mechanical characteristics obtained by the binary system. Some successful applications on dentin recovery, revascularization in open apex periodontitis treatment, and root caries inhibition were reported. Collagen-based nanocomposites using bioactive glass are related to indice cell proliferation and osteogenic differentiation in dental pulp.

Cellulose

Cellulose is the most abundant high molecular weight polysaccharide in nature, and for this reason with relatively low cost. Cellulose is widely used for a large number of applications in the chemical, food, and pharmaceutical industry. The form and order of the crystallite give interesting properties such as tensile strength, rigidity, heat resistance, and also elastic properties which are very important materials aspects for biomedical purposes including reinforce of materials and because of its good biocompatibility, cellulose-based membrane has been recently proposed for medical applications.^{3,79} It has been reported that electrospun cellulose with nanohydroxyapatite results in good mechanical properties and biocompatibility with dental cells with possible applications in TE.80 Cellulose derivatives such as methylcellulose and cellulose acetate are alternatives to biomedical applications when cellulose dissolution is complicated, for instance, to generate cellulose acetate membrane for dentistry.81

Oral applications

Next, main applications of fibers produced by electrospinning in oral areas: drug delivery systems, tissue regeneration and modification of materials, are presented. Functional fibers could be achieved by blending polymers mixtures and active molecules or by dispersing nano or microfillers within fibrous matrix. These composites are attractive since novel materials fabricated with electrospinning demonstrated superior mechanical properties, bioactivity and biochemical properties compared to their constituent phases.

Drug delivery systems

The aim of drug delivery systems is to enable the controlled drug releasing towards alleviating medical conditions at a defined rate over a definite period. The promising use of nanofibers in drug delivery systems might result in salient features such as high loading capacity. Current release of diverse drugs ranging from antibiotics and anticancer agents to proteins, aptamers, DNA has been successfully achieved with electrospun fibers. Different drug delivery assemblies are presented in Figure 4.

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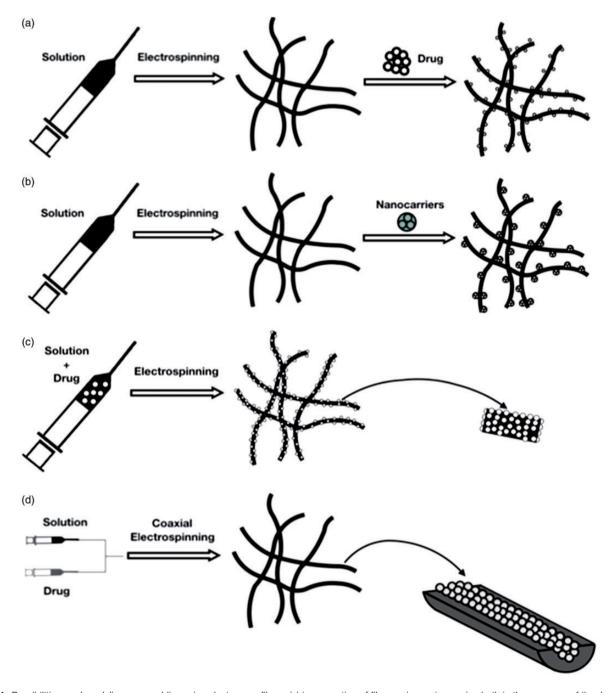


Figure 4. Possibilities on drug delivery assemblies using electrospun fibers: (a) impregnation of fibers using an immersion bath in the presence of the drug; (b) nanocarriers attachment to the fiber; (c) polymeric fiber matrix loaded with the drug, produced by electrospinning from a solution containing the drug; (d) coaxial electrospun fiber obtained by the encapsulation of the drug (core) inside a polymeric structure (shell).

Pharmaceutical properties as bioavailability, solubility, and stability could be enhanced with this strategy.⁸² Electrospun fibers have shown their advantages in the field of drug delivery due to their high surface area and volume ratio with interconnected pores in the fibers.

