Minireview

Sickle cell vaso-occlusion: The dialectic between red cells and white cells

Nicola Conran¹ b and Stephen H Embury²

¹Hematology Center, University of Campinas-UNICAMP, Barão Geraldo 13083-8, Campinas, SP, Brazil; ²Vanguard Therapeutics, Inc., California, CA 94019, USA

Corresponding author: Nicola Conran. Email: conran@unicamp.br

Impact statement

Sickle cell disease (SCD) is a disease of abnormal blood flow, in which genetic modification of hemoglobin results in red blood cell alterations that are primarily responsible for the acute painful vasoocclusive episodes and, eventually, organ damage that severely diminish the quality and expectancy of life of patients with the disease. Sickle red blood cells (SRBC) are significantly modified by, and contribute to, imbalanced redox physiology, accelerated erythropoiesis, and the rheological changes associated with SCD, thereby playing a fundamental role in vasoocclusive processes. Clear contributions by chronic inflammation, pan-cellular adhesion, thromboinflammation, oxidative stress, and reduced nitric oxide bioavailability have all been established in SCD pathophysiology; moreover, anti-Pselectin therapy and other approaches that abrogate the interactions of the SRBC and other blood cells with the endothelium represent important current and future strategies for treating patients with this devastating disease.

Abstract

The pathophysiology of sickle cell anemia, a hereditary hemoglobinopathy, has fascinated clinicians and scientists alike since its description over 100 years ago. A single gene mutation in the HBB gene results in the production of abnormal hemoglobin (Hb) S, whose polymerization when deoxygenated alters the physiochemical properties of red blood cells, in turn triggering pan-cellular activation and pathological mechanisms that include hemolysis, vaso-occlusion, and ischemia-reperfusion to result in the varied and severe complications of the disease. Now widely regarded as an inflammatory disease, in recent years attention has included the role of leukocytes in vaso-occlusive processes in view of the part that these cells play in innate immune processes, their inherent ability to adhere to the endothelium when activated, and their sheer physical and potentially obstructive size. Here, we consider the role of sickle red blood cell populations in elucidating the importance of adhesion vis-a-vis polymerization in vaso-occlusion, review the direct adhesion of sickle red cells to the endothelium in vaso-occlusive processes, and discuss how red cell- and leukocyte-centered mechanisms are not mutually exclusive. Given the initial clinical success of crizanlizumab, a specific anti-P selectin therapy, we suggest that it is appropriate to take a holistic approach to understanding and exploring the complexity of vaso-occlusive mechanisms and the adhesive roles of the varied cell types, including endothelial cells, platelets, leukocytes, and red blood cells.

Keywords: Acute vaso-occlusive crisis, adhesion, blood flow, erythrocytes, sickle red blood cells, leukocytes

Experimental Biology and Medicine 2021; 246: 1458-1472. DOI: 10.1177/15353702211005392

Introduction

Sickle cell disease (SCD) is ultimately a disease of abnormal blood flow, in which genetic modification of the hemoglobin protein results in red blood cell (RBC) alterations that are primarily responsible for vaso-occlusive mechanisms resultant from cellular activation, cell-cell adhesive interactions, and inflammation. Caused by a single base-pair substitution in the *HBB* gene (encoding β globin), the sickle mutation produces abnormal sickle hemoglobin (HbS) and in homozygosity results in sickle cell anemia (SCA).¹

Compound variations of sickle cell inheritance, denominated as sickle cell disease (SCD), result from the co-inheritance of the beta S allele with other β -globin mutations. ² Polymerization of HbS, under conditions of deoxygenation, leads to the formation of deoxy-HbS fibers in the RBC, altering the properties of these cells and leaving them less deformable and often sickle shaped.³ The ensuing and complex pathophysiology of SCD vaso-occlusion involves multiple molecular mechanisms beyond polymerization and sickling, which we will cover in this review.

The clinical course of SCD is characterized predominantly by painful vaso-occlusive episodes that vary in frequency and severity, can often lead to hospitalization of the patient,⁴ and are the feature of SCD that are the greatest concern of patients with the disease.⁵ Other important and varied complications of the disease, including acute complications and end-organ damage are the result of abnormal blood flow leading to ischemia and consequent inflammatory processes.⁶ Currently available treatments for SCD include hematopoietic stem cell therapy, which can be curative but is not available for all;⁷ hydroxyurea therapy, which is a cytostatic drug with antipolymerization and anti-inflammatory properties;⁸ and drugs that have been developed specifically based on SCD pathophysiology but are relatively new additions to the therapeutic options for SCD, such as L-glutamine supplementation, crizanlizumab, and Voxelotor.9

Vaso-occlusion: What, where, and how?

Vaso-occlusion, central to the pathology of SCD, consists of the obstruction of a blood vessel such that the velocity of blood flow considerably slows or stops. In SCD, vasoocclusive events arise predominantly in the microvasculature and are generally believed to occur mainly in the venules,^{10,11} given the slower flow rate and lower oxygen concentrations in these post-capillary regions, but there is evidence that in some organs vaso-occlusion arises in arterioles, provoking the formation of "bottlenecks" and deregulation of blood flow.¹²

Initiation and propagation of vaso-occlusion

Molecular and physical triggers drive the initiation and propagation of cellular recruitment and aggregation in the vasculature. Known such stimuli in individuals with SCD include infections, hypoxia, dehydration, stress, acidosis, cold temperature, and even pain itself.^{13–15} In mouse models of SCD, inflammatory molecules (such as tumor necrosis factor- α (TNF) cytokine, hemolysis products, lipopolysaccharide), agonists of endothelial and platelet protease active receptors such as thrombin, immunomodulatory epinephrine, and physiological stress (that induces autonomic nervous system reactivity¹⁶) have all been employed to induce and study vaso-occlusive mechanisms.^{12,17–22}

Components of vaso-occlusion

The obstruction of the microvasculature in SCD is a direct result of cellular recruitment to the vascular wall and multicellular aggregate formation in the blood vessel lumen and vascular dysfunction, in conjunction with SRBC sickling and rheological changes.²³ The adhesion of blood cells, that is SRBC as well as leukocytes and platelets, to the vascular wall is the consequence of cell membrane molecular alterations and endothelial cell activation (see Table 1) that are caused by physical changes in HbS-containing RBC^{24,25} and by molecular triggers generated during the inflammatory processes that characterize SCD pathophysiology.²⁶

SRBC and the rheological component of vasoocclusion

As a direct effect of their HbS content, SRBC suffer cellular dehydration, HbS autooxidation, and membrane alterations.²⁷ Dehydration of SRBC results in extreme elevations of intracellular concentrations of HbS and seismic increases in HbS polymerization and sickling.^{28–30} In addition, pathological roles for adenosine signaling, erythrocyte hypoxic metabolic programming, and hypoxia-induced impairments in phospholipid metabolism may all participate in HbS deoxygenation and consequent SRBC sickling.³¹ In turn, SRBC sickling reduces SRBC deformability, increases blood viscosity, and is an iconic effector of impaired blood rheology and therefore SCD vaso-occlusion.^{32,33} Increases in blood viscosity under normal circumstances can elevate nitric oxide (NO) production by the endothelium through shear-stress dependent mechanisms.³⁴ However, as a result of the vascular dysfunction associated with SCD, increases in blood viscosity can instead trigger vaso-occlusive processes in individuals with SCD.^{34,35} Additionally, the autonomic nervous system (ANS), a significant regulator of microvascular blood flow, may be dysfunctional in SCD, many patients demonstrating peripheral vasoconstriction without significant increases in heart rate.³⁶ Thus, modulation of the SRBC microvascular transit time in small vessels by rheological factors¹¹ can facilitate SRBC sickling and obstruction of blood flow in small vessels. Of importance, stoppage of microvasculature blood flow can also alter the adhesion of cells to the blood vessel wall.¹⁴ For instance, all three selectins (P-, E-, and L-selectin) lack adhesivity in the absence of shear stress.37,38

Molecular mechanisms driving vaso-occlusion

Major molecular mechanisms that induce cellular changes and the initiation and propagation of vaso-occlusion include oxidative stress and inflammatory processes. Enhanced NADPH activity and HbS autooxidation in SRBC contribute to the increased production of reactive oxygen species (ROS) in RBC and the oxidation of membrane proteins.^{39–43} Oxidative stress also has a role in vesiculation and membrane shedding⁴⁴ by SRBC, which produce large numbers of microparticles that carry the pro-inflammatory hemoglobin products that play a role in cellular activation and the initiation of vaso-occlusion.^{45,46}

Accordingly, intravascular hemolysis is a major source and generator of inflammatory molecules in SCD. SRBC have a significantly reduced half-life compared to normal RBC,²³ leading to extravascular and intravascular hemolysis, the latter of which accounts for up to 30% of red cell lysis.^{23,47,48} Besides making an important contribution to hemolytic anemia and accelerating erythropoiesis, intravascular hemolysis is a major source of the inflammatory signals that take part in the initiation and propagation of vaso-occlusion in SCD. Initially, oxidative reactions of the hemoglobin that is released from ruptured cells into the circulation inactivates vascular NO, thereby causing endothelial dysfunction and endothelial cell activation as a result of the anti-inflammatory properties of NO. Additionally, cell-free hemoglobin, and its released heme

Cell population	Properties	Role in vaso-occlusion	Major molecular interactions	Refs
Lower density red blood cells	 Increased adhesive properties Secondary recruitment to vascular wall mediated by leukocytes Participation in heterocellular aggregates 	 Adhesion to activated endothelium Physical obstruction of microvessels Augmentation of SRBC transit time 	Major SRBC molecules involved in adhesion: CD36, α4β1 integrin, ICAM-4, PS, P-selectin binding deter- minants, Lu/BCAM	235, 21, 236, 142
Higher density red blood cells	- Less deformable, sickling-prone cells	 Increased likelihood of sickling and trapping in cellular agglomerates Reduction of blood flow 	Fewer molecular interactions, but evidence of Lu/BCAM- dependent interactions	107, 108, 124
Leukocytes	 Increased leukocyte counts in SCD Innate immune cells Adhere to endothelium of the microvasculature, especially when activated Large cells Participation in heterocellular aggregates 	 Potential trigger of physical obstruction of microvessels Augmentation of SRBC transit time Contribute to inflammatory mediator production in SCD 	Major molecules involved in adhesion: Selectins and selectin ligands (PSGL-1, E-selectin ligand), β2 integrins, especially Mac-1 and LFA-1	10, 19, 237
Platelets	 Platelet activation is associated with SCD Participation in leukocyte/platelet/ RBC aggregates Participation in innate immunity 	 Platelets are recruited to the vascular wall when it is damaged Contribute to vaso-occlusive mechanisms in microvessels by activating endothelium and leukocytes, and participating in aggregate formation Major producer of inflamma- tory mediators in SCD 	 Major platelet molecules involved in aggregates and adhesion: P-selectin, α_{iib}β₃, GPIbα, α₅β₁, α₂β₁, ICAM-2 	85, 171, 62, 175
Endothelial cells	 Present surface adhesion molecules when activated Participation in immune responses 	 Tether and capture SRBC, leukocytes, and platelets to the microvascular wall Contribute to inflammatory mediator production and thromboinflammation in SCD 	Major endothelial surface molecules involved in cell capture: P-selectin, E-selectin, ICAM-1, VCAM-1, vWF, CD36, α _ν β ₃ , Glb-IX-V Subendothelial matrix proteins: Laminin, fibronectin, fibrinogen	235, 148, 62, 196, 238,239

Table 1. Involvement of cell populations in the vaso-occlusive process in sickle cell disease.

