# **Minireview**

# Nonalcoholic fatty liver disease and the gut microbiome: Are bacteria responsible for fatty liver?

## Tien S Dong<sup>1,2</sup> and Jonathan P Jacobs<sup>1,2,3</sup>

<sup>1</sup>Division of Gastroenterology, Hepatology and Parenteral Nutrition, VA Greater Los Angeles Healthcare System, Los Angeles, CA 90025, USA; <sup>2</sup>The Vatche and Tamar Manoukian Division of Digestive Diseases, Department of Medicine, David Geffen School of Medicine at University of California Los Angeles, Los Angeles, CA 90095, USA; <sup>3</sup>University of California Los Angeles Microbiome Center, David Geffen School of Medicine, Los Angeles, CA 90095, USA

Corresponding author: Jonathan P Jacobs. Email: JJacobs@mednet.ucla.edu

#### Impact statement

This invited minireview for the upcoming thematic issue on the microbiome addresses the role of the microbiome in nonalcoholic fatty liver disease (NAFLD). The incidence of NAFLD has increased greatly in recent years in parallel with the rise in obesity and is now believed to have a population prevalence of 20-40%. It is anticipated to soon become the primary cause of liver-related morbidity and mortality, and unfortunately, there are few treatment options. Therefore, there is a critical need for improved understanding of NAFLD pathophysiology to provide new avenues for therapeutic intervention. In this paper, we have reviewed evidence from human and animal model studies that have associated microbiome composition and microbial metabolites with development and progression of NAFLD. We have also discussed proposed mechanisms by which the microbiome could contribute to NAFLD pathogenesis and addressed future directions for this field.

#### **Abstract**

Over the last several years, a growing body of literature has linked the gut microbiome to human health and diseases such as obesity, metabolic syndrome, and nonalcoholic fatty liver disease (NAFLD). This paper will review the current literature investigating the influence of diets associated with metabolic disorders on the microbiome and how those changes promote susceptibility to metabolic disorders. It will then focus in-depth on the role of the gut microbiome in NAFLD. The review will highlight associations of microbial composition and function with progression of NAFLD in patients and discuss potential mechanisms that link the gut microbiome to NAFLD. Finally, it will address limitations of existing studies along with future directions for microbiome research in NAFLD, including potential microbe-related treatments.

Keywords: Microbiota, fatty liver, fibrosis, obesity, nutrition

Experimental Biology and Medicine 2019; 244: 408-418. DOI: 10.1177/1535370219836739

#### Introduction

The human microbiome represents all microorganisms residing on or within the human body, including bacteria, archaea, fungi, protozoans, and viruses. The human microbiome has the same number of cells and about 100 times more genes than the human body. These genes encode a wide array of pathways that produce bioactive molecules derived from dietary or metabolic precursors. While the gut microbiome has many beneficial functions such as the extraction of energy from otherwise indigestible dietary

fiber, there is increasing evidence connecting the microbiome and its metabolites to the development of certain diseases including obesity, metabolic syndrome, and non-alcoholic fatty liver disease (NAFLD).<sup>5</sup>

The incidence of NAFLD is rapidly growing in conjunction with the epidemic of obesity and metabolic disorders.<sup>6</sup> The risk factors associated with NAFLD include central obesity, insulin resistance, hyperlipidemia, and metabolic syndrome. Epidemiological studies have suggested that NAFLD is more prevalent in men as compared to women and more prevalent in those with Asian or Hispanic

heritage relative to other racial/ethnic groups. <sup>7,8</sup> NAFLD is now the one of the most common causes of chronic liver disease in the Western world and the top two reasons for cirrhosis and liver transplantation. 9,10 NAFLD is a term that encompasses two distinct diseases: nonalcoholic fatty liver (NAFL) and nonalcoholic steatohepatitis (NASH). While patients with NAFL have only bland steatosis on liver biopsy, patients with NASH will also have lobular inflammation and/or hepatocyte ballooning, a sign of hepatocyte damage. Patients with NAFL will often remain stable for many years and will rarely ever progress further. 11,12 Patients with NASH, however, are more likely to progress to fibrosis, cirrhosis, and hepatocellular carcinoma. 11,13 The progression from NAFL to NASH is associated with metabolic syndrome and insulin resistance.<sup>14</sup> Because of the interplay between the microbiome and energy metabolism, there have been many recent studies that have investigated the relationship between the human microbiome and NAFLD development and progression.

This review will explore how diets associated with obesity and NAFLD affect the microbiome and how the microbiome in turn can influence the pathogenesis of these diseases. We will also review potential mechanisms and pathways that link the microbiome to the development and progression of NAFLD. Finally, we will discuss limitations of current research and explore potential future directions including therapeutic applications.

#### Methods

A comprehensive literature review was performed of studies published from 1995 to the present using the following key terms in PubMed: nonalcoholic fatty liver disease, nonalcoholic steatohepatitis, cirrhosis, fibrosis, obesity, metabolic syndrome, diabetes, fat, adipose, bacteria, and microbiome. Particular emphasis was given to articles published within the last five years.

#### Diet and the microbiome

Diet plays a critical role in the development of obesity, metabolic syndrome, and NAFLD. Epidemiological studies have consistently shown associations of diets high in fat and refined sugar with incidence of obesity and NAFLD. In experimental models, such diets have been shown to increase adiposity, hepatic steatosis, and inflammation. Here, we will discuss how the diets most commonly associated with NAFLD and obesity affect the gut microbiome.

#### Western diet

A Western diet is often defined as a diet that is high in sugar, fat, processed meats, and simple grains while being low in fiber.<sup>17</sup> It has been linked to many negative health outcomes including obesity, insulin resistance, metabolic syndrome, and NAFLD.<sup>17,18</sup> There is now growing evidence that the negative effects of a Western diet may be mediated by shifts in the microbiome.<sup>17</sup> Patients on this diet have been reported to have significantly lower microbial diversity and species richness than those on a more

agrarian diet, features that are associated with gut dysbiosis.<sup>19</sup> The Western diet microbiome is often described as having a higher abundance of Firmicutes with a relatively lower abundance of Bacteroidetes.<sup>20</sup> The high Firmicutes to Bacteroides ratio decreases in subjects who lose weight on either a carbohydrate-restricted or fat-restricted diet. 21,22 At a genus level, a Western diet is associated with depletion of Bidobacterium and Lactobacillus and enrichment of Enterobacter.<sup>23</sup> These perturbations of the microbiome may cause metabolic changes in the host by altering short-chain fatty acid (SCFA) production, release of gut hormones such as peptide YY and glucagon-like peptide, and Toll-like receptor (TLR) signaling induced by lipopolysaccharides (LPSs) and other bacterial products.<sup>22</sup> The role of the microbiome in mediating the link between a Western diet and obesity has been examined extensively in several mouse models. Colonization of germ-free mice with the microbiota of obese mice (induced by leptin-deficiency or a Western diet) results in increased body fat accumulation compared to colonization with microbiota from lean controls. 24,25 Similarly, germ-free mice colonized with feces from obese humans had increased adiposity on a high-fat diet compared to germ-free mice colonized with feces from lean humans in weight discordant twin pairs.<sup>26</sup> Mice deficient in TLR 5 and inflammasome components develop susceptibility to Western diet-induced obesity that can be transmitted to other mice by fecal transplantation, demonstrating that genetic factors can modulate the diet-microbiome interaction.<sup>26</sup>

#### High saturated fat

Similar to the Western diet, diets high in saturated fats can also have deleterious effects on health that may be attributable to the microbiome. Epidemiological studies have shown that diets high in saturated and trans-fat are associated with obesity, cardiovascular disease, and NAFLD.<sup>27,28</sup> Mice that are fed a high saturated fat diet develop similar hepatic steatosis and inflammation as seen in patients with NAFLD.<sup>28</sup> However, not all fats have similar consequences. Diets high in polyunsaturated fats, such as those seen in a Mediterranean diet, have been associated with reduced cardiovascular events and a lower prevalence of obesity. 26-29,31 To examine the role of different fats on the microbiome, one study randomized subjects with risk factors for metabolic syndrome to receive either saturated or monosaturated fats. 32 The authors found that a diet high in saturated fats led to an overabundance of Faecalibacterium prausnitzii, which was not seen in patients on the diet high in monounsaturated fats.<sup>32</sup> However, this association with *F. praus*nitzii should not be construed as negative since the introduction of *F. prausnitzii* was protective against hepatic steatosis and adipose tissue inflammation in mice fed a high-fat diet.<sup>33</sup> In one of the largest studies comparing the effects of saturated and polyunsaturated fats on the microbiome, Menni et al. demonstrated in 876 women that higher polyunsaturated fat intake was associated with higher microbial diversity and expansion of members of the Lachnospiraceae family.<sup>34</sup> These findings suggest that the specific composition of fat may be more critical than

the total amount of fat. This idea is supported by a recent study demonstrating that mice fed on a lard fat diet had a greater abundance of Bacteroides and Bilophila, and a lower abundance of Lactobacillus and Akkermansia, than mice fed a diet with an equal amount of fat derived from fish oil.<sup>35</sup> Mice on a lard fat diet also had higher TLR signaling, white adipocyte inflammation, and insulin resistance as compared to mice on a fish oil diet.<sup>35</sup> Fecal transplantation of the lard fat-associated microbiome into germ-free animals induced the donor's metabolic phenotype in recipients, suggesting that these pathways are in part mediated by the microbiome.<sup>35</sup>