Strategies to avoid bacterial and fungal diseases are important in the oral-facial field. 40 For this reason, scaffolds containing antibiotics have been increasingly used in oral studies^{22,27,37,40,49,51} and mucoadhesion strategies are also reported. 83,84 The presence of anti-inflammatory agents in such composites could activate a signaling cascade and trigger the healing process.82

A combination of ampicillin (AMP), MNZ (20/20%w/ w), and PLA single fiber mats were described to suppress periodontopathogenic species A. actinomycetemcomitans within an elution time of 24 h³⁷ and the effect of MNZ (0.1-40% w/w) with electrospun PLA fibers decreased viability of F. nucleatum and P. gingivalis up to 28 days and for A. actinomycetemcomitans up to 2 days. 40 In Reise et al., 40 it is described that highly loaded fiber mat with 40% of MNZ (w/w) with a weight of 35 mg contains 14 mg of MNZ and the amount of MNZ when administered systemically adds up to $8400 \,\mathrm{mg}$ (3 × $400 \,\mathrm{mg}$ daily over seven days). However, toxic effects are expected if local drug administration provides highly concentrated antibiotics doses. Gradually drug release could be a more biologically friendly approach to be tested such as amounts that promote the elimination bacteria associated with the reduction adverse effects.44

Regenerative endodontics requires effective root canal disinfection with no or minimal harm to stem cells and growth factor. Triple antibiotic (ciprofloxacin, MNZ, and minocycline) or double highly concentrated antibiotic (minocycline-free) pastes are typically accomplished for root canal disinfection.⁵¹ However, potentially toxic effects of highly concentrated antibiotic pastes on dental pulp stem cells and dental pulp fibroblasts should be concerned. Controlled amounts of antibiotics could be achieved with electrospun antibiotic-containing scaffolds. For purpose, PDS fibers were fabricated with MNZ, minocycline, and ciprofloxacin via electrospinning and tested in vitro against a dual-species biofilm (A. naeslundii and Enterococcus faecalis, bacteria prevalent in immature infected root canals and responsible for secondary infections in necrotic teeth after treatment, respectively) with a significant bacterial death, whereas they did not affect dental pulp stem cells attachment and proliferation on dentin.44

Controlled localized delivery of antibacterial agents to dental implants site using a biodegradable electrospun material could be designed in order to prevent bacterial infection, one of the causes of dental implant fail. PCL/ alginate associated with MNZ rings were custom designed and inserted around dental implant prior to their placement procedure. This strategy minimized burst release effects with MNZ release over 30 days.²²

Reported studies suggest the use of these materials for periodontal and endodontical infections and for dental implants. Drug release from nanofibers could be due to desorption of drug from the surface, diffusion from pores and/or matrix degradation, all processes likely to get affected by polymer type, porosity, morphology, and also by geometry of nanofibers.⁸⁵ Other antibiotics have been used in drug delivery inside electrospun materials: amoxiciline, ²⁷ AMP, ³⁷ minocycline, and ciprofloxacin. ^{49,51}

Antifungal and mucoadhesives properties are also reported in this area. Poly-ε-caprolactone nanofibers functionalized with ketoconazole were produced by electrospinning and tested against filamentous fungi in Veras et al. 86 and the functionalized nanofibers showed antifungal activity against Aspergillus avus, A. carbonarius, A. niger, Aspergillus sp. A29, Fusarium oxysporum and Penicillium citrinum by agar diffusion test with inhibitory zones ranging from 6 to 44 mm.

Mucoadhesivity of polymers is sometimes a desired characteristic in drug delivery systems because it could enhance drug retention time at the application and increase bioavailability.83 The chitosanethylenediaminetetraacetic acid/polyvinyl (CS-EDTA/PVA) was reported as mucoadhesive and electrospinnable composite that associated with clotrimazole (CZ)-loaded microemulsion could be used for antifungal purposes,84 demonstrating sustained release with antifungal activity in vitro.