ICAM: intercellular adhesion molecule; Lu/BCAM: Lutheran/Basal cell adhesion molecule; PS: phosphatidylserine; PSGL-1: P-selectin glycoprotein ligand-1; SCD: sickle cell disease; SRBC: sickle red blood cell; VCAM: vascular cell adhesion molecule; vWF: von Willebrand factor.

group are recognized as erythrocytic damage-associated molecular patterns (DAMPs) that activate innate immune responses with consequent generation of pro-inflammatory cytokine processing and oxidant reactions.⁴⁹ Heme release also plays a role in direct activation of the endothelium, mobilizing von Willebrand factor and translocating P-selectin from storage in Weibel Palade bodies to the endothelial surface, and in the expression of other adhesion molecules that promote cellular recruitment and consequent vaso-occlusion.^{50,51}

SCD also generates a prothrombotic state;^{52–55} platelet activation is an important consequence of hemolysis, due to the reduction in vascular NO (an inhibitor of platelet aggregation), erythrocyte-derived ADP and heme release,^{56–58} and dense SRBC phosphatidylserine exposure,⁵⁹ which enhances thrombin generation.^{60,61}

Activated platelets readily adhere to the vascular wall, participate in heterocellular aggregate formation and contribute to the release of the pro-inflammatory milieu that drives the vaso-occlusive process,^{12,53,62,63} although the overall pathophysiological relevance of platelet activation may be called to question by the consistent failure of plateletinhibiting compounds to ameliorate pain crises.^{64–70}

Finally, a major inducer of vaso-occlusive processes is vaso-occlusion itself. In a vicious cycle, constant vasoocclusive processes in the microvasculature can lead to oxidative damage, ischemia-reperfusion injury, generation of inflammatory responses, and activation of innate immune pathways. Vaso-occlusion disrupts vascular flow, promoting tissue ischemia and, thereby, hypoxia with consequent RBC sickling, DAMP and other inflammatory molecule generation, and further endothelial activation with P- selectin expression.⁷¹⁻⁷⁵ In turn, the reoxygenation of the vasculature, upon resumption of blood flow,⁷⁶ generates more damaging ROS,^{76,77} which together with enhanced inflammatory pathway activation^{26,78,79} generates further vaso-oclusive processes. Hypoxia itself is used as a trigger for vaso-occlusion in murine models of SCD.^{75,80}

.....

Are vaso-occlusive mechanisms the same in different tissues and organs?

It is probable that the profile of triggering and propagating mechanisms and also cellular involvement may differ depending on the organ or tissue. In the kidney, for example, vaso-occlusion may be more frequent in the renal medulla, which is more hypoxic, hyperosmotic, and acidic⁸¹ than in the hyperperfused renal cortex,⁸² although these sickling dynamics may be changed by acute kidney injury.⁸² The bone marrow and the liver both have hypoxic and sinusoidal environments that promote SRBC sickling,^{83,84} and cellular obstructions comprised of erythroid and myeloid cells and nucleated erythroid precursor cells have been reported in the bone marrow of SCD mice.⁸³ In contrast to other organs, the lung is highly oxygenated and, at least in mice with SCD, a defining feature of pulmonary vaso-occlusion may be the formation of occlusive neutrophil-platelet aggregates in the arterioles rather than in the venules.¹² The expression of the P-selectin adhesion molecule on the surface of endothelial cells and platelets appears to play a significant role in lung vaso-occlusion,⁸⁵ and may contribute to acute chest syndrome,^{12,86} one of the most common causes of death in patients with SCD.

Insights from differential roles of SRBC populations in vaso-occlusion

In addition to the paradigmatic HbS polymerization and SRBC sickling considered above, other SRBC changes are important to SCD,^{4,87} including abnormalities of redox, hemoglobin stability, oxygen affinity, metabolism, membrane integrity, adhesivity, and cation homeostasis. A focus on SRBC dehydration and its effects on HbS polymerization and SRBC sickling provides insights into other vaso-occlusive processes resultant from abnormal cation homeostasis.

The broad clinical heterogeneity among patients with SCD is commonly attributed to variability in the amounts of fetal hemoglobin (HbF) within SRBC,^{88,89} A separate powerful determinant of SRBC pathobiology is abnormal SRBC dehydration,³⁰ which increases cell density, intracellular concentrations of HbS (mean cellular Hb concentration; MCHC),²⁸ HbS polymerization, and cell sickling.²⁹ Density of RBC, as measured by density gradient separation,⁹⁰ is much greater and more heterogeneous in SRBC than in normal RBC.^{91,92} Those SRBC that are most dehydrated are the most dense and have the highest MCHC,²⁸ which decreases the deformability of even oxygenated SRBC as measured by laser scattering viscometry,⁹³ and increases immensely the rate of HbS polymerization in deoxygenated SRBC.⁹⁴ These relationships are essential tenets of the polymerization paradigm that presumes to

account for the totality of SCD clinical features. Leading authorities have attributed all the clinical heterogeneity of SCD to differences in SRBC density because of the powerful effect that MCHC has on HbS polymerization and SRBC sickling;⁹⁵ indeed the densest SRBC are the most sicklingprone. This paradigm predicts a direct association between the number of dense SRBC and the clinical severity of SCD.^{96,97}

However, attribution of the severity of all facets of disease to HbS polymerization is challenged by the indirect relationship of the severity of hemolytic anemia and vasoocclusive crises (VOC). In a large study of the natural history of SCD, milder anemia was associated with more frequent VOC.⁹⁸ Similar associations of milder anemia and more frequent VOC were observed with coexistent α thalassemia and SCD^{99,100} and in therapeutic trials of the cell-hydrating, polymerization-inhibiting drug Senicapoc,¹⁰¹ which together indicated opposite effects of polymerization on the degree of hemolytic anemia and the frequency of VOC.

One astute analysis concluded that the initiation of VOC is not directly related to the major determinants of HbS polymerization, SRBC density or HbF levels.¹⁰² A seminal report had found no correlation of the number of dense SRBC with the frequency of VOC,⁹⁹ which subsequently was supported by the finding of a paradoxical association of α thalassemia, a coinherited condition associated with fewer dense SRBC,⁹² with increased VOC.¹⁰⁰ These observations demonstrated that SCD could not be understood by simple extrapolations from polymerization paradigms alone. A crucial discovery critical to elucidating the relevance of disparate sickle cell pathophysiologies was that low-density SRBC populations had greater adhesivity in micropipet and flow-adhesion assays compared to high-density SRBC.^{103–105}

Designation of the most-dense SRBC as the most sickling-prone, least adhesive subpopulation and of the least dense SRBC as the most adhesive, least sicklingprone subpopulation provided the foundation for reinterpreting the polymerization paradigm. For example, a study in children with SCD found a positive correlation between the fraction of better hydrated, more deformable SRBC and the incidence of VOC,²⁷ and a study in adults with SCD found a positive correlation between incidence of VOC and the fraction of better hydrated, more deformable SRBC and a negative correlation with the percentage of more dense SRBC.¹⁰⁶ Additional evidence that polymerization alone did not explain VOC was provided by the discovery that patients with greater numbers of more deformable, less dense SRBC had more frequent VOC compared to those with greater numbers of less deformable, more dense SRBC.¹⁰⁷

A touchstone study of different sickle cell pathophysiologies and their effects on the flow of human SRBC populations in a rat vascular flow system discovered that the most dense, sickling-prone SRBC fraction neither stuck within the vasculature nor caused stoppage of flow, that the least dense, least sickling-prone SRBC fraction stuck within vessels but did not cause stoppage of flow, and that serial infusion of the two fractions with the least dense followed by the most dense resulted in initial adhesion of the former followed by complete stoppage of flow by the latter.¹⁰⁸ These findings indicated that adhesion of stickier SRBC is the initiatory step in vaso-occlusion and that physical trapping of more sickling-prone SRBC is the second step.

Crucial to these considerations are the delays that occur in SRBC deoxygenation and HbS polymerization during transit through the low oxygen environment of small blood vessels, which usually delay sickling until SRBC have entered large blood vessels where they do no harm, thereby mitigating the detrimental effects of occluded blood flow in small blood vessels.^{94,109} Adhesion of the less dense, stickier SRBC prolongs their transit time as well as the transit time of nonadherant sickling-prone SRBC, resulting in polymerization and sickling in small vessels where occlusion of blood flow elicits tissue damage.

Correlations of VOC severity with adhesive properties of SRBC have established the relative importance of SRBC adhesion compared to HbS polymerization on SCD severity.

Adhesive interactions of SRBC with the endothelium

Abnormal blood flow is the cause of most of the major clinical consequences of SCD.^{94,110-113} Even asymptomatic patients have impaired blood flow, and exacerbations thereof result in acute VOC.¹¹⁴⁻¹¹⁶ The frequency of VOC correlates with mortality in adult patients,⁹⁸ is a major determinant of quality of life of patients with SCD,¹¹⁷ and is the feature of disease about which patients have the greatest concern.⁵

Abnormal sickle cell blood flow is fundamentally related to adhesion of SRBC to the vascular endothelium. The first studies of SRBC adhesion revealed that SRBC have greater adhesion to cultured endothelial cells compared to normal RBC,¹¹⁸ and subsequent studies disclosed that the adhesivity of SRBC correlated with the vaso-occlusive severity of patients with SCD.¹¹⁹ Further perspective for this observation was derived from the appreciation of vaso-occlusion as a two-step process initiated by SRBC adhesion to the vascular endothelium.¹⁰⁸

SRBC adhesion mediates chronic impairment of sickle cell blood flow in unperturbed experimental animals and in asymptomatic patients with $\text{SCD}^{21,120}$ and initiates acute vascular occlusion in experimental animals^{21,75} and in patients with SCD.^{27,99,106,107}

Numerous adhesion molecules on SRBC and endothelial cells and in the plasma, including integrins, members of the immunoglobulin superfamily, and others, participate in SRBC adhesion (Table 1).¹²¹⁻¹²³ Recently, oxidation of RBCs was found to induce post-translational modification of the Lu/BCAM adhesion molecule on the surface of dense SRBCs, signifying that this molecule could mediate adhesion of this specific RBC population to laminin on blood vessel walls, even in the absence of sickling.¹²⁴ Many of the molecules involved in the tethering of SRBC to the endothelium also mediate the molecular cascade of leukocyte-endothelial adhesion during inflammation,^{125,126} which is an apt model for SRBC adhesion. That cascade is

initiated by and dependent upon the endothelial cell adhesion molecule, P-selectin.^{127,128} Of particular importance to the inflammatory processes is the P-selectin dependence of slow rolling adhesion of leukocytes, which is a necessary antecedent for their firm adhesion and extravasation.^{129–131}

Endothelial P-selectin is critical to the adhesion of SRBC to vascular endothelium

The expression of P-selectin on platelets and endothelial cells is generally regarded as acute and transient following exposure to agonists.¹³¹ However, in certain systems,¹³² including sickle cell mouse models ¹³³ and asymptomatic patients with SCD,¹²⁰ P-selectin is expressed chronically on the vascular endothelium. Mechanisms of chronic P-selectin expression include increased transcription^{134,135} and reutilization involving invagination and recycling of the molecule onto the cell surface.¹³⁶ Chronically elevated P-selectin expression on the surface of the vascular endothelium in sickle cell mouse models has been shown to be inducible to higher levels by use of an endothelial cell agonist.²¹

P-selectin is the only selectin that has been shown to support the direct adhesion of SRBC to endothelial cells,¹³⁷ and unpublished results from the Vanguard Therapeutics, Inc. laboratory have shown that SRBC adhere *in vitro* to P-selectin but not E- or L-selectin. Under both static and flow conditions SRBC adhere abnormally to recombinant P-selectin and to cultured endothelial cells that have been activated to express P-selectin.^{137,138} Most importantly, P-selectin has been demonstrated to support the initial contact and slow rolling adhesion of SRBC.¹³⁸

