

TYRO3: A potential therapeutic target in cancer

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Impact statement

Cancer is among the leading causes of death worldwide. In 2016, 8.9 million people are estimated to have died from various forms of cancer. The current treatments, including surgery with chemotherapy and/or radiation therapy, are not effective enough to provide full protection from cancer, which highlights the need for developing novel therapy strategies. In this review, we summarize the molecular biology of a unique member of a subfamily of receptor tyrosine kinase, TYRO3 and discuss the new insights in TYRO3-targeted treatment for cancer therapy.

Abstract

TYRO3 belongs to the TAM (TYRO3, AXL, and MER) receptor family, a unique subfamily of the receptor tyrosine kinases. Members of TAM family share the same ligand, growth arrest-specific 6, and protein S. Although the signal transduction pathways of TYRO3 have not been evaluated in detail, overexpression and activation of TYRO3 receptor tyrosine kinase have been reported to promote cell proliferation, survival, tumorigenesis, migration, invasion, epithelial-mesenchymal transition, or chemoresistance in several human cancers. Targeting TYRO3 could break the kinase signaling, stimulate antitumor immunity, reduce tumor cell survival, and regain drug sensitivity. To date, there is no specific TYRO3-targeted drug, the effectiveness of targeting TYRO3 in cancer is worthy of further investigations. In this review, we present an update on molecular biology of TYRO3, summarize the development of potential inhibitors of TAM family members, and provide new insights in TYRO3-targeted treatment.

Keywords: Cancer, drug discovery, TAM, therapeutic potential, TYRO3

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Introduction

Cancer is among the leading causes of death worldwide. In 2016, 8.9 million people are estimated to have died from various forms of cancer. The current treatments, including surgery with chemotherapy and/or radiation therapy, are not effective enough to provide full protection from cancer, which highlights the need for novel therapeutic approaches. The TAM family (TYRO3, AXL, and MER), a subfamily of the receptor tyrosine kinases (RTKs), has been reported to regulate different cellular functions, including platelet aggregation, immune responses, and cell growth and differentiation.¹ These receptors share common ligands such as growth arrest-specific gene 6 (Gas6) and protein S (Pros1). Among these RTKs, TYRO3 was first shown to express in tissues associated with myelination in the brain.^{2,3} However, emerging evidence has demonstrated the oncogenic effect of TYRO3 in promoting the survival, chemoresistance, tumorigenesis, and metastasis of cancer cells.¹ This review summarizes recent advances about the mechanisms regulated by the TYRO3 to promote oncogenesis. In addition, we will also discuss

possible strategies of targeting TYRO3 as an anti-cancer regimen.

Molecular biology of TYRO3 receptor

TYRO3 belongs to the TAM family of RTKs. Structurally, TYRO3 exhibits two fibronectin type III domains and two immunoglobulin (Ig)-like domains in the extracellular portion, a transmembrane portion, and a kinase domain in the cytoplasm (Figure 1(a)). TYRO3 receptors bind to their ligands through the Ig-like domains. Gas6 and Pros1 are established nature ligands for TYRO3. After ligand binding, tyrosine residues of TYRO3 are autophosphorylated and downstream signaling is activated. In some cases, high levels of cytoplasmic TYRO3 can be activated even in the absence of a ligand. Under this condition, it functions as a dimeric tyrosine kinase and transforms RatB1a fibroblasts.⁴ These data suggest that even in the absence of its ligand, TYRO3 retains all the properties of the full-length TYRO3 kinase.

There are many aliases for *Tyro3* gene as it was cloned from multiple species by different research groups. In 1991,

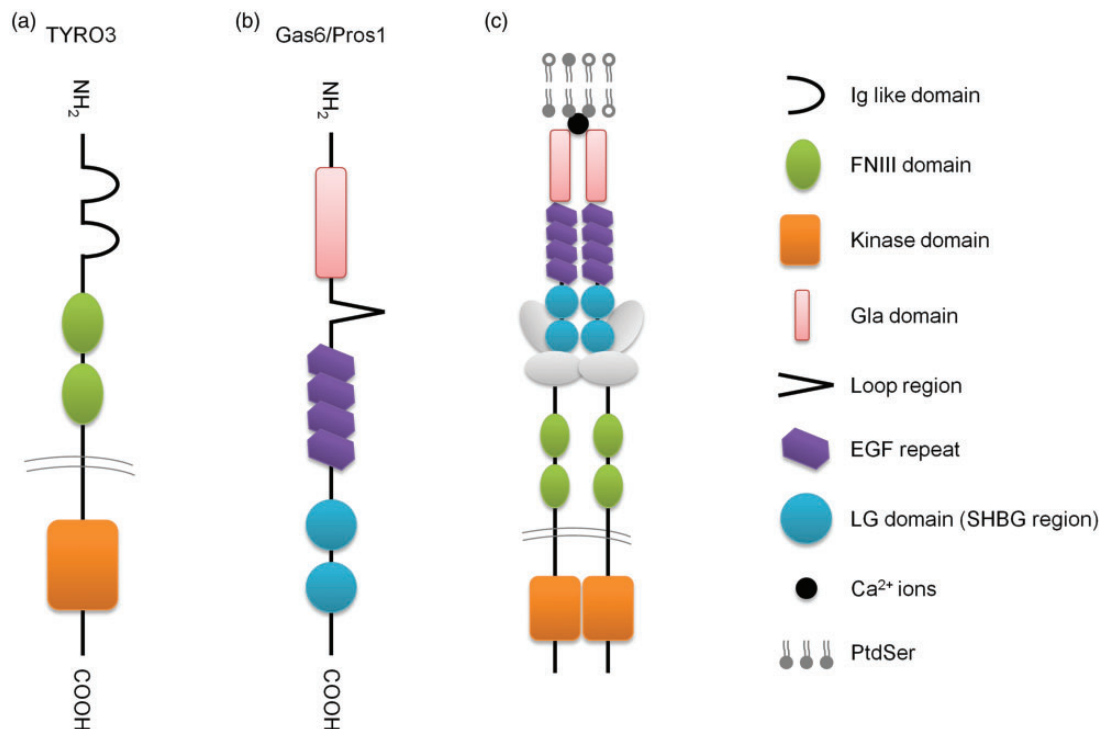


Figure 1. Schematic representation of TYRO3 receptor and ligands protein structure. (a) TYRO3 receptors carry two immunoglobulin (Ig)-like domains in their N-terminus, followed by two fibronectin type III repeats, a transmembrane region and a tyrosine kinase domain in the C-terminal intracellular region. (b) Growth-arrest-specific 6 (GAS6) and protein S (Pros1), TYRO3 ligands, carry a Gla domain in their N-terminus, followed by four EGF repeats and two laminin G (LG) domains in their C-terminus. (c) TYRO3 dimers bind to their ligands through interaction between the two N-terminal Ig-like domains of the receptors and the two C-terminal LG regions, which together make up the sex hormone binding globulin (SHBG) domain of the ligands. γ -carboxylation of glutamic acid residues in the Gla domain and calcium ions (Ca^{2+}), enable Gas6 and Pros1 to bind to phosphatidylserine (PtdSer). The two LG domains form a sex hormone binding globulin-like domain and trigger the activation of TYRO3. (A color version of this figure is available in the online journal.)

Tyro1 to *Tyro13* were found from rat brain.² *Tyro3*, *Tyro7*, and *Tyro12* were grouped into a subfamily based on the unique amino acid sequences found in their kinase domains. Afterwards, it was found that *Tyro7* and *Tyro12* are the same genes as *Axl* and *Mer*, respectively, while *Tyro3* became the third member of the TAM family. In 1993, fragments of murine *Tyro3* were found and named *Etk2*. In 1994, several TAM family genes were cloned from mouse and human by different groups. The murine gene was named *Dtk*,⁵ *Br*,⁶ *Rse*,⁷ or *Tyro3*,⁸ while the human gene was called *Sky*,⁹ *Tif*,¹⁰ or *Rse*.⁷ Two years later, it was learned that *Dtk* and *Br* were encoded by the same gene with alternative splicing.¹¹ There are three splicing variants for *TYRO3* that contain exons 2A, 2B, and 2C, respectively.^{11–13} These exons encode different signaling peptide sequences, indicating that the expression of these alternative splicing variants may affect the subcellular localization and thus the function of *TYRO3*.

Ligands and structures

The endogenous ligands for *TYRO3* receptors are the Gas6 and Pros1. The structure of Gas6 and Pros1 is related to vitamin K. They share approximately 40% sequence identities with an N-terminal γ -carboxyglutamic acid domain, four tandem EGF-like domains, and a C-terminal sex hormone-binding globulin domain (Figure 1(b)).^{14,15} Pros1 is known to regulate anticoagulation and

complement cascades. It can be purified using *TYRO3*-phosphorylating activity as an indicator¹⁶ since purified recombinant murine Pros1 binds to and activates both MER and *TYRO3* (*TYRO3*>MER).¹⁷ Currently, there is no evidence that Pros1 activates AXL. Gas6 was originally identified based on its dramatic upregulation after growth arrest with unknown function.^{18,19} In 1995, it was reported that Gas6 could bind and activate AXL.^{16,20} Shortly thereafter, Gas6 was found to activate all TAM receptors (AXL>*TYRO3*>>>MER).²¹ Since the secretion signal and the γ -carboxyglutamic acid domain are highly conserved in human, mouse, and bovine, Gas6 subfamily members are 74–81% homologous to each other and moderately homologous to human and bovine Pros1.¹⁶

The glutamic acid residue is required for the binding of *TYRO3* to the phosphatidylserine of the cell membrane in a calcium-dependent manner,²² especially when it is γ -carboxylated.^{23,24} The two laminin G motifs within the C-terminal sex hormone-binding globulin domain are required for the binding to *TYRO3* and the activation of downstream signaling pathways including phosphatidylinositol 3-kinase (PI3K)/AKT, ERK, and PLC- γ (Figure 1(c)).^{25–27} The functional importance of other domains of GAS6 and Pros1 awaits further characterization.

Two potential *TYRO3* ligands, tubby-like protein (Tulp) 1 and Tulp2, were identified recently and linked to phagocytosis.²⁸ By co-immunoprecipitation, Tulp1 was found to interact with MER, AXL, and *TYRO3*, while Tulp2 can be

co-precipitated with AXL and TYRO3, but not with MER. These results suggested that Tulp1 and Tulp2 have distinct binding specificities to TYRO3. Unlike Gas6 and Pros1, Tulp ligands lack the signature laminin G motifs for receptor binding but contain minimal phagocytic determinant (MPD) as a new type of TAM-binding motif. It is suggested that the five MPDs of mouse Tulp1 may cause homo- and/or hetero-dimerization of TAM receptors, though it is unclear whether one or multiple receptors will be bound.²⁹ Interestingly, Tulp proteins lack signal peptide and have been identified as intracellular proteins by immunohistochemistry.³⁰ How does intracellular Tulps interact with plasma membrane receptors to facilitate phagocytosis? One explanation for Tulp1 functions as phagocytosis ligand is via active secretion through a non-classical pathway coined unconventional secretion. Similar mechanism has been reported for a number of proteins without a classical signal peptide.³¹ Indeed, Caberoy and Li³² had demonstrated that Tulp1 can be secreted to extracellular space, which cannot be blocked by brefeldin A and monensin, inhibitors that block protein transport via the endoplasmic reticulum-Golgi pathway. This finding supports the notion that Tulp proteins can function as TAM receptor ligands; nevertheless, their functions other than facilitating phagocytosis remain to be characterized.