Tissue regeneration

The synthesis of unique materials to support the regeneration of tissues and lost organs due to trauma and/or diseases and TE remains pivotal to the development and translational impact in regenerative dentistry. 44 TE uses three basic components (cells, scaffolds and biomolecules) to develop biofunctional substitutes for restore and maintenance of tissue function. The regeneration process can be assisted by scaffolds produced by electrospinning method resulting in biocompatible and sometimes biodegradable polymers. 30,87 However, natural tissue integration with functional neovessels can be also obtained (Figure 5). Scaffolds can convey growth factors as well as cells to the target site to assist the injury regeneration⁸⁸ and generate a support to the increase of cell adhesion, growth, migration rate, and differentiation. Biodegradable polymers withdraw surgical implant removal and can improve patient recovery process. 51 Thus, nanostructured materials have been electrospun as mono and multilayered membranes and scaffolds in order to improve tissue regeneration.

Regenerative endodontics includes approaches to treat pulpal and periapical pathologies, for instance, in procedures that could be used for apical closure induction or for teeth that were previously endodontically treated.⁸³ In this area, nanofibrous scaffolds have been designed to stimulate positive cell-ECM interactions, increase cell proliferation, maintain cell phenotype, support differentiation of stem cells, and activate cell-signaling pathways by providing physical and chemical stimuli. 44 In Bae et al., 56 collagen was added into electrospun bioactive glass to improve its poor mechanical properties as high brittleness. Proliferation and odontogenic differentiation of dental pulp cells was tested with enhanced cellular growth and differentiation into odontoblasts and human dental pulp

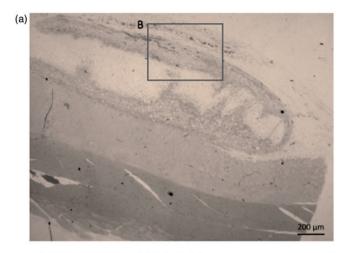




Figure 5. In vivo assay via intramuscular scaffolds implantation in rat. Histological aspect 28 days after implantation of PLGA scaffolds, showing (a) the natural tissue integration capacity and (b) the neovascularization (dark regions) in the implanted region.

cells grew preferentially on the nanocomposite, rather than on type I collagen during culture for 14 days. Additionally, gene expression of the key integrin α2β1 which specifically binds to collagen molecular sequence was highly stimulated in the collagen/electrospun bioglass composite, suggesting that an integrin-mediated process was associated to the odontogenic stimulation.

In Kim et al.,³⁴ a polycaprolactone nanofibrous scaffold was produced by electrospinning and the surface was mineralized with apatite to test cell adhesion, growth, and odontoblastic differentiation. Their results indicated that such scaffold promoted growth and odontogenic differentiation of human dental pulp cells through the integrinmediated signaling pathway.

Healing of periodontal tissues (ligament and surrounding bone) could be improved by TE and nanofibers. 45 In Sundaramy et al., 28 PCL multiscale (micro/nano) membrane was obtained by simultaneous spinning of PCL micro and nanofibers to recover a calcium sulfate/chitosan scaffold to assist in both ligament and bone regeneration. Osteoblastic differentiation of human dental follicle stem cells (hDFCs) on calcium sulfate/chitosan scaffold showed maximum alkaline phosphatase at seventh day followed by

a decline thereafter when compared to chitosan control scaffold and also fibroblastic differentiation of hDFCs.

In Bottino *et al.*, ⁵¹ chitosan for guided bone regeneration membranes was electrospun using chitosan (70% deacetylated, 312 kDa, 5.5 w/v%), with or without the addition of a crosslinking agent (genipin) in order to extend biodegradation. Genipin-crosslinked mats presented lower degradation rates based on mass loss compared with uncrosslinked mats. In addition, genipin-crosslinked mats supported the proliferation of SAOS-2 cells in a five-day growth study, similar to uncrosslinked mats.