#### Role of the microbiome in obesity and insulin resistance

One of the early pivotal studies that linked the microbiome to the development of obesity came from Turnbaugh et al. in 2006.<sup>24</sup> They used 16s rRNA sequencing to demonstrate an increased ratio of Firmicutes to Bacteroidetes in obese humans and experimental mice on a high-fat diet and found that colonization of germ-free mice with this obesity-associated microbial profile could induce an obese phenotype in the recipients. Since then, several other studies have shown that the microbiome can influence weight gain by affecting host gene expression, metabolism, and ingestive behavior. <sup>36–39</sup> Pathways implicated in these metabolic changes included short-SCFA signaling and LPS activation of TLRs, which can induce altered gene expression, hormone secretion, and energy consumption in adipocytes. 40 Additionally, several papers have shown that the efficacy of surgical weight loss interventions may be in part mediated by shifts in the gut microbiome. 41 For example, a study in mice found that gastric bypass led to a persistent increase in Escherichia and Akkermansia, and that microbial transplantation from these mice into nonoperated germfree mice successfully transferred the donor phenotype.<sup>42</sup>

Insulin resistance is another risk factor for NAFLD that is associated with obesity and may also be affected by the microbiome. In support of this concept, two small randomized controlled trials of probiotics for NAFLD—alone or in combination with metformin, an insulin sensitizer-have reported improvement in insulin resistance, hepatic inflammation, and hepatic steatosis. 43,44 This association between the microbiome and insulin resistance led Wu et al. to examine the effects of metformin on the microbiome of diabetic patients.45 They found that metformin greatly alters the microbiome and that some of its metabolic effects on the host could be recapitulated by transferring this altered microbiome into germ-free animals. Wu's and prior studies support the concept that the microbiome influences insulin sensitivity and demonstrates how the microbiome can potentially alter the course of NAFLD through insulinrelated pathways.

### Microbiome associations across the spectrum of NALFD

The growing evidence linking the microbiome to obesity spurred interest in the potential role of the microbiome in other metabolic diseases including NAFLD. Here, we will review evidence from human studies for microbiome associations with NAFL-, NASH-, and NAFLD-related advanced fibrosis.

#### **NAFL**

Studies that have examined the microbiome profile of patients with NAFL as compared to either healthy controls or weight-matched controls have yielded variable results. Pediatric NAFL patients have been reported to have more Prevotella and less Oscillospira than matched controls. 46-48 In studies of adult NAFL patients, Lactobacillus and Escherichia have been enriched while Coprococcus and Prevotella have been depleted (Table 1). These studies utilized 16s rRNA sequencing, which can only provide insight into composition (what bacteria are there) but not function (what products are made by bacteria that may affect disease). Three studies took a multi'omics approach combining microbiome sequencing with metabolomics analysis to evaluate potential microbial metabolic pathways promoting the development of NAFL.<sup>51–53</sup> Raman *et al.* found 18 differentially abundant stool metabolites associated with NAFL in adults, including elevated levels of derivatives of butanoic, propanoic, and acetic acid.<sup>53</sup> Similarly, Da Silva et al. found enrichment of propionate and isobutyric acid in the feces of NAFL patients. 51 These differences were associated with an increase in serum 2-hydroxybutyrate and L-lactic acid. The most convincing data to date that links the microbiome to the development of NAFL comes from Hoyles et al. 52 This study assessed the hepatic transcriptome, gut metagenome, and serum/urine metabolome of a cohort of nondiabetic obese women. NAFL was associated with increased serum levels of several branchedchain and aromatic compounds. Administration of one of these, phenylacetic acid, to mice colonized with human fecal microbiota triggered hepatic steatosis.

#### **NASH**

Studies characterizing the microbiome profile of NASH compared to NAFL or obese controls have found more consistent differences than has been seen for NAFL.54 In children, patients with NASH generally had more Ruminococcus, Dorea, Streptococcus, and Escherichia as compared to their obese counterparts. 46-48,55 In adults, patients with NASH had lower levels of Faecalibacterium, Ruminococcus, and Bidobacterium<sup>51,56</sup> and a higher level of Lactobacillus.<sup>57</sup> Few studies have examined fecal or serum metabolites distinguishing NASH from simple NAFL, most likely due to the fact that a diagnosis of NASH often requires a liver biopsy in order to distinguish it from NAFL. Del Chierico et al. showed higher levels of 4-methyl-2-pentanone and 2-butanone in the serum of children with NASH. 47 Higher levels of 2-butanone were seen in the serum of adults with NAFL,<sup>52</sup> but the functional significance of this metabolite is still unknown. In a cohort of 16 adults with biopsy-proven NASH, patients with NASH had an increased ratio of primary to secondary bile acids, which the author correlated to an increased risk of hepatic injury.<sup>58</sup>

Table 1. Bacteria genera and fecal/serum metabolites associated with different stages of nonalcoholic fatty liver disease in human studies.

NAFLD subtypes	Community composition (genera)	Fecal metabolites	Serum metabolites
NAFL	↑↓ <i>Bifidobacterium</i> <sup>41,47,a</sup>	↑Acetic acid <sup>48</sup>	↑2-butanone <sup>42,a</sup>
	↑ <i>Lactobacillus</i> 41,45,46,48,a	↑Butanoic acid <sup>48</sup>	↑2-hydroxy-butyrate <sup>46</sup>
	↑↓Oscillobacter <sup>42,45,47,48,a</sup>	↑Cholic acid <sup>53</sup>	↑Isoleucine <sup>47</sup>
	↑↓ <i>Prevotella</i> <sup>43,44,a</sup>	↑Ethanol <sup>43,a</sup>	↑Leucine <sup>47</sup>
	↑Roseburia <sup>48</sup>	↑lsobutyric acid <sup>46</sup>	↑L-lactic acid <sup>46</sup>
	↑↓ <i>Ruminococcus</i> <sup>42,44,46,a</sup>	↑Propanoic acid <sup>48</sup>	↑Phenylacetic acid <sup>47</sup>
	†Blautia <sup>42,44</sup>	↑Propionate <sup>46</sup>	↑Valine <sup>47</sup>
	↑Clostridium <sup>45</sup>	2-butanone <sup>48</sup>	
	<i>Dorea</i> <sup>42,48,a</sup>	·	
	↑Escherichia <sup>44,47</sup>		
	↑Streptococcus <sup>45</sup>		
	↓ <i>Alistipes</i> <sup>45</sup>		
	↓Coprococcus <sup>46,47</sup>		
	√Faecalibacterium <sup>46</sup>		
	↓Odoribacter <sup>45</sup>		
	↓Oscillospira <sup>42,a</sup>		
NASH	↑↓Ruminococcus <sup>41,46,52,a</sup>	↑Chenodeoxycholic acid <sup>53</sup>	↑2-butanone <sup>42,a</sup>
	↑Allisonella <sup>51</sup>	↑Cholic acid <sup>53</sup>	↑4-methyl-2-pentanone <sup>42,a</sup>
	↑Blautia <sup>42,44</sup>	↑Lithocholic acid <sup>53</sup>	↑Ethanol <sup>43</sup>
	↑Clostridium <sup>53</sup>	Litilognolio dola	
	↑Dorea <sup>41</sup>		
	↑Escherichia <sup>43</sup>		
	↑Lactobacillus <sup>41,52</sup>		
	↑Parabacteroides <sup>51</sup>		
	↓Bifidobacterium <sup>41,52</sup>		
	↓Coprococcus <sup>46</sup>		
	↓ <i>Faecalibacterium</i> <sup>46,51,52</sup>		
	↓Oscillospira <sup>41</sup>		
NAFLD-related	↑↓ <i>Prevotella</i> <sup>55,56</sup>		↑3-(4-hydroxyphenyl)lactate <sup>58</sup>
advanced fibrosis	↑↓Ruminococcus <sup>55–57</sup>		†3-phenylpropanoate <sup>57</sup>
advanced horosis	↑Bacteroides <sup>55–58</sup>		- prientylproparioate
	↑Blautia <sup>56</sup>		
	↑Enterococcus <sup>56</sup>		
	↑Escherichia <sup>57,58</sup>		
	↑Klebsiella <sup>55</sup>		
	↑Lactobacillus <sup>56</sup>		
	↑ <i>Parabacteroides</i> <sup>56</sup>		
	↑Roseburia <sup>56</sup>		
	†Streptococcus <sup>56</sup>		
	↓Akkermansia <sup>56</sup>		

<sup>a</sup>Denotes an association that has been reported only in pediatric cases of NAFLD.