The adhesion of SRBC to endothelial cells creates a positive feedback mechanism by increasing intraendothelial ROS levels,¹³⁹ which in turn induces P-selectin transcription¹⁴⁰ and expression of P-selectin on the endothelial cell surface.¹⁴¹ P-selectin binding determinants are found in greater amounts on SRBC membranes than on normal RBC membranes, contain sialyl LewisX, exist on glycoproteins and glycolipids, and exist on both sickle-reticulocytes and older SRBC.¹⁴² SRBC adhesion and increased intraendothelial ROS also induce transcription and expression of VCAM-1,^{143,144} which may account for the elevated plasma levels in patients with SCD of soluble VCAM-1 (sVCAM-1), a marker of endothelial injury, activation, or stimulation.^{145–147}

Leukocytes as participants in the vaso-occlusive process

The discovery of leukocyte adhesion in a mouse model of SCD¹⁴⁸ has led to an avalanche of research into this phenomenon despite unmistakable digressions from physiologic and pathophysiologic norms in this original report. These include a 1:4 capture/minute ratio of SRBC vs. WBC, failure to detect vaso-occlusion, and an elevated baseline serum TNF concentration of 15 pg/mL for this mouse model¹⁴⁹ even before treatment with an additional 500,000 pg. of TNF. While fascinating, this observation does

not nullify established evidence that SRBC adhere directly to the vascular endothelium in vaso-occlusion *in vivo* independently of leukocytes.^{21,150-153}

While leukocytosis had long been known to be a common feature of SCD,¹⁵⁴ a role for leukocytes in the pathophysiology of SCD and vaso-occlusive processes was not appreciated until observations of significant associations between white cell counts with disease severity and early SCD-related death.¹⁵⁵⁻¹⁵⁷ In addition to their role in inflammatory mechanisms, leukocytes play a substantial mechanical and obstructive role in vaso-occlusive process. Ex vivo assays first demonstrated the increased adhesive properties of neutrophils, which adhere abnormally to extracellular matrix components and endothelial cell lavers.158-160 Leukocytes are large (human neutrophils are about 12-15 µm in diameter) rigid cells, and stimuli that elicit their interactions with the walls of microvessels can slow blood flow, thereby increasing the RBC transit time in the vessel and promoting RBC sickling; in vivo studies show that the percentage of temporarily static leukocytes and the time of their stasis in a vessel correlates inversely to RBC velocity.¹⁶¹ Microvascular imaging of sickle cell mice⁷⁵ detected that adherent leukocytes can capture circulating SRBC, further exacerbating vessel obstruction, 148,162 and an in vitro flow model of vaso-occlusion confirmed the capacity of human SCD leukocytes adherent on endothelial cell layers to capture SRBC.¹⁶³ A positive feedback is suggested by proximity of the vessel wall and activated leukocytes augmenting SRBC adhesion to the endothelium¹⁶⁴ and stimulated human SRBC promoting leukocyte recruitment to the vessel wall.^{165,166} The molecular interactions that mediate the recruitment and adhesion of leukocytes to the vessel wall largely involve endothelial surface selectins (P-selectin and E-selectin) and ICAM-1, which bind with selectin ligands and β_2 integrins (Mac-1 and LFA-1) on the leukocyte surface (see Table 1).^{19,74,167,168}

Neutrophils are the most abundant leukocytes and circulate in an activated state in patients with SCD, displaying an increased expression and functionality of the B2 integrins on their surface, due in part to endothelin B receptor mediated signaling and TNF stimulation.^{169,170} In addition to their increased propensity for adhesion, neutrophils form heterocellular aggregates with platelets via interac-tions involving the neutrophil surface;^{10,171-173} a subset of effector neutrophils that express high levels of surface CXCR4¹⁷⁴ and have enhanced Mac-1 integrin activity play a major role in the formation of these aggregates in SCD.¹⁷⁵ Neutrophils also form aggregates with SRBC, sometimes involving platelet "bridges", ¹⁷¹ and SCD mice that are deficient in the Mac-1 integrin have a depletion of neutrophil-RBC aggregates and reduced vaso-occlusive processes in response to trauma and cytokine stimulation.¹⁷⁶ The importance of heterocellular aggregate formation in SCD pathophysiology is illustrated by the aforementioned role that neutrophil-platelet aggregates appear to play in vasoocclusive mechanisms in the pulmonary circulation of SCD mice.^{12,177} A role for activated monocytes also is implicated in SCD pathophysiology,¹⁴⁹ as supported by reports of endothelial cell activation by SCD monocytes and the formation of aggregates between this leukocyte type and

both platelets¹⁷⁸ and RBC.^{179,180} In contrast, patrolling monocytes have actually been shown to protect against vaso-occlusive processes by scavenging endothelial-adherent SRBC.¹⁸¹

Finally, leukocytes are key to mounting innate immune responses and the generation of many of the molecules, including cytokines, chemokines, and growth factors, that participate in the inflammatory mechanisms that drive vaso-occlusive processes in sickle cell disease.26,149 Neutrophils in particular are critical innate immunity effector cells, and the release of neutrophil extracellular traps in response to heme molecules and other inflammatory stimuli¹⁸² may be a factor in painful vaso-occlusive episodes and acute chest syndrome.¹⁸³ Individuals with SCD also present intestinal injury and increased gut permeability that may augment intestinal barrier translocation of lipopolysaccharide,¹⁸⁴ a bacterial membrane component and major activator of innate immune signaling. It may be relevant that the microbiota appears to regulate neutrophil heterogeneity and aging in SCD, while the depletion of the microbiota in mice with SCD decreases vaso-occlusive events.174

As such, the inflammatory nature of leukocytes, their adhesion to the endothelium of the microvasculature, and their participation in SRBC adhesion and heterocellular aggregate formation appear to contribute to increasing the SRBC transit time in small vessels, thereby triggering the sickling of dense SRBC to promote vaso-occlusion.

Therapeutic approaches for reducing both red and white cell adhesive mechanisms in SCD

As mentioned, current therapeutic options for SCD are limited. Approaches for reducing/abolishing HbS production and polymerization include HSCT, gene therapies and the recently—USA Food and Drug Administration (FDA) approved Hb-affinity modulator, Voxelotor.^{185–187} Herein, we discuss some of the drugs and biological agents currently in development or use for SCD in the context of the pathophysiological mechanisms described herein.

Hydroxyurea therapy

Hydroxyurea is frequently used as therapy for SCD as it reduces the incidence of hospitalization, acute pain, acute chest syndrome, and transfusion frequency, and prolongs life among other effects in patients with SCD.¹⁸⁸⁻¹⁹² This cytostatic compound acts principally by inducing the production of HbF, thereby reducing HbS polymerization.¹⁹³ However, hydroxyurea also has extensive effects on both cellular adhesivity and inflammatory pathways, due both to the downstream effects of HbS polymerization inhibition and due to its direct anti-inflammatory, myelosuppressive, and NO-dependent signaling effects.^{189,194,195} Of note, hydroxyurea therapy is associated with significant reductions in the expression and activity of adhesion molecules on the surface of reticulocytes, SRBCs, leukocytes, and the endothelium in SCD.^{196–201} However, hydroxyurea therapy inexplicably has been associated with an extraordinary

degree of patient noncompliance, with 85.9% of initial prescriptions not being filled in one study.²⁰²

Anti-adhesion therapies

We have mentioned that P-selectin expression on activated endothelial cells plays a major role in tethering rolling SRBC and leukocytes in SCD, and it also contributes to the formation of heterocellular aggregates involving platelets.^{12,74,137} Crizanlizumab is a monoclonal antibody that neutralizes the activity of P-selectin. In a phase 2 trial, higher doses of crizanlizumab reduced the incidence of SCD VOC,²⁰³ and this biological agent is now FDA-approved as Adakveo for use in patients aged 16 years or older with SCD but with the requirement of extensive post-marketing clinical testing for the presence and immunogenic effect of anti-drug antibodies (ADAb). Informal high-level comparison of median VOC frequency, time to first VOC, and time to second VOC has determined that Adakveo is at least as effective at reducing the frequency of VOC as was hydroxyurea in its original study.^{203,204} The use of Adakveo requires administration by monthly intravenous infusion which is a challenge to patient compliance, is susceptible to inducing ADAb that have the potential to neutralize drug activity or cause serious reactions such as anaphylaxis, and requires ready venous access that is often unavailable in patients with SCD.

The pan-selectin antagonist, Rivipansel (GMI-1070), was developed with the aim of treating, not preventing, VOC in SCD by decreasing leukocyte interactions with the endothelium and the formation of heterocellular leukocyte aggregates. Its blocking activity is 100-fold less for P- and L-selectin than for E-selectin. The aim of treating existent VOC by blocking the initiatory vaso-occlusive molecules, selectins, is challenged by the action of numerous vasoocclusive mechanisms by the time symptoms of VOC are detected. Phase 2 studies demonstrated that GMI-1070 improved clinical outcomes including time to resolution of pain crisis, opioid use and length of hospital stay.^{205–207} However, a phase 3 clinical trial (NCT02187003) that further assessed the effectiveness and safety of Rivipansel for the treatment of pain crisis in hospitalized SCD patients failed to achieve proposed study outcomes, namely reductions in the time patients spent in hospital or opioid use for pain management. A subsequent retrospective analysis revealed that patients who received the agent earliest in their VOC may have had effective reductions in duration of therapy,²⁰⁸ which has rekindled interest in its use for treating sickle cell VOC.

Modified heparins have anti-adhesive properties that show potential for use in SCD therapy. Low molecular weight heparins (LMWHs) can inhibit collagen-induced platelet activation²⁰⁹ and the adhesion of SRBC to endothelial VCAM-1 by blocking erythrocyte VLA-4 integrin function and by blocking P-selectin mediated adhesion of SRBC.^{120,210,211} Similarly, sevuparin a modified heparin, which retains heparin's anti-adhesive properties while decreasing its anticoagulant activity, also inhibits the adhesion of human SRBC to endothelial cell layers and prevents vaso-occlusion in a mouse model of SCD.²¹² The efficacy and safety of this molecule was recently evaluated in a phase 2 study for the treatment of acute VOC in subjects with SCD (NCT02515838), although to our knowledge results have not yet been published. Other anti-cell adhesion approaches that have been evaluated in SCD include single dose intravenous immunoglobulin, and poloxamer 188.^{205,213–215}

Anti-inflammatory therapies

By reducing the inflammatory milieu capable of cellular activation, anti-inflammatory drugs may be important for abrogating the adhesive interactions of SRBC, leukocytes, and platelets with each other and the endothelium. A vast number of anti-inflammatory approaches are currently in pre-clinical and clinical development for SCD. Blocking the action of the sentinel cytokine, TNF, could be particularly useful for reducing endothelial activation and therefore the initiation and propagation of vaso-occlusion in SCD. In SCD mice, etanercept, a fusion protein that binds to and inhibits the action of TNF, ameliorated endothelial activation, blood biomarkers of inflammation, and hypoxia/reoxygenation-triggered vaso-occlusive processes, among other beneficial effects.¹⁴⁹

Neutralization of cell-free hemoglobin and heme may counteract the effects of hemolysis; infusions of haptoglobin or hemopexin have successfully prevented inflammatory mechanisms, vaso-occlusion, and acute chest syndrome onset in SCD mouse models.^{18,216} CSL889, a plasma-derived hemopexin therapy, has recently received FDA and European Commission designation as an orphan drug and a phase 1 clinical trial of CSL889 hemopexin infusion therapy is currently underway (NCT04285827).