In summary, electrospun of polymeric nanofibers has been used for regeneration of pulp-dentin complex used in combination with biomechanical and biochemical cues in endodontics. 44,56 Nanofibers have been used also for regeneration of periodontal ligament alone, 45 considering alveolar regeneration at the same time²⁸ or combining oriented nanotopological and mechanical stresses.34,51 Multilayer membrane has been produced for bone and soft tissue regeneration at the same time, 29 and barrier membranes in guided bone regeneration.⁵² Testing cell viability with increase or decrease of the content of nanofibers while maintaining the required structural and biological properties could be challenging. Although initial in vitro results are promising, limited use should be taken until clinical further long-term data are obtained.83

Modification of materials

Ceramic, polymeric, and biodegradable materials have been increasingly used in dentistry as aesthetic options for oral rehabilitation. The improvement of mechanical and physical properties of dental composite such as tensile strength, flexibility and elasticity at nanoscale has stimulated studies to investigate methods to produce modified restorative materials⁴⁶ to better support the challenging biological and mechanical oral conditions. The use of electrospinning method for these purposes has been indicated. 33,46,55,58,70,89 Nanofiber morphology, size, anisotropy behavior are important factors for reinforced resin composites.⁶⁰

Recent researches are progressing to further understand how polymers composition influences the success or failure of dental restorations, especially dental resins. Mechanical properties of commercial dental composites modified with electrospun nanopolymers are approaches commonly studied. In Vidotti et al., 60 the effect of bis[4-(methacryloxypropoxy)-phenyl]-propane/triethylene glycol dimethacrylate (BisGMA/TEGDMA) ratio and resin blend to fiber mass ratio on the flexural properties was investigated for electrospun PAN fibers included into resin blends. Their results demonstrated that addition of different ratios of PAN fibers did not affect flexural strength and flexural modulus of the composite beams as compared to neat resin beams; however, the addition of fibers significantly increased the work of fracture of the composite beams, especially for blends with higher TEGDMA ratios.⁶⁰ Surface silanization of electrospun fibers could be a method to prevent detachment of fibers and the resin matrix in BisGMA/TEGDMA reinforced blends with further increase of mechanical

properties as flexural strength, elastic modulus, and work of fracture.⁸⁹ The composite performance is a function of matrix chemistry, fiber diameter, fiber dispersion, and fiber matrix interaction. The performance of simple polymer systems, like acrylates, can be effected by the introduction of small weight percentage (<1 wt.%) of nanoscale reinforcements. 46 Resins for indirect oral application as used in the fabrication of interim restorations⁷⁰ or acrylic denture material⁵⁸ could be also benefited with electrospun method. In Almeida et al., 70 bisacrylic partial fixed dental prostheses with and without electrospun silica-nylon reinforcement were tested for different orientations (horizontal or vertical) before and after thermocycling. Results indicated enhanced the flexural strength values of bisacrylic resin bars and that horizontal orientation provided the highest values of fracture strength for partial fixed dental prostheses.

Electrospun polymers with fiber alignment could be an approach since aligned fibers could present round pattern or crossed lines^{55,58} and dynamic rotating collector is an approach to produce them from different polymers. Fine and nanotexturized biocompatible layers with bioactive compounds could be deposited on dental implants surfaces to increase surface area, reactivity, and interaction with surrounding tissue.⁹⁰ Implant–cement or implant–bone interfaces are required for implant fixation and the filling of possible tissue defects.³³ In Khandaker *et al.*,³³ PCL was deposited with a elestrospinning device in order to create unidirectional fibers on titanium (Ti) plates cemented with poly(methyl methacrylate) (PMMA). The results from fatigue testing and finite-element analysis showed that the addition of the micron- to nanosize PCL fibers on Ti improved the quality of the Ti-PMMA union.

In De Carvalho *et al.*, ⁹¹ cellulose acetate and PEO were electrospun and chlorhexidine (CHX) was incorporated into the mats. The CHX-containing fibers showed antibacterial activity against *S. mutans* and *E. faecalis* based on the test of inhibition halos formed around the nanofiber discs after the bacterial incubation period. Additionally, fibers with CHX increased release over 90 days from the fibers treated with CHX after spinning. ⁹¹ This kind of studies is relevant, because most of the commercial restorative composites do not present bacteriostatic and bactericidal properties and such aspects are desirable since restorative resins facilitate bacterial colonization and biofilm formation. ⁹²

In Dodiuk-Kenig et al., 46 polyvinyl alcohol (PVOH), PLLA, and nylon 6 were electrospun and added at different ratios into Bis-GMA/TEGDMA acrylate resin system with increase of cross linking for 0.05 wt.%, 250 nm PVOH nanofiber-reinforced composite with decrease of linear shrinkage by 50%. Such result was attributed to the incorporation of hyperbranched moieties into the matrix resin assembling to form nanophases that retard motions in the acrylate backbone through the hydroxy-rich chemistry; however, the exact details of the chemical mechanisms were not fully addressed.