NAFLD: nonalcoholic fatty liver disease; NAFL: nonalcoholic fatty liver; NASH: nonalcoholic steatohepatitis.

#### **NAFLD-related advanced fibrosis**

In contrast to NAFL and NASH, which have data from both children and adults, NAFLD-related fibrosis has only been studied in adults due to the slow progression of fibrosis. Advanced fibrosis, defined as a fibrosis stage > 2, is associated with a higher incidence of mortality and liver cancer.<sup>59</sup> Microbiome association studies of NAFLDrelated advanced fibrosis have generally reported a decrease in microbial diversity, often due to expansion of Gram-negative bacteria. 60-62 Multiple studies have found an association between advanced fibrosis and an overabundance of Bacteroides and Escherichia, 60-63 while associations with other genera such as Prevotella have been less consistent.61,64 Utilizing metagenomic sequencing, which allows for species level resolution, Loomba et al. showed that Escherichia coli and Bacteroides vulgatus were higher in patients with NAFLD-related advanced fibrosis.<sup>62</sup> They also examined serum metabolites and showed that 3phenylproanoate was the metabolite with the highest fold increase in advanced fibrosis, though it did not reach significance. Recently, Caussy *et al.* found an association between 3–(4-hydroxyphenyl)lactate, a microbial metabolite involved in amino acid metabolism, and NAFLD-related advanced fibrosis. <sup>63</sup> This metabolite was also strongly correlated with several bacterial species that were associated with hepatic fibrosis, including *Escherichia coli*, *Bacteroides caccae*, and *Clostridium sp.* <sup>62</sup>

# Potential mechanisms that link the microbiome to fatty liver disease

While recent human studies have provided meaningful insights into the composition and possible function of the microbiome in each stage during the development and progression of NAFLD, the findings are largely correlative and do not provide conclusive evidence of whether the microbiome is a critical driver of NALFD or simply responds to

the altered diet and host environment associated with NAFLD. Mechanistic investigation supporting a causative role for the microbiome in NAFLD pathogenesis has largely depended upon animal models. The results of studies evaluating microbial composition and metabolites in animal models of NAFLD are summarized in Table 2.24,65-72 Overall, inflammatory pathways such as TLR signaling, choline deficiency, and bile acid metabolism have been linked to NASH, while SCFAs and amino acid metabolism have been linked more to NAFL. Here we will review the potential mechanisms by which the microbiome influences NAFLD development.

#### Epithelial barrier function, TLR signaling, and endotoxemia

Adult patients with NAFLD as well as healthy patients on a Western diet have both been shown to have a "leaky gut" characterized by higher intestinal permeability and altered tight junctions. 73,74 This disruption in the epithelial gut barrier leads to an increased translocation of bacterial products such as LPS into the portal circulation, potentially inducing hepatic inflammation. One of the first studies to causally link the microbiome to NAFLD demonstrated that mice lacking inflammasome components—which are important to intestinal barrier defense-developed dysbiosis and NASH. Transfer of this dysbiosis to wild-type recipients

could induce NASH via an influx of TLR agonist, specifically TLR4 and TLR9, into the portal circulation. <sup>68</sup> Rahman et al. showed that fibrotic steatohepatitis induced by a highfat, high-cholesterol, and high-fructose diet was exacerbated in mice lacking a gene involved in junctional adhesion molecules, an important component of the intestinal barrier. Administration of antibiotics improved liver histology in these knockout mice, suggesting that products of microbial metabolism crossing an impaired intestinal barrier mediated the phenotype. 75,76 There is also a significant role of the host immune system in modulating gut permeability. Beta7 integrin-deficient mice, which are deficient in intestinal immune populations requiring this integrin for chemotaxis, show decreased insulin resistance on a high-fat diet.<sup>77</sup> Treatment of wild-type mice on a high-fat diet with a local gut anti-inflammatory medication, 5-aminosalicyclic acid, reversed diet-induced bowel inflammation and improved metabolic parameters.<sup>77</sup> The downstream effects of LPS translocation are mediated through induction of TLR signaling in the liver. In several studies, LPS has been shown to induce TLR4, leading to increased NF- $\kappa B$ activation and cytokine production important to the progression from NAFL to NASH. 78,79 Unfortunately, a recent phase 2 trial did not show any significant benefit of TLR4 antagonism in NASH patients. Therefore, the clinical relevance of this pathway remains unclear.<sup>80</sup>

Table 2. Bacterial genera and fecal/serum metabolites associated with NAFL and NASH development in animal models.

NAFLD animal models	Community composition (genera)	Fecal metabolites	Serum metabolites
NAFL (high-fat diet or leptin-deficient mice)	↑Bacteroides <sup>65</sup> ↑Barnesiella <sup>67</sup> ↑Bilophila <sup>64,66</sup> ↑Dorea <sup>66</sup> ↑Helicobacter <sup>65</sup> ↑Oscillospira <sup>65</sup> ↑Roseburia <sup>67</sup> ↑Sutterella <sup>66</sup> ↓↑Allobaculum <sup>67</sup> ↓↑Lactobacillus <sup>62,67</sup> ↓Akkermansia <sup>64–66</sup> ↓Bifidobacterium <sup>66</sup> ↓Flavobacterium <sup>65</sup> ↓Marinitoga <sup>65</sup> ↓Parabacteroides <sup>65,66</sup>	↑Butyrate <sup>21</sup> ↓Deoxycholic acid (relative abundance) <sup>66</sup> ↓Hyodeoxycholic acid (relative abundance) <sup>66</sup>	Taurine-conjugated bile acid <sup>66</sup>
NASH (NASH inducing diet, i.e., methionine-choline deficient diet)	↓Ruminococcus <sup>66</sup> ↑(f) Bacteroidaceae <sup>63</sup> ↑(f) Erysipelotrichaceae <sup>63</sup> ↑(f) Porphyromonadaceae <sup>63</sup> ↑(f) Clostridiaceae <sup>63</sup> ↑Alistipes <sup>60</sup> ↑Bacteroide <sup>60,61,63</sup> ↑Bilophila <sup>61</sup> ↑Blautia <sup>61</sup> ↑Parabacteroides <sup>63</sup> ↑Turicibacter <sup>63</sup> ↓Akkermansia <sup>61</sup> ↓Bifidobacteriu <sup>60,61</sup> ↓Desulfovibrio <sup>61</sup> ↓Enterorhabdus <sup>61</sup> ↓Lactobacillus <sup>63</sup>	↑Hexadecane <sup>60</sup> ↑Tetracosane <sup>60</sup> ↓Arachidic acid <sup>60</sup> ↓Cholic acid <sup>60</sup> ↓Stearic acid <sup>60</sup>	

#### Choline deficiency

The relationship between choline deficiency and NAFLD development has been well established.<sup>81</sup> Deficiency in choline leads to abnormal phospholipid synthesis and alterations in very-low-density lipoprotein secretion, eventually leading to hepatic steatohepatitis. 81 Recently, dietary choline bioavailability was shown to be reduced by the gut microbiome through the production of metabolites such as trimethylamine (TMA). 30,82 Several gut microbes are high utilizers of choline and only low abundance of these microbes is required to greatly reduce host choline levels. 83 Mice fed a high-fat diet have been shown to have increased levels of gut microbes that metabolize choline and produce TMA.  $^{84}$  The liver converts gut-derived TMA to trimethylamine-N-oxide (TMAO) via flavin containing monooxygenase 3.85 Elevated levels of TMAO are associated with cardiovascular disease, which potentially links the extrahepatic manifestations of NAFLD to microbialderived metabolites.86 However, the role of circulating TMAO in NAFLD has not been well studied.