TYRO3 in development

The expression of TYRO3 was found in embryonic developing mouse tissue, including undifferentiated mouse embryonic stem cells. Therefore, TYRO3 was initially named Developmental Tyrosine Kinase.⁵ The TAM receptors play overlapping and unique roles in regulating immunity and inflammation. Inactivation of all TAM receptors does not lead to embryonic lethality, indicating the overlapping and compensatory roles of TAM receptors demonstrated in various cell types for embryogenesis.^{13,33} However, TYRO3 receptor and their ligands are largely expressed during development in gastrointestinal, nervous, and reproductive systems.^{5,13} Inactivation of TYRO3 receptors or their ligands affects postnatal developmental processes. For example, the neural degeneration with seizures and paralysis is observed in *Tyro3* knockout mice.¹³ With aging, *Tyro3*-null mice develop cerebellar ataxia, and immune defects are observed with the progressive loss of the TAM family.²⁹ A previous study indicated that Gas6 supports gonadotropin-releasing hormone (GnRH) neuronal survival through AXL and TYRO3, which are expressed by migratory GnRH neurons. *Axl* and *Tyro3* deficiencies resulted in increased apoptosis and delayed migration with a reduction in GnRH neurons reaching their final destination.³⁴ This strain manifested later in adulthood in delayed sexual maturation and irregular estrus cycles with a significantly prolonged proestrus phase.³⁴ Moreover, triple knockouts of TAM receptors demonstrated multiple debilitating and degenerative traits, most importantly, impaired hemostasis, immunoregulation, spermatogenesis, and blindness.^{13,35}

Disease relevance of TYRO3

TYRO3 appears to have a critical role in the immunity, phagocytosis, hemostasis, and neuronal disease. The first evidence suggests TYRO3 may be involved in platelet aggregation came from studies of *Gas6*-null mice. Knockout of *Gas6* protects mice from thrombosis and causes platelet dysfunction.³⁶ A follow-up study demonstrated that platelet granule secretion was limited in *Tyro3*^{-/-} mice. This strain demonstrated reduced thrombus formation and decreased platelet aggregation stability.³⁷ In neuron, TYRO3 activation due to progranulin reduction results in activation of PKC α via PLC γ , inducing tau phosphorylation, mislocalization of tau to dendritic spines, and spine loss.³⁸ Genetic variants of *TYRO3* were associated with an increased risk for several immune-related disorders. Functional *TYRO3* variants alter gene expression in the pathogenesis of allergic sensitization and allergic rhinitis. Multiple intronic variants in *TYRO3* were also associated with asthma. The most significant association was at a single-nucleotide polymorphism located within several putative transcription factor binding sites.³⁹ Moreover, TYRO3 contributes to anxiety phenotypes and can delay hypothalamic neurodegeneration in *anx/anx* mice. In their study, Kim *et al.*⁴⁰ identified a mutation in the signal sequence of the *Tyro3* (R7W-Tyro3) as an enhancing *anx* phenotype.

Involvement of TYRO3 in cancer

The first study that indicates TYRO3 exerts oncogenic capacity was evidenced by showing *Tyro3*-transfected Rat-2 fibroblasts could grow in soft agar.⁸ Later on, rat fibroblasts overexpressing TYRO3 were shown to be able to form tumors in nude mice and the transcripts of *Tyro3* have been associated with human and mouse mammary tumors further support this notion.^{4,41} Like other TAM receptor family members, TYRO3 and ligand overexpression have been shown in a wide range of cancers, and correlate with poor prognosis in a variety of tumor types (Table 1). Through AKT/NF κ B signaling, TYRO3 exerts pro-survival effects and promotes cancer cell growth.⁴² TYRO3 and AXL protein levels are undetectable in normal thyroid cells but significantly upregulated and activated in thyroid cancer cells.⁴³ TYRO3 also triggers the tyrosyl-phosphorylation of ACTN4, a member of actin binding protein family involved in motility. Knockdown of *Tyro3* by siRNA prevents melanoma cell migration and invasion.⁴⁴

Activated TYRO3 promotes the survival, invasion, migration, proliferation, and transformation of cancer cells (Figure 2).^{45,46} Increasingly, evidence supporting the notion that overexpression of TYRO3 contributes to the resistance to conventional and targeted therapies in thyroid cancer, and blocking its signaling dramatically reduced cell viability and resistance to apoptotic stimuli.⁴³ TYRO3 was also shown to promote cell proliferation and chemoresistance in breast cancer.¹⁴² Increased resistance to platinum and taxol secondary to TYRO3 overexpression has also been reported in ovarian cancer.⁴⁷ Ovarian cancer cells

Table 1. Expression and function of TAM family kinases and ligands in cancer.

TAM kinase or ligand	Ectopic expression or overexpression	Functional roles (proliferation, survival, or tumorigenesis)	Metastatic roles (migration, invasion, or EMT)	Roles in chemoresistance	Prognostic importance	References
TYRO3	Multiple myeloma, lung cancer, breast cancer, melanoma, esophageal cancer, hepatocellular carcinoma, colorectal cancer, prostate cancer, ovarian cancer, endometrial cancer, leiomyosarcoma, thyroid cancer, schwannoma, ALL, AML, CML, and B-CLL	Melanoma, hepatocellular carcinoma, colorectal cancer, breast cancer, ovarian cancer, thyroid cancer	Colorectal cancer, prostate cancer, lung cancer	Melanoma, hepatocellular carcinoma, colorectal cancer, breast cancer, ovarian cancer, and thyroid cancer	Hepatocellular carcinoma, colorectal cancer, breast cancer, breast cancer	41–43,45–69
AXL	Lung cancer, glioblastoma, breast cancer, colorectal cancer, gastric cancer, pancreatic cancer, oesophageal cancer, melanoma, squamous cell skin cancer, prostate cancer, endometrial cancer, ovarian cancer, oral squamous cell carcinoma, thyroid cancer, bladder cancer, renal cancer, schwannoma, mesothelioma, Kaposi's sarcoma, osteosarcoma, AML, CML, and B-CLL, Lung cancer, glioma, melanoma, prostate cancer, schwannoma, mantle cell lymphoma, AML, and ALL	Prostate cancer, ovarian cancer, breast cancer, thyroid cancer, lung cancer, pancreatic cancer, melanoma, hepatocellular carcinoma, glioblastoma, mesothelioma, osteosarcoma, schwannoma, Kaposi's sarcoma, and oesophageal cancer	Breast cancer, lung cancer, melanoma, prostate cancer, pancreatic cancer, ovarian cancer, hepatocellular carcinoma, thyroid cancer, bladder cancer, Kaposi's sarcoma, mesothelioma, oesophageal cancer, glioblastoma, colorectal cancer, cervical cancer, neuroblastoma, and osteosarcoma	Breast cancer, lung cancer, ovarian cancer, oesophageal cancer, AML, and CML	Lung cancer, glioblastoma, osteosarcoma, oral squamous cell carcinoma, breast cancer, head and neck cancer, colorectal cancer, pancreatic cancer, oesophageal cancer, ovarian cancer, gastric cancer, bladder cancer, and AML	43,55,57–59, 70–110
MER	Lung cancer, glioma, melanoma, prostate cancer, schwannoma, mantle cell lymphoma, AML, and ALL	Glioma, lung cancer, melanoma, AML, and ALL	Glioblastoma, and melanoma	Glioma, lung cancer, pancreatic cancer, breast cancer, and ALL	Gastric cancer	57,59,73,79, 83,97,99, 110–118
Gas6	Multiple myeloma, glioblastoma, breast cancer, gastric cancer, endometrial cancer, ovarian cancer, thyroid cancer, renal cancer, schwannoma, AML, ALL, and CML	Lymphoma, breast cancer, prostate cancer, colorectal cancer, pancreatic cancer, thyroid cancer, schwannoma, gastric cancer, osteosarcoma, and renal cancer	Breast cancer, prostate cancer, pancreatic cancer, hepatocellular carcinoma, gastric cancer, osteosarcoma, and renal cancer	ALL	Lung cancer, glioblastoma and renal cancer, and AML	43,57–59,78,86, 91,94,96,101, 119–129
Pros1	Thyroid cancer, colorectal cancer, pancreatic cancer, brain tumours, lung cancer, prostate cancer, ovarian cancer, glioblastoma, osteosarcoma, and AML	Thyroid cancer and glioblastoma	Prostate cancer and thyroid cancer	Prostate cancer	Prostate cancer and glioblastoma	130–141

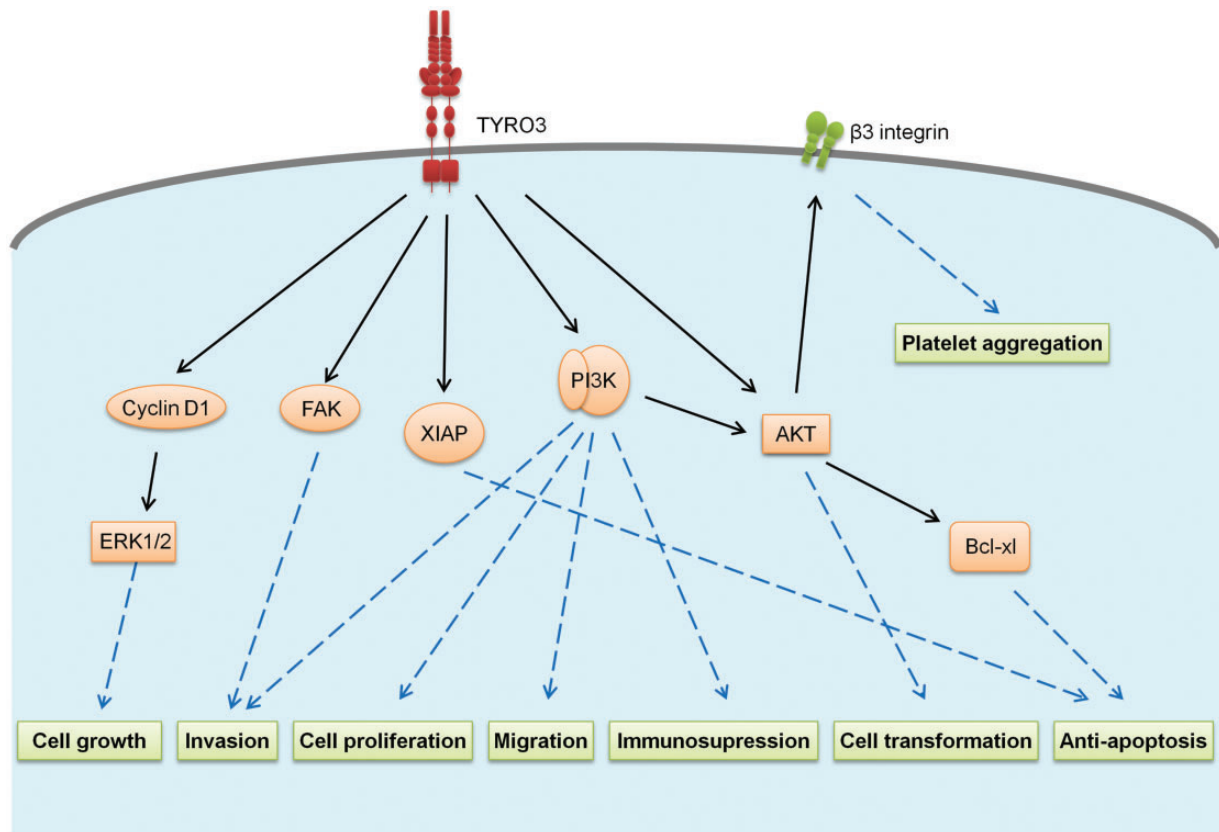


Figure 2. Schematic representation of TYRO3 signaling pathway. TYRO3 has been reported to mediate numerous cellular activities, including platelet aggregation, cell growth, invasion, cell proliferation, migration, immunosuppression, cell transformation, and anti-apoptosis. Molecules in orange have been shown to associate with TYRO3 through either a direct or indirect interaction. (A color version of this figure is available in the online journal.)

overcome treatment resistance via upregulation of TYRO3 and AXL expression, AKT phosphorylation, and Bcl-xl expression.⁴⁷ A growing body of evidence demonstrated that epithelial-mesenchymal transition may be a major mechanism in drug resistance.^{143,144} TYRO3 promotes phagocytosis and inhibits inflammation, allowing resistance to antitumor treatments to further cancer progression.¹⁴⁵ Interestingly, recent study demonstrated that treatment with the human TYRO3 antibody abolished TYRO3-induced EMT process in colon cancer.⁴⁸ Taken together, these studies suggested that inhibition of TYRO3 and its signaling pathways could have therapeutic benefits in cancer development.