To summarize, the increase of the longevity and performance of direct or indirect dental restorations for oral rehabilitations could be assisted by eletrospinning processes. 46,54,55,58,91,93 The achievement of the desirable

benefits of nanofiber reinforcement could be improved with the increase of resin ability to properly wet and penetrate the interfibrillar spaces of the fibrous mesh.

However, not all composites ratios are beneficial and limitations of the bonding between the fibers and the resin matrix are an important problem. The incomplete wetting of the nanofibers by the infiltrating resin could result in air inclusion and voids that ultimately compromises the strength of the material.⁸¹ Another problem is that methacrylate monomers undergo shrinkage when they polymerize generating shrinkage stresses and leaving restorations susceptible to debonding of the tooth/composite interface. This phenomenon could result in gap formation prone to bacterial infiltration and attempt to reduce polymerization shrinkage comprise varying monomer structure, modifying type, size, size distribution, and amount of fillers added to the resin matrix. 94 Finally, another strategy to decrease bacterial infiltration is the incorporation of bacteriostatic and bactericidal agents in dental materials. All promising strategies could be assisted by electrospinning process although further in vitro, in vivo, and clinical investigations are strongly indicated.

Concluding remarks

Collaboration between professionals from different research areas will be essential to the development of new materials and the optimization of techniques for its production, such as electrospinning. Clinical problems and oral health outcomes can be benefited by advances in materials engineering research involving the electrospinning technique.

From the different polymeric materials and applications presented here, it is possible to conclude that the electrospinning technique is increasingly advantageous in oral applications. The improvement of the mechanical properties, the promotion of cell proliferation and differentiation, the degradation rate of different electrospun materials, and the possibility of controlled drug delivery proved the effectiveness of materials produced by this technique. However, some of the analyzed studies require further investigation in order to enable direct human applications. The interaction of nanofiber with biomolecules in animal models may be different in humans and induce different toxic effects. In vivo studies for testing electrospun nanofiber are very low in number compared with in vitro studies. In addition, both in vitro and in vivo toxicity tests must follow well established regulatory guidelines.⁹⁵ This will enhance the performance of products made from fibers and develop new trends and possibilities in oral treatments.

Natural polymers such as cellulose, collagen, and chitosan, appear as potential electrospun materials, low explored, but with high potential for oral applications.

Although several strategies are being developed to achieve materials with superior performance compared to those commercially available, advances in this field of research are incremental and should be combined or optimized for the effectively production of such materials on a larger scale. Despite the controlled production in laboratory is easy to achieve, the large scale production by

electrospinning is in development. For example, in dental and medical applications, the solvent removal from the mat is very important and represents a problem when higher volumes of material are processed. Even though, emerging companies are concerned with these issues, designing innovative devices for industrial scale production. 96 Changes in properties cannot be extrapolated by inverse size analysis of the particular but must be determined through in vitro, in vivo, and clinical studies.

Authors' contributions: This is a multidisciplinary minireview, for this purpose, the professionals that worked were composed by pharmacist, dentist and engineers. All authors participated in design, selection and data analysis and review. ABM, LAGT and DC wrote the manuscript and provided the required information on aspects related to Oral Medicine and Engineering. JVWS helped to discuss polymers from a chemical perspective. ALGM and EB supplied the images from SEM, GEAMB assisted in pharmaceutical aspects described in the minireview.

DECLARATION OF CONFLICTING INTERESTS

All of the authors of this manuscript, we declare that this paper has not been submitted to, nor published previously elsewhere. We disclose any financial and personal relationships with other people or organizations that could inappropriately influence (bias) this work. There are none conflict of interests.

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