#### Short-chain fatty acids

One of the major functions of the human microbiome is the fermentation of indigestible carbohydrates (e.g., fiber) to produce SCFAs. These SCFAs include acetate, propionate, and butyrate, and they act as a major energy source for intestinal epithelial cells. SCFAs also facilitate a wide array of biological activities including hormone production and gene regulation.<sup>87</sup> Obese individuals as well as individuals with NAFL have higher total levels of gut SCFAs as compared to lean controls. 51,53,88 The administration of inulin-type fructan prebiotics was associated with a reduction in SCFAs in obese women along with a reduction of other metabolic markers.<sup>89</sup> Conversely, certain SCFAs may be beneficial against obesity and NAFLD. One mechanism by which SCFAs can affect the host is by binding to highly specific G-protein coupled receptors (GPR), which mediate distinct effects of each SCFA. For example, in a mouse model of diet-induced obesity, a mixture of SCFA predominantly made up of butyrate reduced hepatic expression of GPR41 and GPR43, two receptors that have been shown to promote hepatic lipid accumulation. 90,91 The positive effect of butyrate was further highlighted by Mattace Raso et al. when they demonstrated that butyrate supplementation was able to improve hepatic steatosis induced in mice by a high-fat diet. <sup>92</sup> Furthermore, fecal microbial transplantation from lean human donors to obese patients resulted in improved insulin sensitivity, which was associated with increased abundance of butyrate-producing bacteria.93 The inconsistent findings on SCFAs are most likely due to the distinct biological effects of individual SCFAs on host metabolism.

#### Bile acid metabolism

The recent development and marketing of obeticholic acid, a farnesoid X receptor (FXR) agonist, underscores the importance of bile acids for host metabolism and health. Gut microbes play a critical role in the regulation of the bile acid pool through conversion of primary bile acids to secondary bile acids, which have distinct functional properties mediated by differential binding to bile acid receptors including FXR and G-protein coupled bile acid receptor 1 (GPBAR1).<sup>94</sup> In a murine model of NAFLD, animals with intestine-specific FXR disruption developed changes in their gut microbiome that were associated with reduced triglyceride accumulation in response to a high-fat diet as compared to controls.<sup>95</sup> In mice treated with antibiotics, there was an increase in conjugated bile acid metabolites that inhibited intestinal FXR signaling. 95 GPBAR1 signaling was also found to be necessary for sustained weight loss and improved fatty liver in mice undergoing sleeve gastrectomy. 6 In humans, a phase 2 clinical trial with obeticholic acid in patients with NASH showed improvement by histology after 72 weeks of treatment. 97 The administration of obeticholic acid also led to a reversible induction of Grampositive bacteria in the human small intestine and increased proportion of Firmicutes in mice. 98 While initial results are promising, ongoing studies and phase 3 trials are underway in order to better understand the complex relationship between the gut microbiome, bile acid synthesis, and FXR signaling.

#### Amino acid metabolism

The gut microbiome can also affect the synthesis and metabolism of aromatic and branched-chain amino acids (BCAAs). In patients with insulin resistance, Prevotella copri and Bacteroides vulgatus were identified as the main species associated with increased BCAAs and insulin resistance. 99 The authors also showed that mice gavaged with P. copri developed increased insulin resistance when fed a high-fat diet as compared to controls. 99 In a recent study, Hoyles et al. demonstrated that phenylacetic acid, an aromatic amino acid derived from microbial metabolism, was strongly associated with hepatic steatosis in humans.<sup>52</sup> They also showed that the addition of phenylacetic acid in both primary human hepatocyte cultures and in mice models could trigger hepatic steatosis, implying a causal effect in NAFL.52

#### Therapeutic implications, limitations, and future directions

The growing evidence that links the human microbiome to NAFLD progression has motivated interest in the development of novel microbiome-related therapies for NAFLD. Microbiome-related interventions include gut-specific antibiotics, probiotics, prebiotics, and fecal microbial transplant (FMT).<sup>54</sup> However, large well-designed clinical studies examining microbiome-related interventions in NAFLD are lacking. Several randomized controlled trials involving probiotics in NAFLD have yielded conflicting results due to the lack of standardization across studies. 100 As of yet, no randomized controlled trial involving probiotics has shown any significant changes in body mass index (BMI). 100 Several small trials have shown a potential benefit of probiotics on important markers including insulin resistance, alanine aminotransferase (ALT), aspartate aminotransferase (AST), and

Table 3. Summary of randomized control trials involving NAFLD and probiotics.

Study	Number of patients	Intervention	Major findings
Aller et al. 101	30	Lactobacillus delbrueckii subsp. bulgaricus + Streptococcus thermophilus vs. placebo for 3 months	Decrease in ALT, AST, GGT
Alisi et al. 102	44 <sup>a</sup>	VSL#3 (Bifidobacterium longum, Bifidobacterium infantis, Bifidobacterium breve, Lactobacillus acidophilus, Lactobacillus casei, Lactobacillus delbrueckii subsp. bulgaricus, Lactobacillus plantarum, Streptococcus thermophilus) vs. placebo for 4 months	Ultrasound improvement in fatty liver
Eslamparast et al. <sup>103</sup>	19	Protexin (Bifidobacterium breve, Bifidobacterium longum, Lactobacillus casei, Lactobacillus rhamnosus, Lactobacillus acidophilus, Lactobacillus delbrueckii subsp. bulgaricus, Streptococcus thermophilus) vs. placebo for 28 weeks	Decrease in ALT, AST, GGT, CRP, TNF $\alpha$ , fibrosis score by transient elastography
Malaguarnera et al. <sup>44</sup>	66	Bifidobacterium longum + fructo-oligosaccharides + life-style modification vs. life-style modification alone	Decrease in AST, LDL, CRP, TNFα, HOMA-IR, steatosis, and NASH activity index
Shavakhi et al. <sup>43</sup>	64	Protexin + Metformin vs. Metformin alone	Decrease in ALT, AST, ultrasound grading of steatosis
Wong et al. 109	20	Lepicol (Bifidobacterium bifidum, Lactobacillus plantarum, Lactobacillus delbrueckii subsp. bulgaricus, Lactobacillus acidophilus, Lactobacillus rhamnosus) vs. nothing	Decrease in intrahepatic triglyceride content as measured by proton-magnetic resonance spectroscopy

<sup>&</sup>lt;sup>a</sup>Denotes a pediatric trial.

NASH: nonalcoholic steatohepatitis; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; GGT: Gamma-glutamyl transferase; CRP: C-reactive protein; TNF: Tumor necrosis factor; LDL: low-density lipoprotein; HOMA: Homeostatic model assessment.

histology grade (Table 3).43,44,101-103 For example, a small randomized trial with 66 patients showed that supplementation with Bidobacterium longum and fructo-oligosaccharides improved insulin resistance, hepatic steatosis, and NASH activity index after 24 weeks of treatment. 44 These findings have not yet been reproduced in larger published studies. There is also no data available about the role of FMT in NAFLD, though there are now two actively recruiting clinical trials designed to address this question. 104,105 However, until there is a better understanding of the key mechanistic pathways by which the microbiome promotes NAFLD, the development of microbiome-related therapies will be limited. Nonetheless, a recent multi'omics study has provided initial support for the potential application of microbes and their metabolites as noninvasive biomarkers for diagnosis and prognostication of NAFLD.62

Despite major recent advances in microbiome research, the field is still in its infancy with many areas that can be improved upon. One of the main challenges to interpreting the existing literature on the microbiome and NAFLD is heterogeneity in study design. In particular, there has been wide variation in selection of healthy controls (including inconsistent BMI-matching), age range of study populations, incorporation of diet into the analyses, and sample collection/processing. In addition, early studies examining the gut microbiome and NAFLD have been predominantly association studies. These studies are unable to differentiate whether the microbial profile described was a potential cause of NAFLD or rather a byproduct of the environment. Moreover, relevant microbial metabolites that reach the liver may be produced primarily in the small intestine and/or proximal colon, which may not be well represented by the microbiome and metabolome of feces. At this time, studies are shifting away from these types of analysis and

are moving towards studies focusing on mechanistic pathways. 106 This can be achieved in human studies by using a multi'omics approach combining microbiome analysis with other modalities including metabolomics, proteomics, and host transcriptomics to develop a systems level understanding of NAFLD development and progression. Such studies are complemented by experiments involving transplantation of human microbiota into germ-free or antibiotic-treated animals to establish causal relationships between dysbiosis observed in human cohorts and metabolic outcomes. 41,52

Currently, 16s rRNA sequencing is the most common method for microbiome analysis. 106 It is effective for defining microbial composition and taxonomy to the genus and to some extent species level but does not provide functional data (i.e., presence of bacterial genes and their expression level). In order to achieve this level of specificity, shotgun metagenomic sequencing and/or metatranscriptomics are required. 107 Unfortunately, due to high cost, the sequencing of bacterial metagenomes and metatranscriptomes is still out of reach for many investigators. With ongoing advances in sequencing technology, it is likely that the price of these services will decrease sufficiently to allow for more widespread use in the future, similar to the widespread adoption of 16s rRNA sequencing after the dramatic decrease in sequencing costs early this decade. 108

#### **Conclusions**

In summary, both animal models and human studies have supported the relationship between the gut microbiome and development and progression of NAFLD. By affecting gut barrier function, TLR signaling, choline metabolism, bile acid synthesis, SCFA, and amino acid production, the

gut microbiome appears to play a critical and multifactorial role in NAFLD development. But despite advances in technology and bioinformatics analysis, specific mechanistic pathways are not yet clearly defined. Future large, longitudinal, prospective studies incorporating multi-omics analysis and humanized animal models are needed to better define the multifactorial host-microbiome relationship involved in fatty liver pathogenesis.