Preclinical studies have shown that the amplification of NO-cyclic guanosine monophosphate (cGMP)-dependent signaling using cGMP-modulating drugs, in combination with hydroxyurea, or not, can abrogate endothelial activation, leukocyte recruitment and leukocyte-RBC interactions in mice with SCD.^{195,217,218} However, a recent phase 2 clinical trial in sickle cell anemia for olinciguat, a stimulator of soluble guanylate cyclase (sGC; the enzyme that catalyzes cGMP production), was discontinued for failure to achieve study endpoints (NCT03285178).²¹⁹ Inhibitors of phosphodiesterase 9 (PDE9; an enzyme that specifically degrades intracellular cGMP and is highly expressed in hematopoietic cells)²²⁰ have completed or are in phase 1/2 clinical trials for use in SCD individuals (NCT02114203; NCT04474314); IMR-687 is said to lack the potential side effects of other PDE9 inhibitors because it does not cross the blood brain barrier and it also increases HbF production in erythroid cells.^{221,222} Other anti-inflammatory drugs have been recently reviewed.²⁶

Antioxidants

Control of the intracellular oxidative stress generated in SCD may diminish membrane instability²²³ and cellular activation⁴⁹ and consequently vaso-occlusive processes. L-glutamine supplementation was found to ameliorate the redox potential of SRBC^{224,225} and was approved by the FDA in 2017 for use in treating SCD in the form of

Endari L-glutamine oral powder following findings of a randomized, double-blind, placebo-controlled, multicenter clinical trial that showed that twice-daily administration of this amino acid reduced the frequency of sickle cell crises and hospitalization;²²⁶ however, hurdles in the initiation and adherence to L-glutamine have been reported.²²⁷

NADPH oxidase repression may also represent potential for the reduction of SRBC oxidative stress. Manganese porphyrins are low-molecular-weight synthetic nonpeptides that are commonly known as superoxide dismutase mimics; the administration of a single dose of these redox-active manganese porphyrins to humanized SCD mice, following the establishment of vaso-occlusive processes, was found to reverse and reduce the adhesion of SRBCs and leukocytes to venules, thus restoring blood flow and increasing the survival rate of mice.²²⁸

Finally, small molecule activators of pyruvate kinase-R in SRBC have been found to increase ATP levels and decrease 2,3-DPG levels in RBC,²²⁹ which is predicted to retard SRBC sickling and to mitigate levels of anemia in patients with SCD. In early clinical studies, FT-4202 reduced anemia in 86% of patients and Mitapivat improved anemia in 55%.^{230,231}

Final considerations

Recent decades have witnessed an explosion of evidence of the pathophysiologies of vaso-occlusive mechanisms in SCD, with the recognition of the fundamental roles for chronic inflammation, endothelial and leukocyte stimulation, activation of platelets and coagulation, and alterations in NO bioavailabilty. The critical role of SRBC, which are more affected by imbalanced redox physiology, accelerated erythropoiesis, and rheological modifications than by inflammatory pathways is essential to vaso-occlusive processes. At the end of the day, similar inflammatory processes and cellular activation are observed in other pathologies that display vascular inflammation,²³² but the overwhelming pathological microvascular obstruction seen in SCD also requires participation of the altered SRBC and sickling. We recognize that, in what is commonly referred to as the vicious cycle of SCD pathophysiology, inflammatory triggers generated by SRBC destruction and ischemiareperfusion injury appear to be largely responsible for fueling the cellular alterations that result in the adhesion of both less dense SRBC and activated leukocytes to the adhesion molecule-presenting endothelium and heterocellular aggregate formation in microvessels. The ensuing mechanical obstruction of the vessel alters blood rheology, increases the SRBC transit time, and incurs trapping of the dense, less deformable SRBC population that are prone to sickling and, vaso-occlusion (Figure 1). We suggest that future studies could focus on further understanding how this mechanism adapts in the microvasculature of different organs with different characteristics and in response to different molecular and mechanical stimuli.

Drugs aiming to prevent vaso-occlusive processes in SCD, other than by inhibiting the primary HbS polymerization event, should aim to abrogate leukocyte and SRBC adhesion to the activated endothelial surface with a view to preventing the slowing of blood flow in small vessels and dense SRBC trapping. Indeed, the biological agent crizanlizumab inhibits the activity of P-selectin on activated endothelium and platelets, thereby potentially reducing the adhesion of SRBC and leukocytes to the vessel walls and the formation of heterocellular aggregates, 21,85,233 and has displayed translational success to the clinic. However, it may be that future approaches should ensure not just the prevention of cellular recruitment to the endothelium and cell-cell interactions to prevent VOC, but also the reversal of these interactions to provide badly needed approaches for the treatment of acute VOC. Additionally,



Figure 1. Proposed two-step mechanism for sickle red blood cell involvement in vaso-occlusive processes. Inflammatory mechanisms, caused by intravascular hemolysis and processes of ischemia-reperfusion, among other factors, lead to endothelial cell, leukocyte and platelet activation. Activated endothelium presents multiple adhesion molecules on its surface, including P-selectin, E-selectin and ICAM-1, which mediate cellular tethering to the vascular wall. Less-dense, deformable, less sickling-prone and more adhesive sickle red blood cells (SRBCs) are recruited to activated endothelium, as are activated leukocytes, especially neutrophils. In the microvasculature, especially venules, the mechanical obstruction of the vessel by the adhered SRBCs, and adhered leukocytes, increases the transit time of other SRBCs in the vessel, leading to the trapping of the denser sickling-prone SRBC population, as well as heterocellular aggregates, in these cellular agglomerates. Extensive cellular trapping associated with rheological alterations may result in local SRBC sickling, blood flow arrest and, therefore, vaso-occlusion. (A color version of this figure is available in the online journal.)

the chronic organ damage that is observed in individuals with SCD²³⁴ as they age, is an increasing concern and reducing both vaso-occlusive mechanisms and associated damaging inflammatory processes may be necessary for complete therapy of this population. Such approaches may potentially combine anti-inflammatory approaches with anti-adhesive approaches, or even approaches that target other cellular and adhesive interactions.

AUTHORS' CONTRIBUTIONS

NC and SE wrote the article and have read and approved the final article.

ACKNOWLEDGMENTS

The authors thank Ana Carolina Andrade Vitor Kayano for assistance with the preparation of Figure 1. We are particularly grateful to Dr Bob Hebbel for his thoughtful review of the article.

DECLARATION OF CONFLICTING INTERESTS

NC receives research funding from Novartis Pharma AG. SE is an employee of Vanguard Therapeutics, Inc.

FUNDING

NC acknowledges principal sources of funding from São Paulo Research Foundation (FAPESP grants # 2018/08010-9, 2014/00984-3).

ORCID ID

Nicola Conran D https://orcid.org/0000-0001-5726-7919

REFERENCES

- Pauling L, Itano HA, Singer SJ, Wells IC. Sickle cell anemia a molecular disease. *Science* 1949;110:543–48
- 2. Habara A, Steinberg MH. Genetic basis of heterogeneity and severity in sickle cell disease. *Exp Biol Med* 2016;**241**:689–96
- Henry ER, Cellmer T, Dunkelberger EB, Metaferia B, Hofrichter J, Li Q, Ostrowski D, Ghirlando R, Louis JM, Moutereau S, Galacteros F, Thein SL, Bartolucci P, Eaton WA. Allosteric control of hemoglobin S fiber formation by oxygen and its relation to the pathophysiology of sickle cell disease. *Proc Natl Acad Sci USA* 2020;**117**:15018–27
- Kato GJ, Piel FB, Reid CD, Gaston MH, Ohene-Frempong K, Krishnamurti L, Smith WR, Panepinto JA, Weatherall DJ, Costa FF, Vichinsky EP. Sickle cell disease. *Nat Rev Dis Primers* 2018;4:18010
- FDA. The voice of the patient. In: Public meeting on sickle cell disease patient-focused drug development, 2014, pp. 1–30.
- Howard J, Thein SL. Optimal disease management and health monitoring in adults with sickle cell disease. *Hematology Am Soc Hematol Educ Program* 2019;2019:505–12
- Dalle JH. Hematologist and transplant physicians: how and where to meet for the best of sickle cell disease patients? *Hematol Oncol Stem Cell Ther* 2020;13:58–60
- McGann PT, Ware RE. Hydroxyurea therapy for sickle cell anemia. Expert Opin Drug Saf 2015;14:1749–58
- Matte A, Cappellini MD, Iolascon A, Enrica F, De Franceschi L. Emerging drugs in randomized controlled trials for sickle cell disease: are we on the brink of a new era in research and treatment? *Expert Opin Investig Drugs* 2020;29:23–31

 Hidalgo A, Chang J, Jang JE, Peired AJ, Chiang EY, Frenette PS. Heterotypic interactions enabled by polarized neutrophil microdomains mediate thromboinflammatory injury. *Nat Med* 2009;15:384–91

- Coates TD, Chalacheva P, Zeltzer L, Khoo MCK. Autonomic nervous system involvement in sickle cell disease. *Clin Hemorheol Microcirc* 2018;68:251–62
- Bennewitz MF, Jimenez MA, Vats R, Tutuncuoglu E, Jonassaint J, Kato GJ, Gladwin MT, Sundd P. Lung vaso-occlusion in sickle cell disease mediated by arteriolar neutrophil-platelet microemboli. *JCI Insight* 2017;2:e89761
- Mueller BU, Brugnara C. Prevention of red cell dehydration: a possible new treatment for sickle cell disease. *Pediatr Pathol Mol Med* 2001;20:15–25
- Veluswamy S, Shah P, Denton CC, Chalacheva P, Khoo MCK, Coates TD. Vaso-occlusion in sickle cell disease: is autonomic dysregulation of the microvasculature the trigger? J Clin Med 2019;8
- Stuart J, Johnson CS. Rheology of the sickle cell disorders. Baillieres Clin Haematol 1987;1:747–75
- Shah P, Khaleel M, Thuptimdang W, Sunwoo J, Veluswamy S, Chalacheva P, Kato RM, Detterich J, Wood JC, Zeltzer L, Sposto R, Khoo MCK, Coates TD. Mental stress causes vasoconstriction in subjects with sickle cell disease and in normal controls. *Haematologica* 2020;**105**:83–90
- Xu C, Lee SK, Zhang D, Frenette PS. The gut microbiome regulates psychological-stress-induced inflammation. *Immunity* 2020;53:417–28
- Ghosh S, Adisa OA, Chappa P, Tan F, Jackson KA, Archer DR, Ofori-Acquah SF. Extracellular hemin crisis triggers acute chest syndrome in sickle mice. J Clin Invest 2013;123:4809–20
- Turhan A, Jenab P, Bruhns P, Ravetch JV, Coller BS, Frenette PS. Intravenous immune globulin prevents venular vaso-occlusion in sickle cell mice by inhibiting leukocyte adhesion and the interactions between sickle erythrocytes and adherent leukocytes. *Blood* 2004;103:2397–400
- Zennadi R, De Castro L, Eyler C, Xu K, Ko M, Telen MJ. Role and regulation of sickle red cell interactions with other cells: ICAM-4 and other adhesion receptors. *Transfus Clin Biol* 2008;15:23–28
- Embury SH, Matsui NM, Ramanujam S, Mayadas TN, Noguchi CT, Diwan BA, Mohandas N, Cheung AT. The contribution of endothelial cell P-selectin to the microvascular flow of mouse sickle erythrocytes in vivo. *Blood* 2004;**104**:3378–85
- Sparkenbaugh EM, Chen C, Brzoska T, Nguyen J, Wang S, Vercellotti GM, Key NS, Sundd P, Belcher JD, Pawlinski R. Thrombin activation of PAR-1 contributes to microvascular stasis in mouse models of sickle cell disease. *Blood* 2020;**135**:1783–87
- Steinberg MH. Overview of sickle cell anemia pathophysiology. In: Costa FF and Conran N (eds), Sickle cell anemia: from basic science to clinical practice. Switzerland: Springer International, 2016, pp. 49–75
- Ney PA, Christopher MM, Hebbel RP. Synergistic effects of oxidation and deformation on erythrocyte monovalent cation leak. *Blood* 1990;75:1192–98
- Hebbel RP, Steinberg MH, Eaton JW. Erythrocyte calcium abnormalities in sickle cell disease. Prog Clin Biol Res 1981;51:321–32
- Conran N, Belcher JD. Inflammation in sickle cell disease. Clin Hemorheol Microcirc 2018;68:263–99
- Lande WM, Andrews DL, Clark MR, Braham NV, Black DM, Embury SH, Mentzer WC. The incidence of painful crisis in homozygous sickle cell disease: correlation with red cell deformability. *Blood* 1988;72:2056–59
- Clark MR, Unger RC, Shohet SB. Monovalent cation composition and ATP and lipid content of irreversibly sickled cells. *Blood* 1978;51:1169–78
- Hofrichter J, Ross PD, Eaton WA. Kinetics and mechanism of deoxyhemoglobin S gelation: a new approach to understanding sickle cell disease. *Proc Natl Acad Sci USA* 1974;71:4864–88
- Brugnara C. Sickle cell dehydration: pathophysiology and therapeutic applications. *Clin Hemorheol Microcirc* 2018;68:187–204