TYRO3 signaling pathways

The TYRO3 receptor is the least studied TAM receptors with largely unknown signaling pathways. However, some sporadic studies did report the involvement of different signaling molecules involved in transmitting signals of TYRO3 (Figure 2). One study indicated a possible interaction between TYRO3 and a phosphorylated Src family kinase in COS cells.¹⁴⁶ Previous studies identified multiple proteins that may interact with TYRO3, including Ran binding protein in microtubule organizing center, protein phosphatase 1, and the p85 β -subunit of PI3K.^{147,148} Another study showed that Pros1/TYRO3 activates the

PI3K/AKT pathway, which protects neurons from excitotoxic injury and apoptosis in mouse cortical neurons.^{149,150} Epidermal growth factor receptor (EGFR)/TYRO3 chimeric receptor also showed that the cytoplasmic domain of TYRO3 associated with PI3K and led to transformation of NIH3T3 cells.¹⁴⁷ The mitogen-activated protein kinase (MAPK) signaling pathway has also been linked to TYRO3 activation. It has been shown that Gas6 increased the phosphorylation of AKT and ERK1/2.¹⁵¹ TYRO3 acts on mature osteoclasts through activation of ERK1/2 MAPK, possibly contributing to the bone loss by estrogen deficiency.¹⁵² TYRO3 also regulates the proliferation of MCF-7 breast cancer cells through control of cyclin D1 expression and phosphorylation of ERK1/2 or STAT3.^{142,153} These studies reveal that TYRO3 may utilize distinct signaling pathways to transmit its message in different cell types. Further study is warranted to systemically characterize the second messengers directly downstream of TYRO3.

Therapeutic potential of targeting TYRO3

Because the TAM family has been implicated in the pathogenesis of several cancers, the therapeutic potential of targeting the TAM family has been validated. To date, many pan-TAM inhibitors have proven to be both efficacious and less toxic than standard chemotherapies. These inhibitors

may prevent activation of the TAM family and other RTKs leading to promotion of cell death, thereby preventing cancer metastasis. It is found that the selectivity is primarily determined by the size and configuration of kinase's ATP-binding site. The location and shape of active-site residues of MER and AXL are highly consistent, suggesting that small-molecule inhibitors generally have a low MER-over-AXL selectivity and a high MER-over-TYRO3 selectivity.

Due to the centrality of TYRO3 in immune disease and cancer development, there is interest in targeting TYRO3. A recent study showed that TYRO3 was overexpressed in nearly all melanoma cell lines. Knockdown of TYRO3 by short hairpin RNA led to significantly inhibited cell proliferation.^{49,50} Administration of anti-TYRO3 antibody increased cell death signal and drug sensitivity *in vitro* and *in vivo*.^{48,49} Furthermore, miR-7 was identified as a potent tumor suppressor of human hepatocellular carcinoma. It targets TYRO3 and regulates proliferation, migration, and invasion through the PI3K/protein kinase B pathway.⁴⁵ These findings indicated that TYRO3 is a drugable target in cancer. However, because no specific TYRO3-targeted drugs are available, the role of specific inhibitors against TYRO3 as a therapeutic target has not previously been evaluated in detail. Below is a summary of some representative TAM inhibitors that also work on TYRO3 (Table 2).

Metformin

Since the primary effect of metformin is glucose metabolism modification, it has been widely used to treat type 2 diabetes.¹⁷³ Accumulating evidence suggests it also possesses anticancer effects on cell proliferation in various cancers and tumor growth in xenograft model.^{174–177} The efficacy of metformin for the treatment of endometrial, cervical, breast, and ovarian cancer has been suggested in preclinical studies and clinical trials.¹⁵⁴ The anticancer mechanisms of metformin have been assessed by its ability to inhibit pro-survival and anti-apoptotic signals mediated by mammalian target of rapamycin complex 1, EGFR, and MAPK.^{178–181} Metformin may target TYRO3 to prevent cell proliferation and reduce chemoresistance.¹⁵⁵ Collectively, these studies indicated a potent therapeutic strategy to facilitate the anticancer activity of metformin and overcome chemoresistance in cancer cells.

Compounds 21 and 24

High-throughput screening identified a novel series of spiroindoline-based inhibitors as the first TYRO3-selective tyrosine kinase inhibitors.^{156,182} Among these, compounds 21 and 24, 2,4-diaminopyrimidine-5-carboxamide inhibitors, are potent inhibitors of TYRO3 kinase (Sky IC₅₀ = 0.0007 μ M and 0.015 μ M, respectively).¹⁵⁶ The compound 21, which replaces the entire amide sidechain with a 3-methylisoxazole from an 2, 4-diaminopyrimidine-5-carboxamide inhibitor, exhibited excellent selectivity in 46/48 kinases with some activity in MAP4K4 and Mer. Compound 24 which replaces amide sidechain by a

simple bromine atom has moderate functional P-selectin inhibition, good human liver microsomes, and rat liver microsomes metabolism stability. However, the low aqueous solubility and PAMPA permeability were not predictive of good oral bioavailability.

LDC1267

LDC1267 is an inhibitor of the TAM kinase family in cells at low nanomolar levels with IC₅₀ of <5 nM, 8 nM, and 29 nM for MER, TYRO3, and AXL, respectively.¹⁵⁷ Treatment of wild-type natural killer cells with LDC1267 conferred therapeutic potential, efficiently enhancing anti-metastatic NK cell activity *in vivo*. Administration of LDC1267 markedly reduced murine mammary cancer development and metastasis of melanoma.¹⁵⁷

BMS-777607

BMS-777607 is a small molecule inhibitor for c-Met, AXL, Ron, and TYRO3 with IC₅₀ of 3.9 nM, 1.1 nM, 1.8 nM, and 4.3 nM, 40-fold more selective for Met-related targets versus Lck, VEGFR-2, and TrkA/B, and more than 500-fold greater selectivity versus all other receptor and non-receptor kinases.¹⁵⁸ Previous studies demonstrated the effect of BMS-777607 in antitumor treatment may be mediated by blocking hepatocyte growth factor-stimulated phosphorylation of Met and downstream pathways.^{159,183,184} BMS-777607 inhibits hepatocyte growth factor-stimulated cell scatter, motility, and invasion *in vitro*.^{159,185} Surprisingly, BMS777607 was also reported to have inhibitory effects on TYRO3 in cell-free conditions.¹⁶⁹ However, the efficacy of BMS-777607 against TYRO3 as a therapeutic target has not been evaluated. Due to its potency and promising pharmacokinetic and preclinical safety profiles, BMS-777607 has been advanced into phase 1/2 clinical trials.

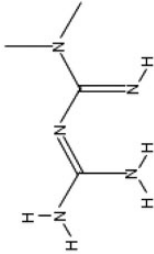
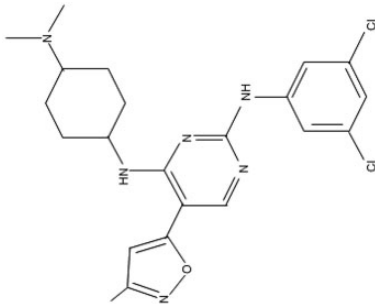
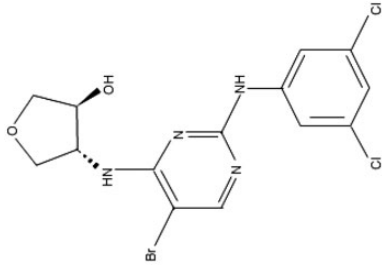
UNC2250

UNC2250 is a potent and selective MER inhibitor with IC₅₀ of 1.7 nM, about 160- and 60-fold selectivity over the closely related kinases AXL/TYRO3.¹⁶⁵ MER inhibition by UNC2250 decreased activation of downstream AKT and p38, inhibited proliferation and invasion in mantle cell lymphoma (MCL), and sensitized MCL cells to treatment with vincristine *in vitro* and doxorubicin *in vitro* and *in vivo*.¹⁶⁶

UNC2881

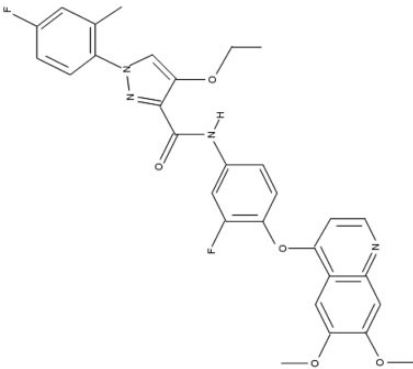
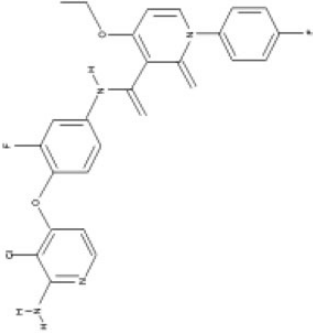
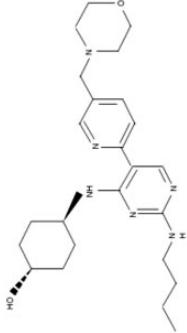
UNC2881 is a specific MER tyrosine kinase inhibitor with IC₅₀ of 4.3 nM, about 83- and 58-fold selectivity over AXL and TYRO3, respectively. UNC2881 inhibits steady-state MER kinase phosphorylation with an IC₅₀ value of 22 nM *in vitro*. Treatment with UNC2881 is also sufficient to block EGF-mediated stimulation of EGFR/MER chimeric receptor. In addition, UNC2881 may have utility for prevention and/or treatment of pathologic thrombosis by inhibiting platelet aggregation.¹⁶⁷

Table 2. TYRO3 as a therapeutic target.

Structure	Name	Target profile		Activity			Development stage/ outcome	References
		Target(s)	TYRO3	AXL	MER			
	Metformin	TYRO3, AXL	-	-	-	Approved (Diabetes) Approved for ovarian cancer cells, endometrial, breast, and ovarian cancer	154,155	
	Compound 21	TYRO3	IC ₅₀ =0.7 nM	-	-	Preclinical	156	
	Compound 24	TYRO3	IC ₅₀ =15 nM	-	-	Preclinical	156	

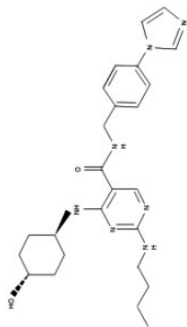
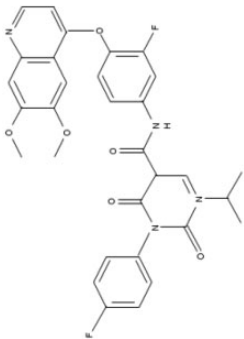
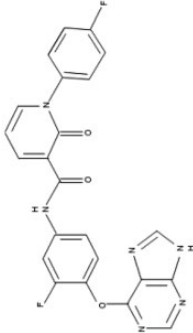
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Table 2. Continued

Structure	Name	Target profile		Activity			Development stage/ outcome	References
		Target(s)		TYRO3	AXL	MER		
	LDC1267	TYRO3,AXL, MER		IC ₅₀ =8 nM	IC ₅₀ =29 nM	IC ₅₀ <5 nM	Preclinical Approved for murine mammary cancer and melanoma metastases dependent on NK cells	157
	BMS-777607	c-Met, TYRO3,AXL, Ron,		IC ₅₀ =4.3 nM	IC ₅₀ =1.1 nM	-	Phase 1/2. Approved for advanced or metastatic solid tumors	158-164
	UNC2250	TYRO3,AXL, MER		IC ₅₀ ~100 nM	IC ₅₀ ~270 nM	IC ₅₀ =1.7 nM	Phase 1 Approved for mantle cell lymphoma	165,166

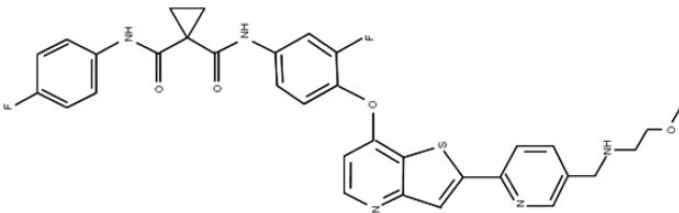
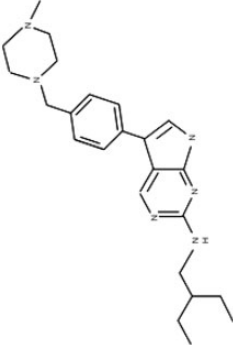
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Table 2. Continued

Structure	Name	Target profile		Activity			Development stage/ outcome	References
		Target(s)		TYRO3	AXL	MER		
	UNC2881	TYRO3,AXL, MER		IC ₅₀ ~250 nM	IC ₅₀ ~350 nM	IC ₅₀ =4.3 nM	Preclinical Approved for pathologic thrombosis	167
	RXDX-106	TYRO3,AXL, MER, c-Met		IC ₅₀ =19 nM	IC ₅₀ =7 nM	IC ₅₀ =29 nM	Phase 1 Approved for immuosuppression of innate immune cells and gastric cancer cell lines	168
	6g	TYRO3,AXL, MER, Met		Kd=200 nM	Kd=39 nM	Kd=42 nM	Preclinical	169

(continued)

Table 2. Continued

Structure	Target profile		Activity			Development stage/ outcome	References
	Name	Target(s)	TYRO3	AXL	MER		
	Sitravatinib (MGCD516)	TYRO3,AXL, MER	IC ₅₀ <1 nM	IC ₅₀ <1 nM	IC ₅₀ <1 nM	Phase 1/2	170,171
	UNC Compound 5	TYRO3	IC ₅₀ =6.7 nM	IC ₅₀ =206 nM	IC ₅₀ =19 nM	Preclinical	172
shRNA		TYRO3	-	-	-	Preclinical	49,50
Antibodies		TYRO3	-	-	-	Approved for leiomyosarcoma and melanoma Preclinical Approved for colon cancer and melanoma cell lines	48,49

Kd=dissociation constant.