**Author Contributions:** TD and JJ contributed equally to the design and writing of this manuscript.

#### **DECLARATION OF CONFLICTING INTERESTS**

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

#### **FUNDING**

This work is funded by National Institues of Health grant T32 DK 07180 (TD) and Veterans Affairs grant CDAII IK2CX001717 (JJ).

#### **ORCID iD**

Tien S Dong http://orcid.org/0000-0003-0105-8063 Jonathan P Jacobs http://orcid.org/0000-0003-4698-0254

#### **REFERENCES**

- Ursell LK, Metcalf JL, Parfrey LW, Knight R. Defining the human microbiome. Nutr Rev 2012;70:S38-44
- Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. PLoS Biol 2016;14:e1002533
- 3. Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li J, Xu J, Li S, Li D, Cao J, Wang B, Liang H, Zheng H, Xie Y, Tap J, Lepage P, Bertalan M, Batto J-M, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu H, Yu C, Li S, Jian M, Zhou Y, Li Y, Zhang X, Li S, Qin N, Yang H, Wang J, Brunak S, Doré J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J, MetaHIT Consortium Bork P, Ehrlich SD, Wang J. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 2010;464:59-65
- Lynch SV, Pedersen O. The human intestinal microbiome in health and disease. N Engl J Med 2016;375:2369–79
- Shreiner AB, Kao JY, Young VB. The gut microbiome in health and in disease. Curr Opin Gastroenterol 2015;31:69-75
- Fabbrini E, Sullivan S, Klein S. Obesity and nonalcoholic fatty liver disease: biochemical, metabolic, and clinical implications. *Hepatology* 2010;51:679–89
- Lim H-W, Bernstein DE. Risk factors for the development of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis, including genetics. Clin Liver Dis 2018;22:39–57
- Ballestri S, Nascimbeni F, Baldelli E, Marrazzo A, Romagnoli D, Lonardo A. NAFLD as a sexual dimorphic disease: role of gender and reproductive status in the development and progression of nonalcoholic fatty liver disease and inherent cardiovascular risk. *Adv Ther* 2017;34:1291–326
- 9. Lazo M, Clark JM. The epidemiology of nonalcoholic fatty liver disease: a global perspective. *Semin Liver Dis* 2008;**28**:339–50
- Kemmer N, Neff GW, Franco E, Osman-Mohammed H, Leone J, Parkinson E, Cece E, Alsina A. Nonalcoholic fatty liver disease epidemic and its implications for liver transplantation. *Transplantation* 2013;96:860-2

- 11. Vernon G, Baranova A, Younossi ZM. Systematic review: the epidemiology and natural history of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in adults. *Aliment Pharmacol Ther* 2011;34:274–85
- 12. Teli MR, James OF, Burt AD, Bennett MK, Day CP. The natural history of nonalcoholic fatty liver: a follow-up study. *Hepatology* 1995:**22**:1714–9
- Ekstedt M, Hagström H, Nasr P, Fredrikson M, Stål P, Kechagias S, Hultcrantz R. Fibrosis stage is the strongest predictor for diseasespecific mortality in NAFLD after up to 33 years of follow-up. Hepatology 2015;61:1547–54
- Losekann A, Weston AC, de Mattos AA, Tovo CV, de Carli LA, Espindola MB, Pioner SR, Coral GP. Non-alcoholic steatohepatitis (NASH): risk factors in morbidly obese patients. *IJMS*. 2015;16:25552-9
- Rodríguez-Monforte M, Sánchez E, Barrio F, Costa B, Flores-Mateo G. Metabolic syndrome and dietary patterns: a systematic review and meta-analysis of observational studies. Eur J Nutr 2017;56:925–47
- 16. Softic S, Cohen DE, Kahn CR. Role of dietary fructose and hepatic de novo lipogenesis in fatty liver disease. *Dig Dis Sci* 2016;61:1282–93
- 17. Turnbaugh PJ, Ridaura VK, Faith JJ, Rey FE, Knight R, Gordon JI. The effect of diet on the human gut microbiome: a metagenomic analysis in humanized gnotobiotic mice. *Sci Transl Med* 2009;1:6ra14
- 18. Zinöcker MK, Lindseth IA. The western diet-microbiome-host interaction and its role in metabolic disease. *Nutrients* 2018;10:piiE365
- Gupta VK, Paul S, Dutta C. Geography, ethnicity or subsistencespecific variations in human microbiome composition and diversity. Front Microbiol 2017;8:1162
- Ley RE, Bäckhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI. Obesity alters gut microbial ecology. Proc Natl Acad Sci U S A 2005;102:11070-5
- Clarke SF, Murphy EF, Nilaweera K, Ross PR, Shanahan F, O'Toole PW, Cotter PD. The gut microbiota and its relationship to diet and obesity: new insights. *Gut Microbes* 2012;3:186–202
- 22. Parekh PJ, Balart LA, Johnson DA. The influence of the gut microbiome on obesity, metabolic syndrome and gastrointestinal disease. *Clin Transl Gastroenterol* 2015;6:e91
- 23. Singh RK, Chang H-W, Yan D, Lee KM, Ucmak D, Wong K, Abrouk M, Farahnik B, Nakamura M, Zhu TH, Bhutani T, Liao W. Influence of diet on the gut microbiome and implications for human health. *J Transl Med* 2017;**15**:73
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006;444:1027–31
- Turnbaugh PJ, Bäckhed F, Fulton L, Gordon JI. Diet-induced obesity is linked to marked but reversible alterations in the mouse distal gut microbiome. Cell Host Microbe 2008;3:213–23
- 26. Ridaura VK, Faith JJ, Rey FE, Cheng J, Duncan AE, Kau AL, Griffin NW, Lombard V, Henrissat B, Bain JR, Muehlbauer MJ, Ilkayeva O, Semenkovich CF, Funai K, Hayashi DK, Lyle BJ, Martini MC, Ursell LK, Clemente JC, Van Treuren W, Walters WA, Knight R, Newgard CB, Heath AC, Gordon JI. Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science* 2013;341:1241214
- Kris-Etherton PM, Harris WS, Appel LJ; American Heart Association. Nutrition Committee. Fish consumption, fish oil, omega-3 fatty acids, and cardiovascular disease. Circulation 2002;106:2747–57
- 28. Nakamura A, Terauchi Y. Lessons from mouse models of high-fat dietinduced NAFLD. *Int J Mol Sci* 2013;**14**:21240–57
- De Filippis F, Pellegrini N, Vannini L, Jeffery IB, La Storia A, Laghi L, Serrazanetti DI, Di Cagno R, Ferrocino I, Lazzi C, Turroni S, Cocolin L, Brigidi P, Neviani E, Gobbetti M, O'Toole PW, Ercolini D. High-level adherence to a Mediterranean diet beneficially impacts the gut microbiota and associated metabolome. *Gut* 2016;65:1812–21
- 30. Velasquez MT, Ramezani A, Manal A, Raj DS. Trimethylamine Noxide: the good, the bad and the unknown. *Toxins (Basel)* 2016;8:piiE326.
- Koloverou E, Panagiotakos DB, Pitsavos C, Chrysohoou C, Georgousopoulou EN, Grekas A, Christou A, Chatzigeorgiou M, Skoumas I, Tousoulis D, Stefanadis C. Attica Study Group