- Adebiyi MG, Manalo JM, Xia Y. Metabolomic and molecular insights into sickle cell disease and innovative therapies. *Blood Adv* 2019;3:1347-55
- Embury SH. The clinical pathophysiology of sickle cell disease. Annu Rev Med 1986;37:361–76
- Valdez JM, Datta YH, Higgins JM, Wood DK. A microfluidic platform for simultaneous quantification of oxygen-dependent viscosity and shear thinning in sickle cell blood. *APL Bioeng* 2019;3:046102
- 34. Nader E, Skinner S, Romana M, Fort R, Lemonne N, Guillot N, Gauthier A, Antoine-Jonville S, Renoux C, Hardy-Dessources MD, Stauffer E, Joly P, Bertrand Y, Connes P. Blood rheology: key parameters, impact on blood flow, role in sickle cell disease and effects of exercise. *Front Physiol* 2019;10:1329
- Vazquez BY, Vazquez MA, Jaquez MG, Huemoeller AH, Intaglietta M, Cabrales P. Blood pressure directly correlates with blood viscosity in diabetes type 1 children but not in normals. *Clin Hemorheol Microcirc* 2010;44:55–61
- 36. Chalacheva P, Kato RM, Shah P, Veluswamy S, Denton CC, Sunwoo J, Thuptimdang W, Wood JC, Detterich JA, Coates TD, Khoo MCK. Sickle cell disease subjects have a distinct abnormal autonomic phenotype characterized by peripheral vasoconstriction with blunted cardiac response to Head-Up tilt. *Front Physiol* 2019;**10**:381
- Finger EB, Puri KD, Alon R, Lawrence MB, von Andrian UH, Springer TA. Adhesion through L-selectin requires a threshold hydrodynamic shear. *Nature* 1996;**379**:266–69
- Lawrence MB, Kansas GS, Kunkel EJ, Ley K. Threshold levels of fluid shear promote leukocyte adhesion through selectins (CD62L,P,E). J Cell Biol 1997;136:717–27
- George A, Pushkaran S, Konstantinidis DG, Koochaki S, Malik P, Mohandas N, Zheng Y, Joiner CH, Kalfa TA. Erythrocyte NADPH oxidase activity modulated by Rac GTPases, PKC, and plasma cytokines contributes to oxidative stress in sickle cell disease. *Blood* 2013;**121**:2099–107
- Hebbel RP. The sickle erythrocyte in double jeopardy: autoxidation and iron decompartmentalization. *Semin Hematol* 1990;27:51–69
- Giulivi C, Hochstein P, Davies KJ. Hydrogen peroxide production by red blood cells. *Free Radic Biol Med* 1994;16:123–29
- Hebbel RP, Leung A, Mohandas N. Oxidation-induced changes in microrheologic properties of the red blood cell membrane. *Blood* 1990;76:1015–20
- Browne P, Shalev O, Hebbel RP. The molecular pathobiology of cell membrane iron: the sickle red cell as a model. *Free Radic Biol Med* 1998;24:1040–08
- Freikman I, Ringel I, Fibach E. Oxidative stress-induced membrane shedding from RBCs is Ca flux-mediated and affects membrane lipid composition. J Membr Biol 2011;240:73–82
- Hebbel RP, Key NS. Microparticles in sickle cell anaemia: promise and pitfalls. Br J Haematol 2016;174:16–29
- 46. Camus SM, De Moraes JA, Bonnin P, Abbyad P, Le Jeune S, Lionnet F, Loufrani L, Grimaud L, Lambry JC, Charue D, Kiger L, Renard JM, Larroque C, Le Clesiau H, Tedgui A, Bruneval P, Barja-Fidalgo C, Alexandrou A, Tharaux PL, Boulanger CM, Blanc-Brude OP. Circulating cell membrane microparticles transfer heme to endothelial cells and trigger vasoocclusions in sickle cell disease. *Blood* 2015;**125**:3805–14
- Piel FB, Steinberg MH, Rees DC. Sickle cell disease. N Engl J Med 2017;376:1561–73
- Reiter CD, Wang X, Tanus-Santos JE, Hogg N, Cannon RO, 3rd, Schechter AN, Gladwin MT. Cell-free hemoglobin limits nitric oxide bioavailability in sickle-cell disease. *Nat Med* 2002;8:1383–89
- Mendonca R, Silveira AA, Conran N. Red cell DAMPs and inflammation. *Inflamm Res* 2016;65:665–78
- Wagener FA, Eggert A, Boerman OC, Oyen WJ, Verhofstad A, Abraham NG, Adema G, van Kooyk Y, de Witte T, Figdor CG. Heme is a potent inducer of inflammation in mice and is counteracted by heme oxygenase. *Blood* 2001;98:1802–11
- True AL, Olive M, Boehm M, San H, Westrick RJ, Raghavachari N, Xu X, Lynn EG, Sack MN, Munson PJ, Gladwin MT, Nabel EG. Heme oxygenase-1 deficiency accelerates formation of arterial thrombosis

through oxidative damage to the endothelium, which is rescued by inhaled carbon monoxide. *Circ Res* 2007;**101**:893–901

- Conran N, De PE. Thromboinflammatory mechanisms in sickle cell disease - challenging the hemostatic balance. *Haematologica* 2020;105:2380–90
- Wun T, Paglieroni T, Rangaswami A, Franklin PH, Welborn J, Cheung A, Tablin F. Platelet activation in patients with sickle cell disease. Br J Haematol 1998;100:741–49
- Ataga KI, Key NS. Hypercoagulability in sickle cell disease: new approaches to an old problem. *Hematology Am Soc Hematol Educ Program* 2007;1:91–96
- Shet AS, Lizarralde-Iragorri MA, Naik RP. The molecular basis for the prothrombotic state in sickle cell disease. *Haematologica* 2020;105:2368–79
- Villagra J, Shiva S, Hunter LA, Machado RF, Gladwin MT, Kato GJ. Platelet activation in patients with sickle disease, hemolysisassociated pulmonary hypertension, and nitric oxide scavenging by cell-free hemoglobin. *Blood* 2007;110:2166–72
- Brzoska T, Vats R, Bennewitz MF, Tutuncuoglu E, Watkins SC, Ragni MV, Neal MD, Gladwin MT, Sundd P. Intravascular hemolysis triggers ADP-mediated generation of platelet-rich thrombi in precapillary pulmonary arterioles. *JCI Insight* 2020;5
- Bourne JH, Colicchia M, Di Y, Martin E, Slater A, Roumenina LT, Dimitrov JD, Watson SP, Rayes J. Heme induces human and mouse platelet activation through C-type-lectin-like receptor-2. *Haematologica* 2021;106:626–29
- Setty BN, Rao AK, Stuart MJ. Thrombophilia in sickle cell disease: the red cell connection. *Blood* 2001;98:3228–33
- Zecher D, Cumpelik A, Schifferli JA. Erythrocyte-derived microvesicles amplify systemic inflammation by thrombin-dependent activation of complement. *Arterioscler Thromb Vasc Biol* 2014;34:313–20
- Setty BN, Betal SG, Zhang J, Stuart MJ. Heme induces endothelial tissue factor expression: potential role in hemostatic activation in patients with hemolytic anemia. J Thromb Haemost 2008;6:2202–09
- 62. Proenca-Ferreira R, Brugnerotto AF, Garrido VT, Dominical VM, Vital DM, Ribeiro MF, dos Santos ME, Traina F, Olalla-Saad ST, Costa FF, Conran N. Endothelial activation by platelets from sickle cell anemia patients. *PLoS One* 2014;9:e89012
- 63. Garrido VT, Proenca-Ferreira R, Dominical VM, Traina F, Bezerra MA, de Mello MR, Colella MP, Araujo AS, Saad ST, Costa FF, Conran N. Elevated plasma levels and platelet-associated expression of the prothrombotic and pro-inflammatory protein, TNFSF14 (LIGHT), in sickle cell disease. Br J Haematol 2012;**158**:788–97
- Greenberg J, Ohene-Frempong K, Halus J, Way C, Schwartz E. Trial of low doses of aspirin as prophylaxis in sickle cell disease. J Pediatr 1983;102:781–84
- Semple MJ, Al-Hasani SF, Kioy P, Savidge GF. A double-blind trial of ticlopidine in sickle cell disease. *Thromb Haemost* 1984;51:303–06
- Cabannes R, Lonsdorfer J, Castaigne JP, Ondo A, Plassard A, Zohoun I. Clinical and biological double-blind-study of ticlopidine in preventive treatment of sickle-cell disease crises. *Agents Actions Suppl* 1984;15:199–212
- 67. Wun T, Soulieres D, Frelinger AL, Krishnamurti L, Novelli EM, Kutlar A, Ataga KI, Knupp CL, McMahon LE, Strouse JJ, Zhou C, Heath LE, Nwachuku CE, Jakubowski JA, Riesmeyer JS, Winters KA. Doubleblind, randomized, multicenter phase 2 study of prasugrel versus placebo in adult patients with sickle cell disease. J Hematol Oncol 2013;6:17
- 68. Styles L, Heiselman D, Heath LE, Moser BA, Small DS, Jakubowski JA, Zhou C, Redding-Lallinger R, Heeney MM, Quinn CT, Rana SR, Kanter J, Winters KJ. Prasugrel in children with sickle cell disease: pharmacokinetic and pharmacodynamic data from an open-label, adaptive-design, dose-ranging study. J Pediatr Hematol Oncol 2015;37:1–9
- 69. Heeney MM, Hoppe CC, Abboud MR, Inusa B, Kanter J, Ogutu B, Brown PB, Heath LE, Jakubowski JA, Zhou C, Zamoryakhin D, Agbenyega T, Colombatti R, Hassab HM, Nduba VN, Oyieko JN, Robitaille N, Segbefia CI, Rees DC, Investigators D. A multinational trial of prasugrel for sickle cell vaso-occlusive events. N Engl J Med 2016;**374**:625-35