RXDX-106

RXDX-106 is an oral immunomodulatory agent that can restore and enhance overall immune function. It inhibits the activity of TAM- and c-Met-induced protumorigenesis by a decrease in downstream MAPK and PI3K signaling and cell viability. RXDX-106 could reverse immunosuppression of innate immune cells, inhibit tumors harboring activating TAM gene fusions, and affect the TAM-expressing tumor microenvironment resulting in a global anti-cancer environment.¹⁶⁸ Phase 1 study is expected to commence in early 2018.

6g

The affinity of 6g for AXL, MER, Met, and TYRO3 is 39, 42, 65, and 200 nM, respectively.¹⁶⁹ The absence of cytotoxicity against several tumor cell lines in culture makes this inhibitor a good candidate for the growth inhibition of tumor cells that would overexpress a gene belonging to the TAM subfamily.

Sitravatinib (MGCD516)

MGCD516 is a novel small molecule inhibitor targeting TAM family ($IC_{50} < 1$ nM) and multiple RTKs involved in driving sarcoma cell growth. MGCD516 treatment induced potent anti-proliferative effects *in vitro* and suppressed tumor growth *in vivo*.¹⁷⁰ As an immuno-oncology agent, MGCD516 may target the tumor microenvironment, resulting in innate and adaptive immune cell changes that augment immune checkpoint blockade.¹⁷¹ MGCD516 is being evaluated in combination with checkpoint blockade (nivolumab) for refractory non-small cell lung cancer in phase 2 clinical study.¹⁷¹

UNC compound 5

More recently, UNC2541-derived compound 5, a TYRO3-selective pyrrolopyrimidine-based inhibitor was mentioned. They reported this inhibitor with more selective against TYRO3 over MER (3-fold) and AXL (31-fold).¹⁷² However, the pharmacokinetic properties of Compound 5 are unclear. Thus, more work is needed in the development of more potent TYRO3 inhibitors.

Current and future development

Despite numerous efforts, many traditional therapies are ineffective due to the pathological and etiological complexity of cancer. As with most drugs, chemotherapy drugs do have side effects. Therefore, it is important to develop effective and safe strategies for cancer prevention and treatment.

The important role of TYRO3 in cancer development has been elucidated. Targeting TYRO3 represents a novel therapeutic approach by suppressing tumor cell survival, proliferation, invasion, chemoresistance, and de-repression of the immune activities. Therefore, therapeutic TYRO3 inhibition may sensitize tumor cells to killing by chemotherapy, radiation, or other targeted agents. Specific TYRO3-targeted drug may enhance immunotherapeutic efficacy in combination with immune checkpoint inhibitors.

If these combination therapies are effective against metastatic disease, then TYRO3-targeted drug could be used in early stages as an adjuvant to provide cancer patients with new options for durable responses. However, development of autoimmunity is a consideration for TYRO3 inhibition treatment.

Although many potent drugs have been developed as mentioned in this review, including different compounds, multi-target TYRO3 inhibitors, and antibody, the ability of these drugs to defeat cancers by TYRO3 and reduce drug resistance is unclear. A better understanding of TYRO3 could lead to more effective anticancer strategies.

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DECLARATION OF CONFLICTING INTERESTS

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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REFERENCES

1. Vouri M, Hafizi S. TAM receptor tyrosine kinases in cancer drug resistance. *Cancer Res* 2017;**77**:2775–8
2. Lai C, Lemke G. An extended family of protein-tyrosine kinase genes differentially expressed in the vertebrate nervous system. *Neuron* 1991;**6**:691–704
3. Goudarzi S, Rivera A, Butt AM, Hafizi S. Gas6 promotes oligodendrogenesis and myelination in the adult central nervous system and after lysocleithin-induced demyelination. *ASN Neuro* 2016;**8**:1–14
4. Taylor IC, Roy S, Varmus HE. Overexpression of the Sky receptor tyrosine kinase at the cell surface or in the cytoplasm results in ligand-independent activation. *Oncogene* 1995;**11**:2619–26
5. Crosier PS, Lewis PM, Hall LR, Vitas MR, Morris CM, Beier DR, Wood CR, Crosier KE. Isolation of a receptor tyrosine kinase (DTK) from embryonic stem cells: structure, genetic mapping and analysis of expression. *Growth Factors* 1994;**11**:125–36
6. Fujimoto J, Yamamoto T. brt, a mouse gene encoding a novel receptor-type protein-tyrosine kinase, is preferentially expressed in the brain. *Oncogene* 1994;**9**:693–8
7. Mark MR, Scadden DT, Wang Z, Gu Q, Goddard A, Godowski PJ. rse, a novel receptor-type tyrosine kinase with homology to Axl/Ufo, is expressed at high levels in the brain. *J Biol Chem* 1994;**269**:10720–8
8. Lai C, Gore M, Lemke G. Structure, expression, and activity of Tyro 3, a neural adhesion-related receptor tyrosine kinase. *Oncogene* 1994;**9**:2567–78
9. Ohashi K, Mizuno K, Kuma K, Miyata T, Nakamura T. Cloning of the cDNA for a novel receptor tyrosine kinase, Sky, predominantly expressed in brain. *Oncogene* 1994;**9**:699–705

10. Dai W, Pan H, Hassanain H, Gupta SL, Murphy MJ, Jr. Molecular cloning of a novel receptor tyrosine kinase, tif, highly expressed in human ovary and testis. *Oncogene* 1994;**9**:975–9
11. Lewis PM, Crosier KE, Wood CR, Crosier PS. Analysis of the murine Dtk gene identifies conservation of genomic structure within a new receptor tyrosine kinase subfamily. *Genomics* 1996;**31**:13–9
12. Biesecker LG, Giannola DM, Emerson SG. Identification of alternative exons, including a novel exon, in the tyrosine kinase receptor gene Etk2/tyro3 that explain differences in 5' cDNA sequences. *Oncogene* 1995;**10**:2239–42
13. Lu Q, Gore M, Zhang Q, Camenisch T, Boast S, Casagrande F, Lai C, Skinner MK, Klein R, Matsushima GK, Earp HS, Goff SP, Lemke G. Tyro-3 family receptors are essential regulators of mammalian spermatogenesis. *Nature* 1999;**398**:723–8
14. Mark MR, Chen J, Hammonds RG, Sadick M, Godowsk PJ. Characterization of Gas6, a member of the superfamily of G domain-containing proteins, as a ligand for Rse and Axl. *J Biol Chem* 1996;**271**:9785–9
15. Sasaki T, Knyazev PG, Cheburkin Y, Gohring W, Tisi D, Ullrich A, Timpl R, Hohenester E. Crystal structure of a C-terminal fragment of growth arrest-specific protein Gas6. Receptor tyrosine kinase activation by laminin G-like domains. *J Biol Chem* 2002;**277**:44164–70
16. Stitt TN, Conn G, Gore M, Lai C, Bruno J, Radziejewski C, Mattsson K, Fisher J, Gies DR, Jones PF, Masiakowski P, Ryan TE, Tobkes NJ, Chen DH, DiStefano PS, Long GL, Basilico C, Goldfarb MP, Lemke G, Glass DJ, Yancopoulos GD. The anticoagulation factor protein S and its relative, Gas6, are ligands for the Tyro 3/Axl family of receptor tyrosine kinases. *Cell* 1995;**80**:661–70
17. Prasad D, Rothlin CV, Burrola P, Burstyn-Cohen T, Lu Q, Garcia de Frutos P, Lemke G. TAM receptor function in the retinal pigment epithelium. *Mol Cell Neurosci* 2006;**33**:96–108
18. Manfioletti G, Brancolini G, Avanzi G, Schneider C. The protein encoded by a growth arrest-specific gene (gas6) is a new member of the vitamin K-dependent proteins related to protein S, a negative coregulator in the blood coagulation cascade. *Mol Cell Biol* 1993;**13**:4976–85
19. Schneider C, King RM, Philipson L. Genes specifically expressed at growth arrest of mammalian cells. *Cell* 1988;**54**:787–93
20. Varnum BC, Young C, Elliott G, Garcia A, Bartley TD, Fridell YW, Hunt RW, Trail G, Clogston C, Toso RJ, Yanagihara D, Bennett L, Sylber M, Merewether LA, Tseng A, Escobar E, Liu ET, Yamane HK. Axl receptor tyrosine kinase stimulated by the vitamin K-dependent protein encoded by growth-arrest-specific gene 6. *Nature* 1995;**373**:623–6
21. Nagata K, Ohashi K, Nakano T, Arita H, Zong C, Hanafusa H, Mizuno K. Identification of the product of growth arrest-specific gene 6 as a common ligand for Axl, Sky, and Mer receptor tyrosine kinases. *J Biol Chem* 1996;**271**:30022–7
22. Huang M, Rigby AC, Morelli X, Grant MA, Huang G, Furie B, Seaton B, Furie BC. Structural basis of membrane binding by Gla domains of vitamin K-dependent proteins. *Nat Struct Mol Biol* 2003;**10**:751–6
23. Tsou WI, Nguyen KQ, Calarese DA, Garforth SJ, Antes AL, Smirnov SV, Almo SC, Birge RB, Kotenko SV. Receptor tyrosine kinases, TYRO3, AXL, and MER, demonstrate distinct patterns and complex regulation of ligand-induced activation. *J Biol Chem* 2014;**289**:25750–63
24. Lew ED, Oh J, Burrola PG, Lax I, Zagorska A, Traves PG, Schlessinger J, Lemke G. Differential TAM receptor-ligand-phospholipid interactions delimit differential TAM bioactivities. *eLife* 2014;**3**:e03385
25. Todd JC, Hu B, Curtis JL. The receptor tyrosine kinase MerTK activates phospholipase C gamma2 during recognition of apoptotic thymocytes by murine macrophages. *J Leukoc Biol* 2004;**75**:705–13
26. Li R, Chen J, Hammonds G, Phillips H, Armanini M, Wood P, Bunge R, Godowski PJ, Sliwkowski MX, Mather JP. Identification of Gas6 as a growth factor for human Schwann cells. *J Neurosci* 1996;**16**:2012–9
27. Fridell YW, Jin Y, Quilliam LA, Burchert A, McCloskey P, Spizz G, Varnum B, Der C, Liu ET. Differential activation of the Ras/extracellular-signal-regulated protein kinase pathway is responsible for the biological consequences induced by the Axl receptor tyrosine kinase. *Mol Cell Biol* 1996;**16**:135–45
28. Caberoy NB, Zhou Y, Li W. Tubby and tubby-like protein 1 are new MerTK ligands for phagocytosis. *EMBO J* 2010;**29**:3898–910
29. Lemke G, Rothlin CV. Immunobiology of the TAM receptors. *Nat Rev Immunol* 2008;**8**:327–36
30. Ikeda S, He W, Ikeda A, Naggert JK, North MA, Nishina PM. Cell-specific expression of tubby gene family members (tub, Tulp1,2, and 3) in the retina. *Invest Ophthalmol Vis Sci* 1999;**40**:2706–12
31. Nickel W, Seedorf M. Unconventional mechanisms of protein transport to the cell surface of eukaryotic cells. *Annu Rev Cell Dev Biol* 2008;**24**:287–308
32. Caberoy NB, Li W. Unconventional secretion of tubby and tubby-like protein 1. *FEBS Lett* 2009;**583**:3057–62
33. Lemke G. Biology of the TAM receptors. *Cold Spring Harb Perspect Biol* 2013;**5**:a009076
34. Pierce A, Bliesner B, Xu M, Nielsen-Preiss S, Lemke G, Tobet S, Wierman ME. Axl and Tyro3 modulate female reproduction by influencing gonadotropin-releasing hormone neuron survival and migration. *Mol Endocrinol* 2008;**22**:2481–95
35. Lu Q, Lemke G. Homeostatic regulation of the immune system by receptor tyrosine kinases of the Tyro 3 family. *Science* 2001;**293**:306–11
36. Angelillo-Scherrer A, de Frutos P, Aparicio C, Melis E, Savi P, Lupu F, Arnout J, Dewerchin M, Hoylaerts M, Herbert J, Collen D, Dahlback B, Carmeliet P. Deficiency or inhibition of Gas6 causes platelet dysfunction and protects mice against thrombosis. *Nat Med* 2001;**7**:215–21
37. Angelillo-Scherrer A, Burnier L, Flores N, Savi P, DeMol M, Schaeffer P, Herbert JM, Lemke G, Goff SP, Matsushima GK, Earp HS, Vesin C, Hoylaerts MF, Plaisance S, Collen D, Conway EM, Wehrle-Haller B, Carmeliet P. Role of Gas6 receptors in platelet signaling during thrombus stabilization and implications for antithrombotic therapy. *J Clin Invest* 2005;**115**:237–46
38. Fujita K, Chen X, Homma H, Tagawa K, Amano M, Saito A, Imoto S, Akatsu H, Hashizume Y, Kaibuchi K, Miyano S, Okazawa H. Targeting Tyro3 ameliorates a model of PGRN-mutant FTLT-TDP via tau-mediated synaptic pathology. *Nat Commun* 2018;**9**:433
39. Chan PY, Carrera Silva EA, De Kouchkovsky D, Joannas LD, Hao L, Hu D, Huntsman S, Eng C, Licona-Limon P, Weinstein JS, Herbert DR, Craft JE, Flavell RA, Repetto S, Correale J, Burchard EG, Torgerson DG, Ghosh S, Rothlin CV. The TAM family receptor tyrosine kinase TYRO3 is a negative regulator of type 2 immunity. *Science* 2016;**352**:99–103
40. Kim DY, Yu J, Mui RK, Niibori R, Taufique HB, Aslam R, Semple JW, Cordes SP. The tyrosine kinase receptor Tyro3 enhances lifespan and neuropeptide Y (Npy) neuron survival in the mouse anorexia (anx) mutation. *Dis Model Mech* 2017;**10**:581–95
41. Taylor IC, Roy S, Yaswen P, Stampfer MR, Varmus HE. Mouse mammary tumors express elevated levels of RNA encoding the murine homology of SKY, a putative receptor tyrosine kinase. *J Biol Chem* 1995;**270**:6872–80
42. Crosier PS, Hall LR, Vitas MR, Lewis PM, Crosier KE. Identification of a novel receptor tyrosine kinase expressed in acute myeloid leukemic blasts. *Leuk Lymphoma* 1995;**18**:443–9
43. Avilla E, Guarino V, Visciano C, Liotti F, Svelto M, Krishnamoorthy G, Franco R, Melillo RM. Activation of TYRO3/AXL tyrosine kinase receptors in thyroid cancer. *Cancer Res* 2011;**71**:1792–804
44. Shao H, Wang A, Lauffenburger D, Wells A. Tyro3-mediated phosphorylation of ACTN4 at tyrosines is FAK-dependent and decreases susceptibility to cleavage by m-Calpain. *Int J Biochem Cell Biol* 2018;**95**:73–84
45. Kabir TD, Ganda C, Brown RM, Beveridge DJ, Richardson KL, Chaturvedi V, Candy P, Epis M, Wintle L, Kalinowski F, Kopp C, Stuart LM, Yeoh GC, George J, Leedman PJ. A microRNA-7/growth arrest specific 6/TYRO3 axis regulates the growth and invasiveness of sorafenib-resistant cells in human hepatocellular carcinoma. *Hepatology* 2018;**67**:216–31
46. Schmitz R, Valls AF, Yerbes R, von Richter S, Kahlert C, Loges S, Weitz J, Schneider M, de Almodovar CR, Ulrich A, Schmidt T. TAM receptors Tyro3 and Mer as novel targets in colorectal cancer. *Oncotarget* 2016;**7**:56355–70