- Adherence to Mediterranean diet and 10-year incidence (2002-2012) of diabetes: correlations with inflammatory and oxidative stress biomarkers in the ATTICA cohort study. Diabetes Metab Res Rev 2016;32:73-81
- 32. Fava F, Gitau R, Griffin BA, Gibson GR, Tuohy KM, Lovegrove JA. The type and quantity of dietary fat and carbohydrate alter faecal microbiome and short-chain fatty acid excretion in a metabolic syndrome 'at-risk' population. Int J Obes (Obes) 2013;37:216-23
- 33. Munukka E, Rintala A, Toivonen R, Nylund M, Yang B, Takanen A, Hänninen A, Vuopio J, Huovinen P, Jalkanen S, Pekkala S. Faecalibacterium prausnitzii treatment improves hepatic health and reduces adipose tissue inflammation in high-fat fed mice. ISME J 2017:11:1667-79
- 34. Menni C, Zierer J, Pallister T, Jackson MA, Long T, Mohney RP, Steves CJ, Spector TD, Valdes AM. Omega-3 fatty acids correlate with gut microbiome diversity and production of N-carbamylglutamate in middle aged and elderly women. Sci Rep 2017;7:11079
- 35. Caesar R, Tremaroli V, Kovatcheva-Datchary P, Cani PD, Bäckhed F. Crosstalk between gut microbiota and dietary lipids aggravates WAT inflammation through TLR signaling. Cell Metab 2015;22:658-68
- Ussar S, Griffin NW, Bezy O, Fujisaka S, Vienberg S, Softic S, Deng L, Bry L, Gordon JI, Kahn CR. Interactions between gut microbiota, host genetics and diet modulate the predisposition to obesity and metabolic syndrome. Cell Metab 2015;22:516-30
- 37. Clapper JR, Hendricks MD, Gu G, Wittmer C, Dolman CS, Herich J, Athanacio J, Villescaz C, Ghosh SS, Heilig JS, Lowe C, Roth JD. Dietinduced mouse model of fatty liver disease and nonalcoholic steatohepatitis reflecting clinical disease progression and methods of assessment. Am J Physiol Gastrointest Liver Physiol 2013;305:G483-95
- 38. Tillisch K, Labus J, Kilpatrick L, Jiang Z, Stains J, Ebrat B, Guyonnet D, Legrain-Raspaud S, Trotin B, Naliboff B, Mayer EA. Consumption of fermented milk product with probiotic modulates brain activity. Gastroenterology 2013;144:1394-401, 1401.e1-4
- 39. Maruvada P, Leone V, Kaplan LM, Chang EB. The human microbiome and obesity: moving beyond associations. Cell Host Microbe
- 40. Bohan R, Tianyu X, Tiantian Z, Ruonan F, Hongtao H, Qiong W, Chao S. Gut microbiota: a potential manipulator for host adipose tissue and energy metabolism. J Nutr Biochem 2018;64:206-17
- 41. Tremaroli V, Karlsson F, Werling M, Ståhlman M, Kovatcheva-Datchary P, Olbers T, Fändriks L, Le Roux CW, Nielsen J, Bäckhed F. Roux-en-Y gastric bypass and vertical banded gastroplasty induce long-term changes on the human gut microbiome contributing to fat mass regulation. Cell Metab 2015:22:228-38
- 42. Liou AP, Paziuk M, Luevano J-M, Machineni S, Turnbaugh PJ, Kaplan LM. Conserved shifts in the gut microbiota due to gastric bypass reduce host weight and adiposity. Sci Transl Med 2013;5:178ra41
- 43. Shavakhi A, Minakari M, Firouzian H, Assali R, Hekmatdoost A, Ferns G. Effect of a probiotic and metformin on liver aminotransferases in non-alcoholic steatohepatitis: a double blind randomized clinical trial. Int J Prev Med 2013;4:531-7
- 44. Malaguarnera M, Vacante M, Antic T, Giordano M, Chisari G, Acquaviva R, Mastrojeni S, Malaguarnera G, Mistretta A, Li Volti G, Galvano F. Bifidobacterium longum with fructo-oligosaccharides in patients with non alcoholic steatohepatitis. Dig Dis Sci 2012;57:545-53
- 45. Wu H, Esteve E, Tremaroli V, Khan MT, Caesar R, Mannerås-Holm L, Ståhlman M, Olsson LM, Serino M, Planas-Fèlix M, Xifra G, Mercader JM, Torrents D, Burcelin R, Ricart W, Perkins R, Fernàndez-Real JM, Bäckhed F. Metformin alters the gut microbiome of individuals with treatment-naive type 2 diabetes, contributing to the therapeutic effects of the drug. Nat Med 2017;23:850-58
- 46. Nobili V, Putignani L, Mosca A, Chierico F, Del Vernocchi P, Alisi A, Stronati L, Cucchiara S, Toscano M, Drago L. Bifidobacteria and lactobacilli in the gut microbiome of children with non-alcoholic fatty liver disease: which strains act as health players? AOMS 2018;14:81-7
- 47. Del Chierico F, Nobili V, Vernocchi P, Russo A, Stefanis C, De Gnani D, Furlanello C, Zandonà A, Paci P, Capuani G, Dallapiccola B, Miccheli A, Alisi A, Putignani L. Gut microbiota profiling of pediatric

nonalcoholic fatty liver disease and obese patients unveiled by an integrated meta-omics-based approach. Hepatology 2017;65:451-64

.....

- 48. Michail S, Lin M, Frey MR, Fanter R, Paliy O, Hilbush B, Reo NV. Altered gut microbial energy and metabolism in children with nonalcoholic fatty liver disease. FEMS Microbiol Ecol 2015;91:1-9
- 49. Shen F, Zheng R-D, Sun X-Q, Ding W-J, Wang X-Y, Fan J-G. Gut microbiota dysbiosis in patients with non-alcoholic fatty liver disease. Hepatobiliary Pancreat Dis Int 2017;16:375-81
- 50. Jiang W, Wu N, Wang X, Chi Y, Zhang Y, Qiu X, Hu Y, Li J, Liu Y. Dysbiosis gut microbiota associated with inflammation and impaired mucosal immune function in intestine of humans with non-alcoholic fatty liver disease. Sci Rep 2015;5:8096
- 51. Da Silva HE, Teterina A, Comelli EM, Taibi A, Arendt BM, Fischer SE, Lou W, Allard JP. Nonalcoholic fatty liver disease is associated with dysbiosis independent of body mass index and insulin resistance. Sci
- 52. Hoyles L, Fernández-Real J-M, Federici M, Serino M, Abbott J, Charpentier J, Heymes C, Luque JL, Anthony E, Barton RH, Chilloux J, Myridakis A, Martinez-Gili L, Moreno-Navarrete JM, Benhamed F, Azalbert V, Blasco-Baque V, Puig J, Xifra G, Ricart W, Tomlinson C, Woodbridge M, Cardellini M, Davato F, Cardolini I, Porzio O, Gentileschi P, Lopez F, Foufelle F, Butcher SA, Holmes E, Nicholson JK, Postic C, Burcelin R, Dumas M-E. Molecular phenomics and metagenomics of hepatic steatosis in non-diabetic obese women. Nat Med 2018;24:1070-80
- 53. Raman M, Ahmed I, Gillevet PM, Probert CS, Ratcliffe NM, Smith S, Greenwood R, Sikaroodi M, Lam V, Crotty P, Bailey J, Myers RP, Rioux KP. Fecal microbiome and volatile organic compound metabolome in obese humans with nonalcoholic fatty liver disease. Clin Gastroenterol Hepatol 2013;11:868-875.e3
- 54. Sharpton SR, Ajmera V, Loomba R. Emerging role of the gut microbiome in nonalcoholic fatty liver disease: from composition to function. Clin Gastroenterol Hepatol 2019;17:296-306
- 55. Zhu L, Baker SS, Gill C, Liu W, Alkhouri R, Baker RD, Gill SR. Characterization of gut microbiomes in nonalcoholic steatohepatitis (NASH) patients: a connection between endogenous alcohol and NASH. Hepatology 2013;57:601-9
- 56. Wong VW-S, Tse C-H, Lam TT-Y, Wong GL-H, Chim AM-L, Chu WC-W, Yeung DK-W, Law PT-W, Kwan H-S, Yu J, Sung JJ-Y, Chan HL-Y. Molecular characterization of the fecal microbiota in patients with nonalcoholic steatohepatitis - a longitudinal study. PLoS One 2013;8:e62885
- 57. Duarte SMB, Stefano JT, Miele L, Ponziani FR, Souza-Basqueira M, Okada LSRR, de Barros Costa FG, Toda K, Mazo DFC, Sabino EC, Carrilho FJ, Gasbarrini A, Oliveira CP. Gut microbiome composition in lean patients with NASH is associated with liver damage independent of caloric intake: a prospective pilot study. Nutr Metab Cardiovasc Dis 2018;28:369-84
- 58. Mouzaki M, Wang AY, Bandsma R, Comelli EM, Arendt BM, Zhang L, Fung S, Fischer SE, McGilvray IG, Allard JP. Bile acids and dysbiosis in non-alcoholic fatty liver disease. PLoS One 2016;11:e0151829
- 59. Arulanandan A, Loomba R. Noninvasive testing for NASH and NASH with advanced fibrosis: are we there yet? Curr Hepatol Rep 2015;14:109-18
- 60. Million M, Maraninchi M, Henry M, Armougom F, Richet H, Carrieri P, Valero R, Raccah D, Vialettes B, Raoult D. Obesity-associated gut microbiota is enriched in Lactobacillus reuteri and depleted in Bidobacterium animalis and Methanobrevibacter smithii. Int J Obes Relat Metab Disord 2012:36:817-25
- 61. Ponziani FR, Bhoori S, Castelli C, Putignani L, Rivoltini L, Del Chierico F, Sanguinetti M, Morelli D, Paroni Sterbini F, Petito V, Reddel S, Calvani R, Camisaschi C, Picca A, Tuccitto A, Gasbarrini A, Pompili M, Mazzaferro V. Hepatocellular carcinoma is associated with gut microbiota profile and inflammation in nonalcoholic fatty liver disease. Hepatology 2019;69:107-20
- 62. Loomba R, Seguritan V, Li W, Long T, Klitgord N, Bhatt A, Dulai PS, Caussy C, Bettencourt R, Highlander SK, Jones MB, Sirlin CB, Schnabl B, Brinkac L, Schork N, Chen C-H, Brenner DA, Biggs W, Yooseph S, Venter JC, Nelson KE. Gut microbiome-based metagenomic signature