- Conran N, Rees DC. Prasugrel hydrochloride for the treatment of sickle cell disease. *Expert Opin Investig Drugs* 2017;26:865–72
- 71. Bertheloot D, Latz E. HMGB1, IL-1alpha, IL-33 and S100 proteins: dual-function alarmins. *Cell Mol Immunol* 2017;**14**:43–64
- Ye Y, Zeng Z, Zhang JT, Xiong H, Gu X. L. The role of high mobility group box 1 in ischemic stroke. *Front Cell Neurosci* 2019;13:127
- Aufradet E, DeSouza G, Bourgeaux V, Bessaad A, Campion Y, Canet-Soulas E, Pialoux V, Chirico EN, Chevrier AM, Godfrin Y, Martin C. Hypoxia/reoxygenation stress increases markers of vaso-occlusive crisis in sickle SAD mice. *Clin Hemorheol Microcirc* 2013;54:297–312
- Gutsaeva DR, Parkerson JB, Yerigenahally SD, Kurz JC, Schaub RG, Ikuta T, Head CA. Inhibition of cell adhesion by anti-P-selectin aptamer: a new potential therapeutic agent for sickle cell disease. *Blood* 2011;117:727–35
- Kaul DK, Hebbel RP. Hypoxia/reoxygenation causes inflammatory response in transgenic sickle mice but not in normal mice. J Clin Invest 2000;106:411–20
- Hebbel RP. Ischemia-reperfusion injury in sickle cell anemia: relationship to acute chest syndrome, endothelial dysfunction, arterial vasculopathy, and inflammatory pain. *Hematol Oncol Clin North Am* 2014;28:181–98
- Osarogiagbon UR, Choong S, Belcher JD, Vercellotti GM, Paller MS, Hebbel RP. Reperfusion injury pathophysiology in sickle transgenic mice. *Blood* 2000;96:314–20
- Silva DG, Belini Junior E, de Almeida EA, Bonini-Domingos CR. Oxidative stress in sickle cell disease: an overview of erythrocyte redox metabolism and current antioxidant therapeutic strategies. *Free Radic Biol Med* 2013;65:1101–09
- Hebbel RP, Belcher JD, Vercellotti GM. The multifaceted role of ischemia/reperfusion in sickle cell anemia. J Clin Invest 2020;130:1062–72
- Embury SH, Mohandas N, Paszty C, Cooper P, Cheung ATW. *In vivo* blood flow abnormalities in the transgenic knockout sickle cell mouse. *J Clin Invest* 1999;103:915–20
- Nath KA, Hebbel RP. Sickle cell disease: renal manifestations and mechanisms. Nat Rev Nephrol 2015;11:161–71
- Nath KA, Katusic ZS. Vasculature and kidney complications in sickle cell disease. J Am Soc Nephrol 2012;23:781–84
- Park SY, Matte A, Jung Y, Ryu J, Anand WB, Han EY, Liu M, Carbone C, Melisi D, Nagasawa T, Locascio JJ, Lin CP, Silberstein LE, De Franceschi L. Pathologic angiogenesis in the bone marrow of humanized sickle cell mice is reversed by blood transfusion. *Blood* 2020;135:2071–84
- 84. Nombela-Arrieta C, Pivarnik G, Winkel B, Canty KJ, Harley B, Mahoney JE, Park SY, Lu J, Protopopov A, Silberstein LE. Quantitative imaging of haematopoietic stem and progenitor cell localization and hypoxic status in the bone marrow microenvironment. *Nat Cell Biol* 2013;15:533–43
- Bennewitz MF, Tutuncuoglu E, Gudapati S, Brzoska T, Watkins SC, Monga SP, Pradhan-Sundd T, Sundd P. P-selectin-deficient mice to study pathophysiology of sickle cell disease. *Blood Adv* 2020;4:266–73
- Anea CB, Lyon M, Lee IA, Gonzales JN, Adeyemi A, Falls G, Kutlar A, Brittain JE. Pulmonary platelet thrombi and vascular pathology in acute chest syndrome in patients with sickle cell disease. *Am J Hematol* 2016;91:173–78
- Embury SH. The not-so-simple process of sickle cell vasoocclusion. Microcirculation 2004;11:101–13
- Steinberg MH, Chui DH, Dover GJ, Sebastiani P, Alsultan A. Fetal hemoglobin in sickle cell anemia: a glass half full? *Blood* 2014;123:481–85
- Sewchand LS, Johnson CS, Meiselman HJ. The effect of fetal hemoglobin on the sickling dynamics of SS erythrocytes. *Blood Cells* 1983;9:147–59
- Clark MR, Greenquist AC, Shohet SB. Stabilization of the shape of sickled cells by calcium and A23187. *Blood* 1976;48:899–909
- Kaul DK, Fabry ME, Windisch P, Baez S, Nagel RL. Erythrocytes in sickle cell anemia are heterogeneous in their rheological and hemodynamic characteristics. J Clin Invest 1983;72:22–29

 Embury SH, Clark MR, Monroy GM, Mohandas N. Concurrent sickle cell anemia and α thalassemia: effect on pathological properties of sickle red cells. J Clin Invest 1984;73:116–23

- Clark MR, Mohandas N, Shohet SB. Deformability of oxygenated irreversibly sickled cells. J Clin Invest 1980;65:189–96
- Eaton WA, Hofrichter J. Hemoglobin S gelation sickle cell disease. Blood 1987;70:1245–66
- Brittenham GM, Schechter AN, Noguchi CT. Hemoglobin S polymerization: primary determination of the hemolytic and clinical severity of the sickling syndromes. *Blood* 1985;65:283–89
- Keidan AJ, Sowter MC, Johnson CS, Noguchi CT, Girling AJ, Stevens SM, Stuart J. Effect of polymerization tendency on haematological, rheological and clinical parameters in sickle cell anaemia. *Br J Haematol* 1989;71:551–57
- 97. Di Liberto G, Kiger L, Marden MC, Boyer L, Poitrine FC, Conti M, Rakotoson MG, Habibi A, Khorgami S, Vingert B, Maitre B, Galacteros F, Pirenne F, Bartolucci P. Dense red blood cell and oxygen desaturation in sickle-cell disease. *Am J Hematol* 2016;**91**:1008–13
- Platt OS, Thorington BD, Brambilla DJ, Milner PF, Rosse WF, Vichinsky E, Kinney TR. Pain in sickle cell disease. Rates and risk factors. N Engl J Med 1991;325:11–16
- Billett HH, Kim K, Fabry ME, Nagel RL. The percentage of dense red cells does not predict incidence of sickle cell painful crisis. *Blood* 1986;68:301–03
- Billett HH, Nagel RL, Fabry ME. Paradoxical increase of painful crises in sickle cell patients with alpha-thalassemia. *Blood* 1995;86:4382
- 101. Ataga KI, Reid M, Ballas SK, Yasin Z, Bigelow C, James LS, Smith WR, Galacteros F, Kutlar A, Hull JH, Stocker JW, Investigators ICAS. Improvements in haemolysis and indicators of erythrocyte survival do not correlate with acute vaso-occlusive crises in patients with sickle cell disease: a phase III randomized, placebo-controlled, double-blind study of the Gardos channel blocker senicapoc (ICA-17043). Br J Haematol 2011;153:92–104
- 102. Nagel RL. Sickle cell anemia is a multigene disease: sickle painful crises, a case in point. *Am J Hematol* 1993;**42**:96–101
- Mohandas N, Evans E. Adherence of sickle erythrocytes to vascular endothelial cells: requirement for both cell membrane changes plasma factors. *Blood* 1984;64:282–87
- Barabino GA, McIntire LV, Eskin SG, Sears DA, Udden M. Rheological studies of erythrocyte-endothelial cell interactions in sickle cell disease. *Progr Clin Biol Res* 1987;240:113–27
- Barabino GA, McIntire LV, Eskin SG, Sears DA, Udden M. Endothelial cell interactions with sickle cell, sickle trait, mechanically injured, and normal erythrocytes under controlled flow. *Blood* 1987;70:152–57
- Ballas SK, Larner J, Smith ED, Surrey S, Schwartz E, Rappaport EF. Rheologic predictors of the severity of the painful sickle cell crisis. *Blood* 1988;72:1216–23
- 107. Ballas SK. Sickle cell anemia with few painful crises is characterized by decreased red cell deformability and increased number of dense cells. Am J Hematol 1991;36:122–30
- Kaul DK, Fabry ME, Nagel RL. Microvascular sites and characteristics of sickle cell adhesion to vascular endothelium in shear flow conditions: pathophysiological implications. *Proc Natl Acad Sci USA* 1989;86:3356–60
- Ferrone FA. Polymerization and sickle cell disease: a molecular view. Microcirculation 2004;11:115–28
- Powars D, Chan LS, Schroeder WA. The variable expression of sickle cell disease is genetically determined. *Sem Hematol* 1990;27:360–76
- 111. Thomas AN, Pattison C, Serjeant GR. Causes of death in sickle-cell disease in Jamaica. Br Med J 1982;285:633-5
- 112. Francis RB, Jr., Johnson CS. Vascular occlusion in sickle cell disease: current concepts and unanswered questions. *Blood* 1991;77:1405–14
- Ballas SK, Mohandas N. Sickle red cell microrheology and sickle blood rheology. *Microcirculation* 2004;11:209–25
- Lipowsky HH, Sheikh NU, Katz DM. Intravital microscopy of capillary microdynamics in sickle cell disease. J Clin Invest 1987;80:117–27
- 115. Rodgers GP, Schechter AN, Noguchi CT, Klein HG, Nienhuis AW, Bonner RF. Microcirculatory adaptations in sickle cell anemia: reactive hyperemia response. Am J Physiol 1990;258:H113–20

 Cheung AT, Chen PC, Larkin EC, Duong PL, Ramanujam S, Tablin F, Wun T. Microvascular abnormalities in sickle cell disease: a computerassisted intravital microscopy study. *Blood* 2002;99:3999–4005

- 117. van Tuijn CF, van Beers EJ, Schnog JJ, Biemond BJ. Pain rate and social circumstances rather than cumulative organ damage determine the quality of life in adults with sickle cell disease. *Am J Hematol* 2010;85:532–35
- 118. Hebbel RP, Yamada O, Moldow CF, Jacob HS, White JG, Eaton JW. Abnormal adherence of sickle erythrocytes to cultured vascular endothelium. Possible mechanism for microvascular occlusion in sickle cell disease. J Clin Invest 1980;65:154–60
- Hebbel RP, Boogaerts MAB, Eaton JW, Steinberg MH. Erythrocyte adherence to endothelium in sickle-cell anemia. N Engl J Med 1980;302:992–95
- 120. Kutlar A, Ataga KI, McMahon L, Howard J, Galacteros F, Hagar W, Vichinsky E, Cheung AT, Matsui N, Embury SH. A potent oral P-selectin blocking agent improves microcirculatory blood flow and a marker of endothelial cell injury in patients with sickle cell disease. *Am J Hematol* 2012;87:536–39
- 121. Johnson C, Telen M. Adhesion molecules and hydroxyurea in the pathophysiology of sickle cell disease. *Haematologica* 2008;93:481–86
- 122. Harlan JM. Introduction: anti-adhesion therapy in sickle cell disease. *Blood* 2000;**95**:365–67
- Hebbel RP. Clinical implications of basic research: blockade of adhesion of sickle cells to endothelium by monoclonal antibodies. N Engl J Med 2000;342:1910–12
- 124. Lizarralde-Iragorri MA, Lefevre SD, Cochet S, El Hoss S, Brousse V, Filipe A, Dussiot M, Azouzi S, Le Van Kim C, Rodrigues-Lima F, Francais O, Le Pioufle B, Klei T, van Bruggen R, El Nemer W. Oxidative stress activates red cell adhesion to laminin in sickle cell disease. *Haematologica* 2020 (in press).
- Lawrence MB, Springer TA. Leukocytes roll on a selectin at physiologic flow rates: distinction from and prerequisite for adhesion through integrins. *Cell* 1991;65:859–73
- 126. Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. *Cell* 1994;76:301–14
- 127. Robinson SD, Frenette PS, Rayburn H, Cummiskey M, Ullman-Cullere M, Wagner DD, Hynes RO. Multiple, targeted deficiencies in selectins reveal a predominant role for P-selectin in leukocyte recruitment. *Proc Natl Acad Sci USA* 1999;96:11452–57
- Mayadas TN, Johnson RC, Rayburn H, Hynes RO, Wagner DD. Leukocyte rolling and extravasation are severely compromised in P selectin-deficient mice. *Cell* 1993;74:541–54
- Ley K, Laudanna C, Cybulsky MI, Nourshargh S. Getting to the site of inflammation: the leukocyte adhesion Cascade updated. *Nat Rev Immunol* 2007;7:678–89
- McEver RP, Zhu C. Rolling cell adhesion. Annu Rev Cell Dev Biol 2010;26:363–96
- McEver RP. Regulation of expression of E-selectin and P-selectin. In: Vestweber D (ed.), *The selectins: initiators of leukocyte endothelial adhesion*. Amsterdam: Harwood, 1997, pp. 31–47.
- Barkalow FJ, Goodman MJ, Gerritsen ME, Mayadas TN. Brain endothelium lack one of two pathways of P-selectin-mediated neutrophil adhesion. *Blood* 1996;88:4585–93
- Wood K, Russell J, Hebbel RP, Granger DN. Differential expression of E- and P-selectin in the microvasculature of sickle cell transgenic mice. *Microcirculation* 2004;11:377–85
- 134. Khew-Goodall Y, Butcher CM, Litwin MS, Newlands S, Korpelainen EI, Noack LM, Berndt MC, Lopez AF, Gamble JR, Vadas MA. Chronic expression of P-selectin on endothelial cells stimulated by the T-cell cytokine, interleukin-3. *Blood* 1996;87:1432–38
- 135. Yao L, Pan J, Setiadi H, Patel KD, McEver RP. Interleukin 4 or oncostatin M induces a prolonged increase in P-selectin mRNA and protein in human endothelial cells. J Exp Med 1996;184:81–92
- 136. Subramaniam M, Koedam JA, Wagner DD. Divergent fates of P- and E-selectins after their expression on the plasma membrane. *Mol Biol Cell* 1993;4:791–801