47. Lee C. Overexpression of Tyro3 receptor tyrosine kinase leads to the acquisition of taxol resistance in ovarian cancer cells. *Mol Med Rep* 2015;**12**:1485–92
48. Chien CW, Hou PC, Wu HC, Chang YL, Lin SC, Lin SC, Lin BW, Lee JC, Chang YJ, Sun HS, Tsai SJ. Targeting TYRO3 inhibits epithelial-mesenchymal transition and increases drug sensitivity in colon cancer. *Oncogene* 2016;**35**:5872–81
49. Demarest SJ, Gardner J, Vendel MC, Ailor E, Szak S, Huang F, Doern A, Tan X, Yang W, Grueneberg DA, Richards EJ, Endege WO, Harlow E, Koopman LA. Evaluation of Tyro3 expression, Gas6-mediated Akt phosphorylation, and the impact of anti-Tyro3 antibodies in melanoma cell lines. *Biochemistry* 2013;**52**:3102–18
50. Dantas-Barbosa C, Lesluyes T, Loarer FL, Chibon F, Treilleux I, Coindre JM, Meeus P, Brahmi M, Bally O, Ray-Coquard I, Sunyach MP, Cesne AL, Mir O, Bonvalot S, Toulmonde M, Italiano A, Saintigny P, Jean-Denis M, Ducimetiere F, Ranchere D, El Sayadi H, Alberti L, Blay JY. Expression and role of TYRO3 and AXL as potential therapeutic targets in leiomyosarcoma. *Br J Cancer* 2017;**117**:1787–97
51. Cheng CL, Hou HA, Lee MC, Liu CY, Jhuang JY, Lai YJ, Lin CW, Chen HY, Liu FT, Chou WC, Chen CY, Tang JL, Yao M, Huang SY, Ko BS, Wu SJ, Tsay W, Tien HF. Higher bone marrow LGALS3 expression is an independent unfavorable prognostic factor for overall survival in patients with acute myeloid leukemia. *Blood* 2013;**121**:3172–80
52. De Vos J, Couderc G, Tarte K, Jourdan M, Requirand G, Delteil MC, Rossi JF, Mechti N, Klein B. Identifying intercellular signaling genes expressed in malignant plasma cells by using complementary DNA arrays. *Blood* 2001;**98**:771–80
53. Wimmel A, Rohner I, Ramaswamy A, Heidtmann HH, Seitz R, Kraus M, Schuermann M. Synthesis and secretion of the anticoagulant protein S and coexpression of the Tyro3 receptor in human lung carcinoma cells. *Cancer* 1999;**86**:43–9
54. Rikova K, Guo A, Zeng Q, Possemato A, Yu J, Haack H, Nardone J, Lee K, Reeves C, Li Y, Hu Y, Tan Z, Stokes M, Sullivan L, Mitchell J, Wetzel R, Macneill J, Ren JM, Yuan J, Bakalarski CE, Villen J, Kornhauser JM, Smith B, Li D, Zhou X, Gygi SP, Gu TL, Polakiewicz RD, Rush J, Comb MJ. Global survey of phosphotyrosine signaling identifies oncogenic kinases in lung cancer. *Cell* 2007;**131**:1190–203
55. Tworowski K, Singhal G, Szpakowski S, Zito CI, Bacchiocchi A, Muthusamy V, Bosenberg M, Krauthammer M, Halaban R, Stern DF. Phosphoproteomic screen identifies potential therapeutic targets in melanoma. *Mol Cancer Res* 2011;**9**:801–12
56. Zhu S, Wurdak H, Wang Y, Galkin A, Tao H, Li J, Lyssiotis CA, Yan F, Tu BP, Miraglia L, Walker J, Sun F, Orth A, Schultz PG, Wu X. A genomic screen identifies TYRO3 as a MITF regulator in melanoma. *Proc Natl Acad Sci U S A* 2009;**106**:17025–30
57. Shiozawa Y, Pedersen EA, Patel LR, Ziegler AM, Havens AM, Jung Y, Wang J, Zalucha S, Loberg RD, Pienta KJ, Taichman RS. GAS6/AXL axis regulates prostate cancer invasion, proliferation, and survival in the bone marrow niche. *Neoplasia* 2010;**12**:116–27
58. Sun WS, Fujimoto J, Tamaya T. Coexpression of growth arrest-specific gene 6 and receptor tyrosine kinases Axl and Sky in human uterine endometrial cancers. *Ann Oncol* 2003;**14**:898–906
59. Ammoun S, Provenzano L, Zhou L, Barczyk M, Evans K, Hilton DA, Hafizi S, Hanemann CO. Axl/Gas6/NFkappaB signalling in schwannoma pathological proliferation, adhesion and survival. *Oncogene* 2014;**33**:336–46
60. Duan Y, Wong W, Chua SC, Wee HL, Lim SG, Chua BT, Ho HK. Overexpression of Tyro3 and its implications on hepatocellular carcinoma progression. *Int J Oncol* 2016;**48**:358–66
61. Qin A, Qian W. MicroRNA-7 inhibits colorectal cancer cell proliferation, migration and invasion via TYRO3 and phosphoinositide 3-kinase/protein B kinase/mammalian target of rapamycin pathway suppression. *Int J Mol Med* 2018;**42**:2503–14
62. Nemoto T, Ohashi K, Akashi T, Johnson JD, Hirokawa K. Overexpression of protein tyrosine kinases in human esophageal cancer. *Pathobiology* 1997;**65**:195–203
63. Thomson S, Petti F, Sujka-Kwok I, Mercado P, Bean J, Monaghan M, Seymour SL, Argast GM, Epstein DM, Haley JD. A systems view of epithelial-mesenchymal transition signaling states. *Clin Exp Metastasis* 2011;**28**:137–55
64. Jansen FH, van Rijswijk A, Teubel W, van Weerden WM, Reneman S, van den Bemd GJ, Roobol MJ, Bangma CH, Staal FJ, Jenster G. Profiling of antibody production against xenograft-released proteins by protein microarrays discovers prostate cancer markers. *J Proteome Res* 2012;**11**:728–35
65. Alexander PB, Chen R, Gong C, Yuan L, Jasper JS, Ding Y, Markowitz GJ, Yang P, Xu X, McDonnell DP, Song E, Wang XF. Distinct receptor tyrosine kinase subsets mediate anti-HER2 drug resistance in breast cancer. *J Biol Chem* 2017;**292**:748–59
66. Kasikara C, Kumar S, Kimani S, Tsou WI, Geng K, Davra V, Sriram G, Devoe C, Nguyen KN, Antes A, Krantz A, Rymarczyk G, Wilczynski A, Empig C, Freimark B, Gray M, Schlunegger K, Hutchins J, Kotenko SV, Birge RB. Phosphatidylserine sensing by TAM receptors regulates AKT-dependent chemoresistance and PD-L1 expression. *Mol Cancer Res* 2017;**15**:753–64
67. Taichman RS, Patel LR, Bedenis R, Wang J, Weidner S, Schumann T, Yumoto K, Berry JE, Shiozawa Y, Pienta KJ. GAS6 receptor status is associated with dormancy and bone metastatic tumor formation. *PLoS One* 2013;**8**:e61873
68. Ammoun S, Cunliffe CH, Allen JC, Chiriboga L, Giancotti FG, Zagzag D, Hanemann CO, Karajannis MA. ErbB/HER receptor activation and preclinical efficacy of lapatinib in vestibular schwannoma. *Neurooncology* 2010;**12**:834–43
69. Sinha S, Boysen J, Nelson M, Secreto C, Warner SL, Bearss DJ, Lesnick C, Shanafelt TD, Kay NE, Ghosh AK. Targeted Axl inhibition primes chronic lymphocytic leukemia B cells to apoptosis and shows synergistic/additive effects in combination with BTK inhibitors. *Clin Cancer Res* 2015;**21**:2115–26
70. Neubauer A, Burchert A, Maiwald C, Gruss HJ, Serke S, Huhn D, Wittig B, Liu E. Recent progress on the role of Axl, a receptor tyrosine kinase, in malignant transformation of myeloid leukemias. *Leuk Lymphoma* 1997;**25**:91–6
71. Ghosh AK, Secreto C, Boysen J, Sassoon T, Shanafelt TD, Mukhopadhyay D, Kay NE. The novel receptor tyrosine kinase Axl is constitutively active in B-cell chronic lymphocytic leukemia and acts as a docking site of nonreceptor kinases: implications for therapy. *Blood* 2011;**117**:1928–37
72. Wimmel A, Glitz D, Kraus A, Roeder J, Schuermann M. Axl receptor tyrosine kinase expression in human lung cancer cell lines correlates with cellular adhesion. *Eur J Cancer* 2001;**37**:2264–74
73. Keating AK, Kim GK, Jones AE, Donson AM, Ware K, Mulcahy JM, Salzberg DB, Foreman NK, Liang X, Thorburn A, Graham DK. Inhibition of Mer and Axl receptor tyrosine kinases in astrocytoma cells leads to increased apoptosis and improved chemosensitivity. *Mol Cancer Ther* 2010;**9**:1298–307
74. Zhang YX, Knyazev PG, Cheburkin YV, Sharma K, Knyazev YP, Orfi L, Szabadkai I, Daub H, Keri G, Ullrich A. AXL is a potential target for therapeutic intervention in breast cancer progression. *Cancer Res* 2008;**68**:1905–15
75. Craven RJ, Xu LH, Weiner TM, Fridell YW, Dent GA, Srivastava S, Varnum B, Liu ET, Cance WG. Receptor tyrosine kinases expressed in metastatic colon cancer. *Int J Cancer* 1995;**60**:791–7
76. Mudduluru G, Vajkoczy P, Allgayer H. Myeloid zinc finger 1 induces migration, invasion, and in vivo metastasis through Axl gene expression in solid cancer. *Mol Cancer Res* 2010;**8**:159–69
77. Lin WC, Li AF, Chi CW, Chung WW, Huang CL, Lui WY, Kung HJ, Wu CW. tie-1 protein tyrosine kinase: a novel independent prognostic marker for gastric cancer. *Clin Cancer Res* 1999;**5**:1745–51
78. Sawabu T, Seno H, Kawashima T, Fukuda A, Uenoyama Y, Kawada M, Kanda N, Sekikawa A, Fukui H, Yanagita M, Yoshibayashi H, Satoh S, Sakai Y, Nakano T, Chiba T. Growth arrest-specific gene 6 and Axl signaling enhances gastric cancer cell survival via Akt pathway. *Mol Carcinog* 2007;**46**:155–64
79. Song X, Wang H, Logsdon CD, Rashid A, Fleming JB, Abbruzzese JL, Gomez HF, Evans DB, Wang H. Overexpression of receptor tyrosine kinase Axl promotes tumor cell invasion and survival in pancreatic ductal adenocarcinoma. *Cancer* 2011;**117**:734–43