for non-invasive detection of advanced fibrosis in human nonalcoholic fatty liver disease. Cell Metab 2017;25:1054-62.e5

.....

- 63. Caussy C, Hsu C, Lo M-T, Liu A, Bettencourt R, Ajmera VH, Bassirian S, Hooker J, Sy E, Richards L, Schork N, Schnabl B, Brenner DA, Sirlin CB, Chen C-H, Loomba R. Genetics of NAFLD in Twins Consortium. Link between gut-microbiome derived metabolite and shared geneeffects with hepatic steatosis and fibrosis in NAFLD. *Hepatology*. Epub ahead of print 23 March 2018. DOI: 10.1002/hep.29892
- 64. Boursier J, Mueller O, Barret M, Machado M, Fizanne L, Araujo-Perez F, Guy CD, Seed PC, Rawls JF, David LA, Hunault G, Oberti F, Calès P, Diehl AM. The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. *Hepatology* 2016;63:764–75
- 65. Ye J-Z, Li Y-T, Wu W-R, Shi D, Fang D-Q, Yang L-Y, Bian X-Y, Wu J-J, Wang Q, Jiang X-W, Peng C-G, Ye W-C, Xia P-C, Li L-J. Dynamic alterations in the gut microbiota and metabolome during the development of methionine-choline-deficient diet-induced nonalcoholic steatohepatitis. WJG 2018;24:2468–81
- 66. Ye J, Lv L, Wu W, Li Y, Shi D, Fang D, Guo F, Jiang H, Yan R, Ye W, Li L. Butyrate protects mice against methionine-choline-deficient dietinduced non-alcoholic steatohepatitis by improving gut barrier function, attenuating inflammation and reducing endotoxin levels. Front Microbiol 2018;9:1967
- Zeng H, Liu J, Jackson MI, Zhao F-Q, Yan L, Combs GF. Fatty liver accompanies an increase in lactobacillus species in the hind gut of C57BL/6 mice fed a high-fat diet. J Nutr 2013;143:627–31
- Henao-Mejia J, Elinav E, Jin C, Hao L, Mehal WZ, Strowig T, Thaiss CA, Kau AL, Eisenbarth SC, Jurczak MJ, Camporez J-P, Shulman GI, Gordon JI, Hoffman HM, Flavell RA. Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. *Nature* 2012;482:179–85
- 69. Schneeberger M, Everard A, Gómez-Valadés AG, Matamoros S, Ramírez S, Delzenne NM, Gomis R, Claret M, Cani PD. Akkermansia muciniphila inversely correlates with the onset of inflammation, altered adipose tissue metabolism and metabolic disorders during obesity in mice. Sci Rep 2015;5:16643
- Moreira GV, Azevedo FF, Ribeiro LM, Santos A, Guadagnini D, Gama P, Liberti EA, Saad M, Carvalho C. Liraglutide modulates gut microbiota and reduces NAFLD in obese mice. J Nutr Biochem 2018;62:143–54
- Natividad JM, Lamas B, Pham HP, Michel M-L, Rainteau D, Bridonneau C, da Costa G, van Hylckama Vlieg J, Sovran B, Chamignon C, Planchais J, Richard ML, Langella P, Veiga P, Sokol H. Bilophila wadsworthia aggravates high fat diet induced metabolic dysfunctions in mice. Nat Commun 2018;9:2802
- 72. Le Roy T, Llopis M, Lepage P, Bruneau A, Rabot S, Bevilacqua C, Martin P, Philippe C, Walker F, Bado A, Perlemuter G, Cassard-Doulcier A-M, Gérard P. Intestinal microbiota determines development of non-alcoholic fatty liver disease in mice. *Gut* 2013;62:1787–94
- Pendyala S, Walker JM, Holt PR. A high-fat diet is associated with endotoxemia that originates from the gut. Gastroenterology 2012;142:1100-01.e2
- Miele L, Valenza V, La Torre G, Montalto M, Cammarota G, Ricci R, Mascianà R, Forgione A, Gabrieli ML, Perotti G, Vecchio FM, Rapaccini G, Gasbarrini G, Day CP, Grieco A. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. Hepatology 2009;49:1877–87
- 75. Rahman K, Desai C, Iyer SS, Thorn NE, Kumar P, Liu Y, Smith T, Neish AS, Li H, Tan S, Wu P, Liu X, Yu Y, Farris AB, Nusrat A, Parkos CA, Anania FA. Loss of junctional adhesion molecule a promotes severe steatohepatitis in mice on a diet high in saturated fat, fructose, and cholesterol. *Gastroenterology* 2016;151:733–46.e12
- Bergheim I, Weber S, Vos M, Krämer S, Volynets V, Kaserouni S, McClain CJ, Bischoff SC. Antibiotics protect against fructoseinduced hepatic lipid accumulation in mice: role of endotoxin. *I Hepatol* 2008;48:983–92
- Luck H, Tsai S, Chung J, Clemente-Casares X, Ghazarian M, Revelo XS, Lei H, Luk CT, Shi SY, Surendra A, Copeland JK, Ahn J, Prescott D, Rasmussen BA, Chng MHY, Engleman EG, Girardin SE, Lam TKT,