- Matsui NM, Borsig L, Rosen SD, Yaghmai M, Varki A, Embury SH. Pselectin mediates the adhesion of sickle erythrocytes to the endothelium. *Blood* 2001;98:1955–62
- Matsui NM, Varki A, Embury SH. Heparin inhibits the flow adhesion of sickle red blood cells to P-selectin. *Blood* 2002;100:3790–96
- 139. Sultana C, Shen Y, Rattan V, Johnson C, Kalra VK. Interaction of sickle erythrocytes with endothelial cells in the presence of endothelial cell conditioned medium induces oxidant stress leading to transendothelial migration of monocytes. *Blood* 1998;92:3924–35
- 140. Patel KD, Zimmerman GA, Prescott SM, McEver RP, McIntyre TM. Oxygen radicals induce human endothelial cells to express GMP-140 and bind neutrophils. J Cell Biol 1991;112:749–59
- 141. Takano M, Meneshian A, Sheikh E, Yamakawa Y, Wilkins KB, Hopkins EA, Bulkley GB. Rapid upregulation of endothelial P-selectin expression via reactive oxygen species generation. Am J Physiol Heart Circ Physiol 2002;283:H2054–61
- 142. Embury SH, Baran CE, Hefner CA, Seto CK, Matsui NM. The nature of P-selectin ligands on sickle cells. *Blood* 2004;**104**:107a
- 143. Marui N, Offermann MK, Swerlick R, Kunsch C, Rosen CA, Ahmad M, Alexander RW, Medford RM. Vascular cell adhesion molecule-1 (VCAM-1) gene transcription and expression are regulated through an antioxidant-sensitive mechanism in human vascular endothelial cells. J Clin Invest 1993;92:1866–74
- 144. Shiu YT, Udden MM, McIntire LV. Perfusion with sickle erythrocytes up-regulates ICAM-1 and VCAM-1 gene expression in cultured human endothelial cells. *Blood* 2000;**95**:3232–41
- 145. Duits AJ, Pieters RC, Saleh AW, van Rosmalen E, Katerberg H, Berend K, Rojer RA. Enhanced levels of soluble VCAM-1 in sickle cell patients and their specific increment during vasoocclusive crisis. *Clin Immunol Immunopathol* 1996;81:96–98
- 146. Schnog JB, Rojer RA, Mac Gillavry MR, ten Cate H, Brandjes DP, Duits AJ. Steady-state sVCAM-1 serum levels in adults with sickle cell disease. Ann Hematol 2003;82:109–13
- 147. Blann AD. Endothelial cell activation, injury, damage and dysfunction: separate entities or mutual terms? *Blood Coagul Fibrinolysis* 2000;**11**:623–30
- 148. Turhan A, Weiss LA, Mohandas N, Coller BS, Frenette PS. Primary role for adherent leukocytes in sickle cell vascular occlusion: a new paradigm. *Proc Natl Acad Sci USA* 2002;99:3047–51
- 149. Solovey A, Somani A, Belcher JD, Milbauer L, Vincent L, Pawlinski R, Nath KA, Kelm RJ, Jr, Mackman N, O'Sullivan MG, Gupta K, Vercellotti GM, Hebbel RP. A monocyte-TNF-endothelial activation axis in sickle transgenic mice: therapeutic benefit from TNF blockade. *Am J Hematol* 2017;92:1119–30
- 150. French JA, Kenny D, Scott JP, Hoffmann RG, Wood JD, Hudetz AG, Hillery CA. Mechanisms of stroke in sickle cell disease: sickle erythrocytes decrease cerebral blood flow in rats after nitric oxide synthase inhibition. *Blood* 1997;89:4591–99
- 151. Lutty GA, Taomoto M, Cao J, McLeod DS, Vanderslice P, McIntyre BW, Fabry ME, Nagel RL. Inhibition of TNF-alpha-induced sickle RBC retention in retina by a VLA-4 antagonist. *Invest Ophthalmol Vis Sci* 2001;42:1349–55
- 152. Lutty GA, Otsuji T, Taomoto M, Merges C, McLeod DS, Kim SY, Vanderslice P, Suzuka S, Fabry ME, Nagel RL. Mechanisms for sickle red blood cell retention in choroid. *Curr Eye Res* 2002;25:163–71
- 153. Kaul DK, Fabry ME, Costantini F, Rubin EM, Nagel RL. In vivo demonstration of red cell-endothelial interaction, sickling and altered microvascular response to oxygen in the sickle transgenic mouse. J Clin Invest 1995;96:2845–53
- West MS, Wethers D, Smith J, Steinberg M. Laboratory profile of sickle cell disease: a cross-sectional analysis. J Clin Epidemiol 1992;45:893–909
- 155. Castro O, Brambilla DJ, Thorington B, Reindorf CA, Scott RB, Gillette P, Vera JC, Levy PS. The acute chest syndrome in sickle cell disease: incidence and risk factors. The cooperative study of sickle cell disease. *Blood* 1994;84:643–49
- 156. Platt OS, Brambilla DJ, Rosse WF, Milner PF, Castro O, Steinberg MH, Klug PP. Mortality in sickle cell disease. Life expectancy and risk factors for early death. N Engl J Med 1994;330:1639–44

- 157. Anyaegbu CC, Okpala IE, Akren'Ova YA, Salimonu LS. Peripheral blood neutrophil count and candidacidal activity correlate with the clinical severity of sickle cell anaemia (SCA). *Eur J Haematol* 1998;60:267–68
- 158. Fadlon E, Vordermeier S, Pearson TC, Mire-Sluis AR, Dumonde DC, Phillips J, Fishlock K, Brown KA. Blood polymorphonuclear leukocytes from the majority of sickle cell patients in the crisis phase of the disease show enhanced adhesion to vascular endothelium and increased expression of CD64. *Blood* 1998;91:266–74
- Kasschau MR, Barabino GA, Bridges KR, Golan DE. Adhesion of sickle neutrophils and erythrocytes to fibronectin. *Blood* 1996;87:771–80
- Assis A, Conran N, Canalli AA, Lorand-Metze I, Saad ST, Costa FF. Effect of cytokines and chemokines on sickle neutrophil adhesion to fibronectin. *Acta Haematol* 2005;113:130–36
- 161. Kuhnle GE, Kuebler WM, Groh J, Goetz AE. Effect of blood flow on the leukocyte-endothelium interaction in pulmonary microvessels. *Am J Respir Crit Care Med* 1995;152:1221–28
- Frenette PS. Sickle cell vaso-occlusion: multistep and multicellular paradigm. Curr Opin Hematol 2002;9:101–06
- Finnegan EM, Turhan A, Golan DE, Barabino GA. Adherent leukocytes capture sickle erythrocytes in an in vitro flow model of vasoocclusion. *Am J Hematol* 2007;82:266–75
- 164. Haynes J, Jr, Obiako B, King JA, Hester RB, Ofori-Acquah S. Activated neutrophil-mediated sickle red blood cell adhesion to lung vascular endothelium: role of phosphatidylserine-exposed sickle red blood cells. Am J Physiol Heart Circ Physiol 2006;291:H1679–85
- Zennadi R, Chien A, Xu K, Batchvarova M, Telen MJ. Sickle red cells induce adhesion of lymphocytes and monocytes to endothelium. *Blood* 2008;112:3474–83
- 166. Zennadi R, Moeller BJ, Whalen EJ, Batchvarova M, Xu K, Shan S, Delahunty M, Dewhirst MW, Telen MJ. Epinephrine-induced activation of LW-mediated sickle cell adhesion and vaso-occlusion in vivo. *Blood* 2007;110:2708–17
- 167. Canalli AA, Proenca RF, Franco-Penteado CF, Traina F, Sakamoto TM, Saad ST, Conran N, Costa FF. Participation of Mac-1, LFA-1 and VLA-4 integrins in the in vitro adhesion of sickle cell disease neutrophils to endothelial layers, and reversal of adhesion by simvastatin. *Haematologica* 2011;96:526–33
- Chang J, Patton JT, Sarkar A, Ernst B, Magnani JL, Frenette PS. GMI-1070, a novel pan-selectin antagonist, reverses acute vascular occlusions in sickle cell mice. *Blood* 2010;116:1779–86
- 169. Koehl B, Nivoit P, El Nemer W, Lenoir O, Hermand P, Pereira C, Brousse V, Guyonnet L, Ghinatti G, Benkerrou M, Colin Y, Le Van Kim C, Tharaux PL. The endothelin B receptor plays a crucial role in the adhesion of neutrophils to the endothelium in sickle cell disease. *Haematologica* 2017;**102**:1161–72
- 170. Canalli AA, Franco-Penteado CF, Saad ST, Conran N, Costa FF. Increased adhesive properties of neutrophils in sickle cell disease may be reversed by pharmacological nitric oxide donation. *Haematologica* 2008;93:605–09
- 171. Dominical VM, Samsel L, Nichols JS, Costa FF, McCoy JP, Jr, Conran N, Kato GJ. Prominent role of platelets in the formation of circulating neutrophil-red cell heterocellular aggregates in sickle cell anemia. *Haematologica* 2014;99:e214–17
- 172. Pircher J, Engelmann B, Massberg S, Schulz C. Platelet-neutrophil crosstalk in atherothrombosis. *Thromb Haemost* 2019;**119**:1274–82
- 173. Polanowska-Grabowska R, Wallace K, Field JJ, Chen L, Marshall MA, Figler R, Gear AR, Linden J. P-selectin-mediated platelet-neutrophil aggregate formation activates neutrophils in mouse and human sickle cell disease. *Arterioscler Thromb Vasc Biol* 2010;**30**:2392–99
- 174. Zhang D, Chen G, Manwani D, Mortha A, Xu C, Faith JJ, Burk RD, Kunisaki Y, Jang JE, Scheiermann C, Merad M, Frenette PS. Neutrophil ageing is regulated by the microbiome. *Nature* 2015;**525**:528–32
- 175. Garcia F, Mendonca R, Miguel LI, Dominical VM, Saad STO, Costa FF, Conran N. CXCR4(hi) effector neutrophils in sickle cell anemia: potential role for elevated circulating serotonin (5-HT) in CXCR4(hi) neutrophil polarization. *Sci Rep* 2020;**10**:14262