80. Leconet W, Larbouret C, Chardes T, Thomas G, Neiveyans M, Busson M, Jarlier M, Radosevic-Robin N, Pugniere M, Bernex F, Penault-Llorca F, Pasquet JM, Pelegriin A, Robert B. Preclinical validation of AXL receptor as a target for antibody-based pancreatic cancer immunotherapy. *Oncogene* 2014;**33**:5405–14
81. Hector A, Montgomery EA, Karikari C, Canto M, Dunbar KB, Wang JS, Feldmann G, Hong SM, Haffner MC, Meeker AK, Holland SJ, Yu J, Heckrodt TJ, Zhang J, Ding P, Goff D, Singh R, Roa JC, Marimuthu A, Riggins GJ, Eshleman JR, Nelkin BD, Pandey A, Maitra A. The Axl receptor tyrosine kinase is an adverse prognostic factor and a therapeutic target in esophageal adenocarcinoma. *Cancer Biol Ther* 2010;**10**:1009–18
82. van Ginkel PR, Gee RL, Shearer RL, Subramanian L, Walker TM, Albert DM, Meisner LF, Varnum BC, Polans AS. Expression of the receptor tyrosine kinase Axl promotes ocular melanoma cell survival. *Cancer Res* 2004;**64**:128–34
83. Tworowski KA, Platt JT, Bacchiocchi A, Bosenberg M, Boggon TJ, Stern DF. MERTK controls melanoma cell migration and survival and differentially regulates cell behavior relative to AXL. *Pigment Cell Melanoma Res* 2013;**26**:527–41
84. Green J, Ikram M, Vyas J, Patel N, Proby CM, Ghali L, Leigh IM, O'Toole EA, Storey A. Overexpression of the Axl tyrosine kinase receptor in cutaneous SCC-derived cell lines and tumours. *Br J Cancer* 2006;**94**:1446–51
85. Pacez JD, Vasques GJ, Correa RG, Vasconcellos JF, Duncan K, Gu X, Bhasin M, Libermann TA, Zerbin LF. The receptor tyrosine kinase Axl is an essential regulator of prostate cancer proliferation and tumor growth and represents a new therapeutic target. *Oncogene* 2013;**32**:689–98
86. Sun W, Fujimoto J, Tamaya T. Coexpression of Gas6/Axl in human ovarian cancers. *Oncology* 2004;**66**:450–7
87. Chen PX, Li QY, Yang Z. Axl and prostaticin are biomarkers for prognosis of ovarian adenocarcinoma. *Ann Diagn Pathol* 2013;**17**:425–9
88. Rankin EB, Fuh KC, Taylor TE, Krieg AJ, Musser M, Yuan J, Wei K, Kuo CJ, Longacre TA, Giaccia AJ. AXL is an essential factor and therapeutic target for metastatic ovarian cancer. *Cancer Res* 2010;**70**:7570–9
89. Lee CH, Yen CY, Liu SY, Chen CK, Chiang CF, Shiah SG, Chen PH, Shieh YS. Axl is a prognostic marker in oral squamous cell carcinoma. *Ann Surg Oncol* 2012;**19**:S500–8
90. Yeh CY, Shin SM, Yeh HH, Wu TJ, Shin JW, Chang TY, Raghavaraju G, Lee CT, Chiang JH, Tseng VS, Lee YC, Shen CH, Chow NH, Liu HS. Transcriptional activation of the Axl and PDGFR- α by c-Met through a ras- and Src-independent mechanism in human bladder cancer. *BMC Cancer* 2011;**11**:139
91. Gustafsson A, Martuszewska D, Johansson M, Ekman C, Hafizi S, Ljungberg B, Dahlback B. Differential expression of Axl and Gas6 in renal cell carcinoma reflecting tumor advancement and survival. *Clin Cancer Res* 2009;**15**:4742–9
92. Ou WB, Corson JM, Flynn DL, Lu WP, Wise SC, Bueno R, Sugarbaker DJ, Fletcher JA. AXL regulates mesothelioma proliferation and invasiveness. *Oncogene* 2011;**30**:1643–52
93. Liu R, Gong M, Li X, Zhou Y, Gao W, Tulpule A, Chaudhary PM, Jung J, Gill PS. Induction, regulation, and biologic function of Axl receptor tyrosine kinase in Kaposi sarcoma. *Blood* 2010;**116**:297–305
94. Ishikawa M, Sonobe M, Nakayama E, Kobayashi M, Kikuchi R, Kitamura J, Imamura N, Date H. Higher expression of receptor tyrosine kinase Axl, and differential expression of its ligand, Gas6, predict poor survival in lung adenocarcinoma patients. *Ann Surg Oncol* 2013;**20**:S467–76
95. Giles KM, Kalinowski FC, Candy PA, Epis MR, Zhang PM, Redfern AD, Stuart LM, Goodall GJ, Leedman PJ. Axl mediates acquired resistance of head and neck cancer cells to the epidermal growth factor receptor inhibitor erlotinib. *Mol Cancer Ther* 2013;**12**:2541–58
96. Han J, Tian R, Yong B, Luo C, Tan P, Shen J, Peng T. Gas6/Axl mediates tumor cell apoptosis, migration and invasion and predicts the clinical outcome of osteosarcoma patients. *Biochem Biophys Res Commun* 2013;**435**:493–500
97. Wu CW, Li AF, Chi CW, Lai CH, Huang CL, Lo SS, Lui WY, Lin WC. Clinical significance of AXL kinase family in gastric cancer. *Anticancer Res* 2002;**22**:1071–8
98. Jiao Y, Ou W, Meng F, Zhou H, Wang A. Targeting HSP90 in ovarian cancers with multiple receptor tyrosine kinase coactivation. *Mol Cancer* 2011;**10**:125
99. Zhao Y, Sun X, Jiang L, Yang F, Zhang Z, Jia L. Differential expression of Axl and correlation with invasion and multidrug resistance in cancer cells. *Cancer Invest* 2012;**30**:287–94
100. Li Y, Ye X, Tan C, Hongo JA, Zha J, Liu J, Kallop D, Ludlam MJ, Pei L. Axl as a potential therapeutic target in cancer: role of Axl in tumor growth, metastasis and angiogenesis. *Oncogene* 2009;**28**:3442–55
101. He L, Zhang J, Jiang L, Jin C, Zhao Y, Yang G, Jia L. Differential expression of Axl in hepatocellular carcinoma and correlation with tumor lymphatic metastasis. *Mol Carcinog* 2010;**49**:882–91
102. Rettew AN, Young ED, Lev DC, Kleiner ES, Abdul-Karim FW, Getty PJ, Greenfield EM. Multiple receptor tyrosine kinases promote the in vitro phenotype of metastatic human osteosarcoma cell lines. *Oncogenesis* 2012;**1**:e34
103. Holland SJ, Powell MJ, Franci C, Chan EW, Frier AM, Atchison RE, McLaughlin J, Swift SE, Pali ES, Yam G, Wong S, Lasaga J, Shen MR, Yu S, Xu W, Hitoshi Y, Bogenberger J, Nor JE, Payan DG, Lorens JB. Multiple roles for the receptor tyrosine kinase axl in tumor formation. *Cancer Res* 2005;**65**:9294–303
104. Mishra A, Wang J, Shiozawa Y, McGee S, Kim J, Jung Y, Joseph J, Berry JE, Havens A, Pienta KJ, Taichman RS. Hypoxia stabilizes GAS6/Axl signaling in metastatic prostate cancer. *Mol Cancer Res* 2012;**10**:703–12
105. Sayan AE, Stanford R, Vickery R, Grigorenko E, Diesch J, Kulbicki K, Edwards R, Pal R, Greaves P, Jariel-Encontre I, Piechaczyk M, Kriajevska M, Mellon JK, Dhillon AS, Tulchinsky E. Fra-1 controls motility of bladder cancer cells via transcriptional upregulation of the receptor tyrosine kinase AXL. *Oncogene* 2012;**31**:1493–503
106. Duijkers FA, Meijerink JP, Pieters R, van Noesel MM. Downregulation of Axl in non-MYC amplified neuroblastoma cell lines reduces migration. *Gene* 2013;**521**:62–8
107. Hong CC, Lay JD, Huang JS, Cheng AL, Tang JL, Lin MT, Lai GM, Chuang SE. Receptor tyrosine kinase AXL is induced by chemotherapy drugs and overexpression of AXL confers drug resistance in acute myeloid leukemia. *Cancer Lett* 2008;**268**:314–24
108. Lay JD, Hong CC, Huang JS, Yang YY, Pao CY, Liu CH, Lai YP, Lai GM, Cheng AL, Su IJ, Chuang SE. Sulfasalazine suppresses drug resistance and invasiveness of lung adenocarcinoma cells expressing AXL. *Cancer Res* 2007;**67**:3878–87
109. Hong J, Peng D, Chen Z, Sehdev V, Belkhir A. ABL regulation by AXL promotes cisplatin resistance in esophageal cancer. *Cancer Res* 2013;**73**:331–40
110. Linger RM, Cohen RA, Cummings CT, Sather S, Migdall-Wilson J, Middleton DH, Lu X, Baron AE, Franklin WA, Merrick DT, Jedlicka P, DeRyckere D, Heasley LE, Graham DK. Mer or Axl receptor tyrosine kinase inhibition promotes apoptosis, blocks growth and enhances chemosensitivity of human non-small cell lung cancer. *Oncogene* 2013;**32**:3420–31
111. Lee-Sherick AB, Eisenman KM, Sather S, McGranahan A, Armistead PM, McGary CS, Hunsucker SA, Schlegel J, Martinson H, Cannon C, Keating AK, Earp HS, Liang X, DeRyckere D, Graham DK. Aberrant Mer receptor tyrosine kinase expression contributes to leukemogenesis in acute myeloid leukemia. *Oncogene* 2013;**32**:5359–68
112. Graham DK, Dawson TL, Mullaney DL, Snodgrass HR, Earp HS. Cloning and mRNA expression analysis of a novel human protooncogene, c-mer. *Cell Growth Different* 1994;**5**:647–57
113. Brandao LN, Wings A, Christoph S, Sather S, Migdall-Wilson J, Schlegel J, McGranahan A, Gao D, Liang X, Deryckere D, Graham DK. Inhibition of MerTK increases chemosensitivity and decreases oncogenic potential in T-cell acute lymphoblastic leukemia. *Blood Cancer J* 2013;**3**:e101
114. Wang Y, Moncayo G, Morin P, Jr., Xue G, Grzmil M, Lino MM, Clement-Schatlo V, Frank S, Merlo A, Hemmings BA. Mer receptor tyrosine kinase promotes invasion and survival in glioblastoma multiforme. *Oncogene* 2013;**32**:872–82