- Croitoru K, Dunn S, Philpott DJ, Guttman DS, Woo M, Winer S, Winer DA. Regulation of obesity-related insulin resistance with gut anti-inflammatory agents. *Cell Metab* 2015;21:527–42
- 78. Ye D, Li FYL, Lam KSL, Li H, Jia W, Wang Y, Man K, Lo CM, Li X, Xu A. Toll-like receptor-4 mediates obesity-induced non-alcoholic steato-hepatitis through activation of X-box binding protein-1 in mice. *Gut* 2012;61:1058–67
- Sharifnia T, Antoun J, Verriere TGC, Suarez G, Wattacheril J, Wilson KT, Peek RM, Abumrad NN, Flynn CR. Hepatic TLR4 signaling in obese NAFLD. Am J Physiol Gastrointest Liver Physiol 2015;309:G270–8
- Diehl A, Harrison S, Caldwell S, Rinella M, Paredes A, Moylan C, Guy C, Bashir M, Wang Y, Miller L, Chang A, Wu E, Abdelmalek M. JKB-121 in patients with nonalcoholic steatohepatitis: a phase 2 double blind randomized placebo control study. J Hepatol 2018;68:S103
- 81. Sherriff JL, O'Sullivan TA, Properzi C, Oddo J-L, Adams LA. Choline, its potential role in nonalcoholic fatty liver disease, and the case for human and bacterial genes. *Adv Nutr* 2016;7:5–13
- 82. Stremmel W, Schmidt KV, Schuhmann V, Kratzer F, Garbade SF, Langhans C-D, Fricker G, Okun JG. Blood trimethylamine-N-oxide originates from microbiota mediated breakdown of phosphatidylcholine and absorption from small intestine. *PLoS One* 2017;12:e0170742
- 83. Rath S, Heidrich B, Pieper DH, Vital M. Uncovering the trimethylamine-producing bacteria of the human gut microbiota. *Microbiome* 2017;5:54
- 84. Dumas M-E, Barton RH, Toye A, Cloarec O, Blancher C, Rothwell A, Fearnside J, Tatoud R, Blanc V, Lindon JC, Mitchell SC, Holmes E, McCarthy MI, Scott J, Gauguier D, Nicholson JK. Metabolic profiling reveals a contribution of gut microbiota to fatty liver phenotype in insulin-resistant mice. *Proc Natl Acad Sci U S A* 2006;**103**:12511–6
- 85. Treacy EP, Akerman BR, Chow LM, Youil R, Bibeau C, Lin J, Bruce AG, Knight M, Danks DM, Cashman JR, Forrest SM. Mutations of the flavin-containing monooxygenase gene (FMO3) cause trimethylaminuria, a defect in detoxication. *Hum Mol Genet* 1998;7:839–45
- Tang WHW, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, Wu Y, Hazen SL. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. N Engl J Med 2013;368:1575–84
- 87. Ríos-Covián D, Ruas-Madiedo P, Margolles A, Gueimonde M, de Los Reyes-Gavilán CG, Salazar N. Intestinal short chain fatty acids and their link with diet and human health. *Front Microbiol* 2016;7:185
- 88. Schwiertz A, Taras D, Schäfer K, Beijer S, Bos NA, Donus C, Hardt PD. Microbiota and SCFA in lean and overweight healthy subjects. *Obesity (Silver Spring)* 2010;18:190–5
- 89. Salazar N, Dewulf EM, Neyrinck AM, Bindels LB, Cani PD, Mahillon J, de Vos WM, Thissen J-P, Gueimonde M, de Los Reyes-Gavilán CG, Delzenne NM. Inulin-type fructans modulate intestinal Bifidobacterium species populations and decrease fecal short-chain fatty acids in obese women. Clin Nutr 2015;34:501-7
- Lu Y, Fan C, Li P, Lu Y, Chang X, Qi K. Short chain fatty acids prevent high-fat-diet-induced obesity in mice by regulating G protein-coupled receptors and gut microbiota. Sci Rep 2016;6:37589
- 91. Lu J, Fang B, Zheng Y, Yu X, Huang G, Wang Z, Deng X, Guan S. 1,3-Dichloro-2-propanol induced lipid accumulation in HepG2 cells through cAMP/protein kinase A and AMP-activated protein kinase pathways via Gi/o-coupled receptors. *Environ Toxicol Pharmacol* 2017;55:118–26
- 92. Mattace Raso G, Simeoli R, Russo R, Iacono A, Santoro A, Paciello O, Ferrante MC, Canani RB, Calignano A, Meli R. Effects of sodium butyrate and its synthetic amide derivative on liver inflammation and glucose tolerance in an animal model of steatosis induced by high fat diet. *PLoS One.* 2013;8:e68626
- 93. Vrieze A, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman Jfwm Dallinga-Thie GM, Ackermans MT, Serlie MJ, Oozeer R, Derrien M, Druesne A, Van Hylckama Vlieg JET, Bloks VW, Groen AK, Heilig Hghj Zoetendal EG, Stroes ES, de Vos WM, Hoekstra JBL, Nieuwdorp M. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. Gastroenterology 2012;143:913-6.e7

- 94. Ridlon JM, Harris SC, Bhowmik S, Kang D-J, Hylemon PB. Consequences of bile salt biotransformations by intestinal bacteria. Gut Microbes 2016;7:22-39
- 95. Jiang C, Xie C, Li F, Zhang L, Nichols RG, Krausz KW, Cai J, Qi Y, Fang Z-Z, Takahashi S, Tanaka N, Desai D, Amin SG, Albert I, Patterson AD, Gonzalez FJ. Intestinal farnesoid X receptor signaling promotes nonalcoholic fatty liver disease. J Clin Invest 2015;125:386-402
- 96. Ding L, Sousa KM, Jin L, Dong B, Kim B-W, Ramirez R, Xiao Z, Gu Y, Yang Q, Wang J, Yu D, Pigazzi A, Schones D, Yang L, Moore D, Wang Z, Huang W. Vertical sleeve gastrectomy activates GPBAR-1/TGR5 to sustain weight loss, improve fatty liver, and remit insulin resistance in mice. Hepatology 2016;64:760-73
- Neuschwander-Tetri BA, Loomba R, Sanyal AJ, Lavine JE, Van Natta ML, Abdelmalek MF, Chalasani N, Dasarathy S, Diehl AM, Hameed B, Kowdley KV, McCullough A, Terrault N, Clark JM, Tonascia J, Brunt EM, Kleiner DE, Doo E; NASH Clinical Research Network. Farnesoid X nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (FLINT): a multicentre, randomised, placebo-controlled trial. Lancet (London, England) 2015:385:956-65
- 98. Friedman ES, Li Y, Shen T-CD, Jiang J, Chau L, Adorini L, Babakhani F, Edwards J, Shapiro D, Zhao C, Carr RM, Bittinger K, Li H, Wu GD. FXR-dependent modulation of the human small intestinal microbiome by the bile acid derivative obeticholic acid. Gastroenterology 2018;**155**:1741-52.e5.
- 99. Pedersen HK, Gudmundsdottir V, Nielsen HB, Hyotylainen T, Nielsen T, Jensen BAH, Forslund K, Hildebrand F, Prifti E, Falony G, Le Chatelier E, Levenez F, Doré J, Mattila I, Plichta DR, Pöhö P, Hellgren LI, Arumugam M, Sunagawa S, Vieira-Silva S, Jørgensen T, Holm JB, Trošt K, Consortium MHIT, Kristiansen K, Brix S, Raes J, Wang J, Hansen T, Bork P, Brunak S, Oresic M, Ehrlich SD, Pedersen O. Human gut microbes impact host serum metabolome and insulin sensitivity. Nature 2016;535:376-81

- 100. S Lavekar A, V Raje D, Manohar T, A Lavekar A. Role of probiotics in the treatment of nonalcoholic fatty liver disease: a meta-analysis. EIOHG 2017;7:130-37
- 101. Aller R, De Luis DA, Izaola O, Conde R, Gonzalez Sagrado M, Primo D, D La Fuente B, Gonzalez J. Effect of a probiotic on liver aminotransferases in nonalcoholic fatty liver disease patients: a double blind randomized clinical trial. Eur Rev Med Pharmacol Sci 2011;15:1090-5
- 102. Alisi A, Bedogni G, Baviera G, Giorgio V, Porro E, Paris C, Giammaria P, Reali L, Anania F, Nobili V. Randomised clinical trial: the beneficial effects of VSL#3 in obese children with non-alcoholic steatohepatitis. Aliment Pharmacol Ther 2014;39:1276-85
- 103. Eslamparast T, Poustchi H, Zamani F, Sharafkhah M, Malekzadeh R, Hekmatdoost A. Synbiotic supplementation in nonalcoholic fatty liver disease: a randomized, double-blind, placebo-controlled pilot study. Am J Clin Nutr. 2014;99:535-42
- 104. Clinicaltrials.gov. Bethesda (MD). National Library of Medicine (US). Identifier: NCT02721264, Fecal Microbiota Therapy Versus Standard Therapy in NSH related cirrhosis.
- 105. Clinicaltrials.gov. Bethesda (MD). National Library of Medicine (US). Identifier: NCT02469272, Fecal Microbial Transplant (FMT) in Nonalcoholic Steatohepatitis (NASH). A Pilot Study.
- 106. Gilbert JA, Blaser MJ, Caporaso JG, Jansson JK, Lynch SV, Knight R. Current understanding of the human microbiome. Nat Med 2018;24:392-400
- 107. Ranjan R, Rani A, Metwally A, McGee HS, Perkins DL. Analysis of the microbiome: Advantages of whole genome shotgun versus 16S amplicon sequencing. Biochem Biophys Res Commun 2016;469:967-77
- 108. de Muinck EJ, Trosvik P, Gilfillan GD, Hov JR, Sundaram AYM. A novel ultra high-throughput 16S rRNA gene amplicon sequencing library preparation method for the Illumina HiSeq platform. Microbiome 2017;5:68
- 109. Wong VW-S, Won GL-H, Chim AM-L, Chu WC-W, Yeung DK-W, Li KC-T, Chan HL-Y. Treatment of nonalcoholic steatohepatitis with probiotics: a proof-of-concept study. Ann Hepatol 2013;12:256-62