115. Schlegel J, Sambade MJ, Sather S, Moschos SJ, Tan AC, Wings A, DeRyckere D, Carson CC, Trembath DG, Tentler JJ, Eckhardt SG, Kuan PF, Hamilton RL, Duncan LM, Miller CR, Nikolaishvili-Feinberg N, Midkiff BR, Liu J, Zhang W, Yang C, Wang X, Frye SV, Earp HS, Shields JM, Graham DK. MERTK receptor tyrosine kinase is a therapeutic target in melanoma. *J Clin Invest* 2013;**123**:2257–67
116. Wu YM, Robinson DR, Kung HJ. Signal pathways in up-regulation of chemokines by tyrosine kinase MER/NYK in prostate cancer cells. *Cancer Res* 2004;**64**:7311–20
117. Ek S, Hogerkorp CM, Dictor M, Ehinger M. Borrebaeck C A Mantle cell lymphomas express a distinct genetic signature affecting lymphocyte trafficking and growth regulation as compared with subpopulations of normal human B cells. *Cancer Res* 2002;**62**:4398–405
118. Knubel KH, Pernu BM, Sufti A, Nelson S, Pierce AM, Keating AK. MerTK inhibition is a novel therapeutic approach for glioblastoma multiforme. *Oncotarget* 2014;**5**:1338–51
119. Dirks W, Rome D, Ringel F, Jager K, MacLeod RA, Drexler HG. Expression of the growth arrest-specific gene 6 (GAS6) in leukemia and lymphoma cell lines. *Leuk Res* 1999;**23**:643–51
120. Hutterer M, Knyazev P, Abate A, Reschke M, Maier H, Stefanova N, Knyazeva T, Barbieri V, Reindl M, Muigg A, Kostron H, Stockhammer G, Ullrich A. Axl and growth arrest-specific gene 6 are frequently overexpressed in human gliomas and predict poor prognosis in patients with glioblastoma multiforme. *Clin Cancer Res* 2008;**14**:130–8
121. McCormack O, Chung WY, Fitzpatrick P, Cooke F, Flynn B, Harrison M, Fox E, Gallagher E, Goldrick AM, Dervan PA, McCann A, Kerin MJ. Growth arrest-specific gene 6 expression in human breast cancer. *Br J Cancer* 2008;**98**:1141–6
122. Buehler M, Tse B, Leboucq A, Jacob F, Caduff R, Fink D, Goldstein DR, Heinzelmann-Schwarz V. Meta-analysis of microarray data identifies GAS6 expression as an independent predictor of poor survival in ovarian cancer. *Biomed Res Int* 2013;**2013**:238284
123. Ito M, Nakashima M, Nakayama T, Ohtsuru A, Nagayama Y, Takamura N, Demedchik EP, Sekine I, Yamashita S. Expression of receptor-type tyrosine kinase, Axl, and its ligand, Gas6, in pediatric thyroid carcinomas around chernobyl. *Thyroid* 2002;**12**:971–5
124. Whitman SP, Kohlschmidt J, Maharry K, Volinia S, Mrozek K, Nicolet D, Schwind S, Becker H, Metzeler KH, Mendler JH, Eisfeld AK, Carroll AJ, Powell BL, Carter TH, Baer MR, Kolitz JE, Park IK, Stone RM, Caligiuri MA, Marcucci G, Bloomfield CD. GAS6 expression identifies high-risk adult AML patients: potential implications for therapy. *Leukemia* 2014;**28**:1252–8
125. Loges S, Schmidt T, Tjwa M, van Geyte K, Lievens D, Lutgens E, Vanhoutte D, Borgel D, Plaisance S, Hoylaerts M, Luttun A, Dewerchin M, Jonckx B, Carmeliet P. Malignant cells fuel tumor growth by educating infiltrating leukocytes to produce the mitogen Gas6. *Blood* 2010;**115**:2264–73
126. Sainaghi PP, Castello L, Bergamasco L, Galletti M, Bellosta P, Avanzi GC. Gas6 induces proliferation in prostate carcinoma cell lines expressing the Axl receptor. *J Cell Physiol* 2005;**204**:36–44
127. Gustafsson A, Bostrom AK, Ljungberg B, Axelson H, Dahlback B. Gas6 and the receptor tyrosine kinase Axl in clear cell renal cell carcinoma. *PLoS One* 2009;**4**:e7575
128. Lee Y, Lee M, Kim S. Gas6 induces cancer cell migration and epithelial-mesenchymal transition through upregulation of MAPK and Slug. *Biochem Biophys Res Commun* 2013;**434**:8–14
129. Shiozawa Y, Pedersen EA, Taichman RS. GAS6/Mer axis regulates the homing and survival of the E2A/PBX1-positive B-cell precursor acute lymphoblastic leukemia in the bone marrow niche. *Exp Hematol* 2010;**38**:132–40
130. Abe A, Emi N, Kato H, Adachi K, Murate T, Saga S, Ogura M, Kojima T, Tanimoto M, Morishita N. Establishment and characterization of an immature human megakaryoblastic cell line, MEG-A2. *Leukemia* 1995;**9**:341–9
131. Chung KW, Kim SW, Kim SW. Gene expression profiling of papillary thyroid carcinomas in Korean patients by oligonucleotide microarrays. *J Korean Surg Soc* 2012;**82**:271–80
132. Griffith OL, Melck A, Jones SJ, Wiseman SM. Meta-analysis and meta-review of thyroid cancer gene expression profiling studies identifies important diagnostic biomarkers. *J Clin Oncol* 2006;**24**:5043–51
133. Sierko E, Wojtukiewicz MZ, Zawadzki R, Zimnoch L, Kisiel W. Expression of protein C (PC), protein S (PS) and thrombomodulin (TM) in human colorectal cancer. *Thromb Res* 2010;**125**:e71–5
134. Putterman C, Tadmor B, Simer L, Caraco Y. Recurrent venous thrombosis as the presenting sign of two primary lung carcinomas – 15 years apart. *Respir Med* 1992;**86**:521–3
135. Phillips DJ, Greengard JS, Fernandez JA, Ribeiro M, Evatt BL, Griffin JH, Hooper WC. Protein S, an antithrombotic factor, is synthesized and released by neural tumor cells. *J Neurochem* 1993;**61**:344–7
136. Wojtukiewicz MZ, Zacharski LR, Memoli VA, Kisiel W, Kudryk BJ, Rousseau SM, Stump DC. Abnormal regulation of coagulation/fibrinolysis in small cell carcinoma of the lung. *Cancer* 1990;**65**:481–5
137. Saraon P, Musrap N, Cretu D, Karagiannis GS, Batruch I, Smith C, Drabovich AP, Trudel D, van der Kwast T, Morrissey C, Jarvi KA, Diamandis EP. Proteomic profiling of androgen-independent prostate cancer cell lines reveals a role for protein S during the development of high grade and castration-resistant prostate cancer. *J Biol Chem* 2012;**287**:34019–31
138. Faca VM, Ventura AP, Fitzgibbon MP, Pereira-Faca SR, Pitteri SJ, Green AE, Ireton RC, Zhang Q, Wang H, O'Brian KC, Drescher CW, Schummer M, McIntosh MW, Knudsen BS, Hanash SM. Proteomic analysis of ovarian cancer cells reveals dynamic processes of protein secretion and shedding of extra-cellular domains. *PLoS One* 2008;**3**:e2425
139. Maillard C, Berruyer M, Serre CM, Dechavanne M, Delmas PD. Protein-S, a vitamin K-dependent protein, is a bone matrix component synthesized and secreted by osteoblasts. *Endocrinology* 1992;**130**:1599–604
140. Che Mat MF, Abdul Murad NA, Ibrahim K, Mohd Mokhtar N, Wan Ngah WZ, Harun R, Jamal R. Silencing of PROS1 induces apoptosis and inhibits migration and invasion of glioblastoma multiforme cells. *Int J Oncol* 2016;**49**:2359–66
141. Ning P, Zhong JG, Jiang F, Zhang Y, Zhao J, Tian F, Li W. Role of protein S in castration-resistant prostate cancer-like cells. *Endocrine Relat Cancer* 2016;**23**:595–607
142. Ekyalongo RC, Mukohara T, Funakoshi Y, Tomioka H, Kataoka Y, Shimono Y, Chayahara N, Toyoda M, Kiyota N, Minami H. TYRO3 as a potential therapeutic target in breast cancer. *Anticancer Res* 2014;**34**:3337–45
143. Chung JH, Rho JK, Xu X, Lee JS, Yoon HI, Lee CT, Choi YJ, Kim HR, Kim CH, Lee JC. Clinical and molecular evidences of epithelial to mesenchymal transition in acquired resistance to EGFR-TKIs. *Lung Cancer* 2011;**73**:176–82
144. Sequist LV, Waltman BA, Dias-Santagata D, Digumarthy S, Turke AB, Fidias P, Bergethon K, Shaw AT, Gettinger S, Cosper AK, Akhavanfar S, Heist RS, Temel J, Christensen JG, Wain JC, Lynch TJ, Vernovsky K, Mark EJ, Lanuti M, Iafrate AJ, Mino-Kenudson M, Engelman JA. Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. *Sci Transl Med* 2011;**3**:75ra26
145. Liu Y, Zeng G. Cancer and innate immune system interactions: translational potentials for cancer immunotherapy. *J Immunother* 2012;**35**:299–308
146. Tushima J, Ohashi K, Iwashita S, Mizuno K. Autophosphorylation activity and association with Src family kinase of Sky receptor tyrosine kinase. *Biochem Biophys Res Commun* 1995;**209**:656–63
147. Lan Z, Wu H, Li W, Wu S, Lu L, Xu M, Dai W. Transforming activity of receptor tyrosine kinase tyro3 is mediated, at least in part, by the PI3 kinase-signaling pathway. *Blood* 2000;**95**:633–8
148. Hafizi S, Gustafsson A, Stenhoff J, Dahlback B. The Ran binding protein RanBPM interacts with Axl and Sky receptor tyrosine kinases. *Int J Biochem Cell Biol* 2005;**37**:2344–56
149. Guo H, Barrett TM, Zhong Z, Fernandez JA, Griffin JH, Freeman RS, Zlokovic BV. Protein S blocks the extrinsic apoptotic cascade in tissue plasminogen activator/N-methyl D-aspartate-treated neurons via Tyro3-Akt-FKHL1 signaling pathway. *Mol Neurodegener* 2011;**6**:13

150. Zhong Z, Wang Y, Guo H, Sagare A, Fernandez JA, Bell RD, Barrett TM, Griffin JH, Freeman RS, Zlokovic BV. Protein S protects neurons from excitotoxic injury by activating the TAM receptor Tyro3-phosphatidylinositol 3-kinase-Akt pathway through its sex hormone-binding globulin-like region. *J Neurosci* 2010;**30**:15521–34
151. Chen J, Carey K, Godowski PJ. Identification of Gas6 as a ligand for Mer, a neural cell adhesion molecule related receptor tyrosine kinase implicated in cellular transformation. *Oncogene* 1997;**14**:2033–9
152. Katagiri M, Hakeda Y, Chikazu D, Ogasawara T, Takato T, Kumegawa M, Nakamura K, Kawaguchi H. Mechanism of stimulation of osteoclastic bone resorption through Gas6/Tyro 3, a receptor tyrosine kinase signaling, in mouse osteoclasts. *J Biol Chem* 2001;**276**:7376–82
153. Ekyalongo RC, Mukohara T, Kataoka Y, Funakoshi Y, Tomioka H, Kiyota N, Fujiwara Y, Minami H. Mechanisms of acquired resistance to insulin-like growth factor 1 receptor inhibitor in MCF-7 breast cancer cell line. *Invest New Drugs* 2013;**31**:293–303
154. Irie H, Banno K, Yanokura M, Iida M, Adachi M, Nakamura K, Umene K, Nogami Y, Masuda K, Kobayashi Y, Tominaga E, Aoki D. Metformin: a candidate for the treatment of gynecological tumors based on drug repositioning. *Oncol Lett* 2016;**11**:1287–93
155. Kim NY, Lee HY, Lee C. Metformin targets Axl and Tyro3 receptor tyrosine kinases to inhibit cell proliferation and overcome chemoresistance in ovarian cancer cells. *Int J Oncol* 2015;**47**:353–60
156. Powell NA, Hoffman JK, Ciske FL, Kohrt JT, Baxi SM, Peng YW, Zhong M, Catana C, Ohren J, Perrin LA, Edmunds JJ. Optimization of highly selective 2,4-diaminopyrimidine-5-carboxamide inhibitors of Sky kinase. *Bioorgan Med Chem Lett* 2013;**23**:1051–5
157. Paolino M, Choidas A, Wallner S, Pranjic B, Uribealago I, Loeser S, Jamieson AM, Langdon WY, Ikeda F, Fededa JP, Cronin SJ, Nitsch R, Schultz-Fademrecht C, Eickhoff J, Menninger S, Unger A, Torka R, Gruber T, Hinterleitner R, Baier G, Wolf D, Ullrich A, Klebl BM, Penninger JM. The E3 ligase Cbl-b and TAM receptors regulate cancer metastasis via natural killer cells. *Nature* 2014;**507**:508–12
158. Schroeder GM, An Y, Cai ZW, Chen XT, Clark C, Cornelius LA, Dai J, Gullo-Brown J, Gupta A, Henley B, Hunt JT, Jeyaseelan R, Kamath A, Kim K, Lippy J, Lombardo LJ, Manne V, Oppenheimer S, Sack JS, Schmidt RJ, Shen G, Stefanski K, Tokarski JS, Trainor GL, Wautlet BS, Wei D, Williams DK, Zhang Y, Zhang Y, Fargnoli J, Borzilleri RM. Discovery of N-(4-(2-amino-3-chloropyridin-4-yloxy)-3-fluorophenyl)-4-ethoxy-1-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carboxamide (BMS-777607), a selective and orally efficacious inhibitor of the Met kinase superfamily. *J Med Chem* 2009;**52**:1251–4
159. Dai Y, Siemann DW. BMS-777607, a small-molecule met kinase inhibitor, suppresses hepatocyte growth factor-stimulated prostate cancer metastatic phenotype in vitro. *Mol Cancer Ther* 2010;**9**:1554–61
160. Sharma S, Yao HP, Zhou YQ, Zhou J, Zhang R, Wang MH. Prevention of BMS-777607-induced polyploidy/senescence by mTOR inhibitor AZD8055 sensitizes breast cancer cells to cytotoxic chemotherapeutics. *Mol Oncol* 2014;**8**:469–82
161. Patel PR, Sun H, Li SQ, Shen M, Khan J, Thomas CJ, Davis MI. Identification of potent Yes1 kinase inhibitors using a library screening approach. *Bioorg Med Chem Lett* 2013;**23**:4398–403
162. Dai Y, Siemann DW. Constitutively active c-Met kinase in PC-3 cells is autocrine-independent and can be blocked by the Met kinase inhibitor BMS-777607. *BMC Cancer* 2012;**12**:198
163. Dai Y, Bae K, Pampo C, Siemann DW. Impact of the small molecule Met inhibitor BMS-777607 on the metastatic process in a rodent tumor model with constitutive c-Met activation. *Clin Exp Metastasis* 2012;**29**:253–61
164. Mollard A, Warner SL, Call LT, Wade ML, Bearss JJ, Verma A, Sharma S, Vankayalapati H, Bearss DJ. Design, synthesis and biological evaluation of a series of novel Axl kinase inhibitors. *ACS Med Chem Lett* 2011;**2**:907–12
165. Zhang W, Zhang D, Stashko MA, DeRyckere D, Hunter D, Kireev D, Miley MJ, Cummings C, Lee M, Norris-Drouin J, Stewart WM, Sather S, Zhou Y, Kirkpatrick G, Machius M, Janzen WP, Earp HS, Graham DK, Frye SV, Wang X. Pseudo-cyclization through intramolecular hydrogen bond enables discovery of pyridine substituted pyrimidines as new Mer kinase inhibitors. *J Med Chem* 2013;**56**:9683–92
166. Shi C, Li X, Wang X, Ding N, Ping L, Shi Y, Mi L, Lai Y, Song Y, Zhu J. The proto-oncogene Mer tyrosine kinase is a novel therapeutic target in mantle cell lymphoma. *J Hematol Oncol* 2018;**11**:43
167. Zhang W, McIver AL, Stashko MA, DeRyckere D, Branchford BR, Hunter D, Kireev D, Miley MJ, Norris-Drouin J, Stewart WM, Lee M, Sather S, Zhou Y, Di Paola JA, Machius M, Janzen WP, Earp HS, Graham DK, Frye SV, Wang X. Discovery of Mer specific tyrosine kinase inhibitors for the treatment and prevention of thrombosis. *J Med Chem* 2013;**56**:9693–700
168. Kim JE, Kim Y, Li G, Kim ST, Kim K, Park SH, Park JO, Park YS, Lim HY, Lee H, Sohn TS, Kim KM, Kang WK, Lee J. MerTK inhibition by RXDX-106 in MerTK activated gastric cancer cell lines. *Oncotarget* 2017;**8**:105727–34
169. Suarez RM, Chevot F, Cavagnino A, Saettel N, Radvanyi F, Piguel S, Bernard-Pierrot I, Stoven V, Legraverend M. Inhibitors of the TAM subfamily of tyrosine kinases: synthesis and biological evaluation. *Eur J Med Chem* 2013;**61**:2–25
170. Patwardhan PP, Ivy KS, Musi E, de Stanchina E, Schwartz GK. Significant blockade of multiple receptor tyrosine kinases by MGCD516 (Sitravatinib), a novel small molecule inhibitor, shows potent anti-tumor activity in preclinical models of sarcoma. *Oncotarget* 2016;**7**:4093–109
171. Du W, Huang H, Sorrelle N, Brekken R A. Sitravatinib potentiates immune checkpoint blockade in refractory cancer models. *JCI Insight* 2018;**3**:pii: 124184
172. Zhao J, Zhang D, Zhang W, Stashko MA, DeRyckere D, Vasileiadis E, Parker RE, Hunter D, Liu Q, Zhang Y, Norris-Drouin J, Li B, Drewry DH, Kireev D, Graham DK, Earp HS, Frye SV, Wang X. Highly selective MERTK inhibitors achieved by a single methyl group. *J Med Chem* 2018;**61**:10242–54
173. Donato NJ. Imatinib is receptive to a collaboration. *Blood* 2011;**118**:4009–10
174. Gottlieb WH, Saumet J, Beauchamp MC, Gu J, Lau S, Pollak MN, Bruchim I. In vitro metformin anti-neoplastic activity in epithelial ovarian cancer. *Gynecol Oncol* 2008;**110**:246–50
175. Wang LW, Li ZS, Zou DW, Jin ZD, Gao J, Xu GM. Metformin induces apoptosis of pancreatic cancer cells. *World J Gastroenterol* 2008;**14**:7192–8
176. Goodwin PJ, Pritchard KI, Ennis M, Clemons M, Graham M, Fantus IG. Insulin-lowering effects of metformin in women with early breast cancer. *Clin Breast Cancer* 2008;**8**:501–5
177. Ben Sahra I, Laurent K, Loubat A, Giorgetti-Peraldi S, Colosetti P, Auberger P, Tanti JF, Le Marchand-Brustel Y, Bost F. The antidiabetic drug metformin exerts an antitumoral effect in vitro and in vivo through a decrease of cyclin D1 level. *Oncogene* 2008;**27**:3576–86
178. Hadad SM, Appleyard V, Thompson AM. Therapeutic metformin/AMPK activation promotes the angiogenic phenotype in the ERalpha negative MDA-MB-435 breast cancer model. *Breast Cancer Res Treat* 2009;**114**:391
179. Nair V, Sreevalsan S, Basha R, Abdelrahman M, Abudayyeh A, Rodrigues Hoffman A, Safe S. Mechanism of metformin-dependent inhibition of mammalian target of rapamycin (mTOR) and Ras activity in pancreatic cancer: role of specificity protein (Sp) transcription factors. *J Biol Chem* 2014;**289**:27692–701
180. Honjo S, Ajani JA, Scott AW, Chen Q, Skinner HD, Stroehlein J, Johnson RL, Song S. Metformin sensitizes chemotherapy by targeting cancer stem cells and the mTOR pathway in esophageal cancer. *Int J Oncol* 2014;**45**:567–74
181. Ben Sahra I, Regazzetti C, Robert G, Laurent K, Le Marchand-Brustel Y, Auberger P, Tanti JF, Giorgetti-Peraldi S, Bost F. Metformin, independent of AMPK, induces mTOR inhibition and cell-cycle arrest through REDD1. *Cancer Res* 2011;**71**:4366–72
182. Powell NA, Kohrt JT, Filipinski KJ, Kaufman M, Sheehan D, Edmunds JE, Delaney A, Wang Y, Bourbonais F, Lee DY, Schwende F, Sun F, McConnell P, Catana C, Chen H, Ohren J, Perrin LA. Novel and

- selective spiroindoline-based inhibitors of Sky kinase. *Bioorgan Med Chem Lett* 2012;**22**:190–3
183. Zhu H, Naujokas MA, Fixman ED, Torossian K, Park M. Tyrosine 1356 in the carboxyl-terminal tail of the HGF/SF receptor is essential for the transduction of signals for cell motility and morphogenesis. *J Biol Chem* 1994;**269**:29943–8
184. Birchmeier C, Birchmeier W, Gherardi E, Vande Woude GF. Met, metastasis, motility and more. *Nat Rev Mol Cell Biol* 2003;**4**:915–25
185. Comoglio PM, Giordano S, Trusolino L. Drug development of MET inhibitors: targeting oncogene addiction and expedience. *Nat Rev Drug Discov* 2008;**7**:504–16