176. Chen G, Chang J, Zhang D, Pinho S, Jang JE, Frenette PS. Targeting Mac-1-mediated leukocyte-RBC interactions uncouples the benefits for acute vaso-occlusion and chronic organ damage. *Exp Hematol* 2016;44:940–46

- 177. Kim K, Li J, Barazia A, Tseng A, Youn SW, Abbadessa G, Yu Y, Schwartz B, Andrews RK, Gordeuk VR, Cho J. ARQ 092, an orallyavailable, selective AKT inhibitor, attenuates neutrophil-platelet interactions in sickle cell disease. *Haematologica* 2017;102:246–59
- 178. Wun T, Cordoba M, Rangaswami A, Cheung AW, Paglieroni T. Activated monocytes and platelet-monocyte aggregates in patients with sickle cell disease. *Clin Lab Haematol* 2002;24:81–88
- Brittain JE, Knoll CM, Ataga KI, Orringer EP, Parise LV. Fibronectin bridges monocytes and reticulocytes via integrin alpha4beta1. Br J Haematol 2008;141:872–81
- 180. Chaar V, Picot J, Renaud O, Bartolucci P, Nzouakou R, Bachir D, Galacteros F, Colin Y, Le Van Kim C, El Nemer W. Aggregation of mononuclear and red blood cells through an {alpha}4{beta}1-Lu/ basal cell adhesion molecule interaction in sickle cell disease. *Haematologica* 2010;95:1841-48
- 181. Liu Y, Zhong H, Bao W, Mendelson A, An X, Shi P, Chou ST, Manwani D, Yazdanbakhsh K. Patrolling monocytes scavenge endothelialadherent sickle RBCs: a novel mechanism of inhibition of vasoocclusion in SCD. *Blood* 2019;**134**:579–90
- 182. Chen G, Zhang D, Fuchs TA, Manwani D, Wagner DD, Frenette PS. Heme-induced neutrophil extracellular traps contribute to the pathogenesis of sickle cell disease. *Blood* 2014;**123**:3818–27
- 183. Schimmel M, Nur E, Biemond BJ, van Mierlo GJ, Solati S, Brandjes DP, Otten HM, Schnog JJ, Zeerleder S. and Curama Study G. Nucleosomes and neutrophil activation in sickle cell disease painful crisis. *Haematologica* 2013;98:1797–803
- 184. Dutta D, Methe B, Amar S, Morris A, Lim SH. Intestinal injury and gut permeability in sickle cell disease. *J Transl Med* 2019;**17**:183
- Cavazzana M, Antoniani C, Miccio A. Gene therapy for beta-hemoglobinopathies. *Mol Ther* 2017;25:1142–54
- 186. Rai P, Malik P. Gene therapy for hemoglobin disorders a mini-review. J Rare Dis Res Treat 2016;1:25–31
- 187. Vichinsky E, Hoppe CC, Ataga KI, Ware RE, Nduba V, El-Beshlawy A, Hassab H, Achebe MM, Alkindi S, Brown RC, Diuguid DL, Telfer P, Tsitsikas DA, Elghandour A, Gordeuk VR, Kanter J, Abboud MR, Lehrer-Graiwer J, Tonda M, Intondi A, Tong B, Howard J, Investigators HT. A phase 3 randomized trial of voxelotor in sickle cell disease. N Engl J Med 2019;381:509–19
- 188. Steinberg MH, Lu ZH, Barton FB, Terrin ML, Charache S, Dover GJ. Fetal hemoglobin in sickle cell anemia: determinants of response to hydroxyurea. Multicenter study of hydroxyurea. *Blood* 1997;89:1078–88
- 189. Charache S, Barton FB, Moore RD, Terrin ML, Steinberg MH, Dover GJ, Ballas SK, McMahon RP, Castro O, Orringer EP. Hydroxyurea and sickle cell anemia. Clinical utility of a myelosuppressive "switching" agent. The multicenter study of hydroxyurea in sickle cell anemia. *Medicine* 1996;75:300–26
- 190. Ware RE, Davis BR, Schultz WH, Brown RC, Aygun B, Sarnaik S, Odame I, Fuh B, George A, Owen W, Luchtman-Jones L, Rogers ZR, Hilliard L, Gauger C, Piccone C, Lee MT, Kwiatkowski JL, Jackson S, Miller ST, Roberts C, Heeney MM, Kalfa TA, Nelson S, Imran H, Nottage K, Alvarez O, Rhodes M, Thompson AA, Rothman JA, Helton KJ, Roberts D, Coleman J, Bonner MJ, Kutlar A, Patel N, Wood J, Piller L, Wei P, Luden J, Mortier NA, Stuber SE, Luban NL, Cohen AR, Pressel S, Adams RJ. Hydroxycarbamide versus chronic transfusion for maintenance of transcranial doppler flow velocities in children with sickle cell anaemia-TCD with transfusions changing to hydroxyurea (TWiTCH): a multicentre, open-label, phase 3, noninferiority trial. *Lancet* 2016;**387**:661–70
- 191. Bernaudin F, Verlhac S, Arnaud C, Kamdem A, Hau I, Leveille E, Vasile M, Kasbi F, Madhi F, Fourmaux C, Biscardi S, Gluckman E, Socie G, Dalle JH, Epaud R, Pondarre C. Long-term treatment follow-up of children with sickle cell disease monitored with abnormal transcranial Doppler velocities. *Blood* 2016;**127**:1814–22

- 192. Steinberg MH, McCarthy WF, Castro O, Ballas SK, Armstrong FD, Smith W, Ataga K, Swerdlow P, Kutlar A, DeCastro L, Waclawiw MA. The risks and benefits of long-term use of hydroxyurea in sickle cell anemia: a 17.5 year follow-up. Am J Hematol 2010;85:403–08
- 193. Platt OS, Orkin SH, Dover G, Beardsley GP, Miller B, Nathan DG. Hydroxyurea enhances fetal hemoglobin production in sickle cell anemia. J Clin Invest 1984;74:652–56
- 194. Noguchi CT, Rodgers GP, Schechter AN. Intracellular polymerization. Disease severity and therapeutic predictions. Ann NY Acad Sci 1989;565:75–82
- 195. Almeida CB, Scheiermann C, Jang JE, Prophete C, Costa FF, Conran N, Frenette PS. Hydroxyurea and a cGMP-amplifying agent have immediate benefits on acute vaso-occlusive events in sickle cell disease mice. *Blood* 2012;**120**:2879–88
- 196. Chaar V, Laurance S, Lapoumeroulie C, Cochet S, De Grandis M, Colin Y, Elion J, Le Van Kim C, El Nemer W. Hydroxycarbamide decreases sickle reticulocyte adhesion to resting endothelium by inhibiting endothelial lutheran/basal cell adhesion molecule (Lu/BCAM) through phosphodiesterase 4A activation. J Biol Chem 2014;289:11512-21
- 197. Bartolucci P, Chaar V, Picot J, Bachir D, Habibi A, Fauroux C, Galacteros F, Colin Y, Le Van Kim C, El Nemer W. Decreased sickle red blood cell adhesion to laminin by hydroxyurea is associated with inhibition of Lu/BCAM protein phosphorylation. *Blood* 2010;**116**:2152–59
- Cartron JP, Elion J. Erythroid adhesion molecules in sickle cell disease: effect of hydroxyurea. *Transfus Clin Biol* 2008;15:39–50
- 199. Proenca-Ferreira R, Franco-Penteado CF, Traina F, Saad ST, Costa FF, Conran N. Increased adhesive properties of platelets in sickle cell disease: roles for alphaIIb beta3-mediated ligand binding, diminished cAMP signalling and increased phosphodiesterase 3A activity. Br J Haematol 2010;149:280–88
- 200. Canalli AA, Franco-Penteado CF, Traina F, Saad ST, Costa FF, Conran N. Role for cAMP-protein kinase a signalling in augmented neutro-phil adhesion and chemotaxis in sickle cell disease. *Eur J Haematol* 2007;**79**:330–37
- 201. Gambero S, Canalli AA, Traina F, Albuquerque DM, Saad ST, Costa FF, Conran N. Therapy with hydroxyurea is associated with reduced adhesion molecule gene and protein expression in sickle red cells with a concomitant reduction in adhesive properties. *Eur J Haematol* 2007;**78**:144–51
- Lanzkron S, Haywood C, Jr, Fagan PJ, Rand CS. Examining the effectiveness of hydroxyurea in people with sickle cell disease. J Health Care Poor Underserved 2010;21:277–86
- 203. Ataga KI, Kutlar A, Kanter J, Liles D, Cancado R, Friedrisch J, Guthrie TH, Knight-Madden J, Alvarez OA, Gordeuk VR, Gualandro S, Colella MP, Smith WR, Rollins SA, Stocker JW, Rother RP. Crizanlizumab for the prevention of pain crises in sickle cell disease. N Engl J Med 2017;376:429–39
- 204. Charache S, Terrin ML, Moore RD, Dover GJ, Barton FB, Eckert SV, McMahon RP, Bonds DR, Anemia TlotMSoHiSCEffect of hydroxyurea on the frequency of painful crises in sickle cell anemia. N Engl J Med 1995;332:1317–22
- 205. Wun T, Styles L, DeCastro L, Telen MJ, Kuypers F, Cheung A, Kramer W, Flanner H, Rhee S, Magnani JL, Thackray H. Phase 1 study of the E-selectin inhibitor GMI 1070 in patients with sickle cell anemia. *PLoS One* 2014;9:e101301
- 206. Wun T, Telen MJ, Krishnamurti L, McCavit TL, DeCastro LM, Flanner H, Kuypers F, Larkin SK, Rhee S, Magnani JL, Thackray H. Pan-selectin antagonist rivipansel (GMI-1070) reduces soluble E-selectin levels while improving clinical outcomes in SCD vaso-occlusive crisis *blood*. 2014;**124**:2704
- 207. Telen MJ, Wun T, McCavit TL, De Castro LM, Krishnamurti L, Lanzkron S, Hsu LL, Smith WR, Rhee S, Magnani JL, Thackray H. Randomized phase 2 study of GMI-1070 in SCD: reduction in time to resolution of vaso-occlusive events and decreased opioid use. *Blood* 2015;**125**:2656-64
- Dampier CD, Telen MJ, Wun T, Smith WR, Brown C, Desai P, El Rassi FA, Kanter J, Fuh BR, Pastore YD, Rothman JA, Taylor JG, Readett D,

Lozier JN, Magnani JL, Thackray HM, Hassell KL. Early initiation of treatment with rivipansel for acute vaso-occlusive crisis in sickle cell disease (SCD) achieves earlier discontinuation of IV opioids and shorter hospital stay: reset clinical trial analysis. *Blood* 2020;**136**: 18–9

- 209. Fernandez F, N'Guyen P, Van Ryn J, Ofosu FA, Hirsh J, Buchanan MR. Hemorrhagic doses of heparin and other glycosaminoglycans induce a platelet defect. *Thromb Res* 1986;43:491–95
- 210. Lancelot M, White J, Sarnaik S, Hines P. Low molecular weight heparin inhibits sickle erythrocyte adhesion to VCAM-1 through VLA-4 blockade in a standardized microfluidic flow adhesion assay. Br J Haematol 2017;178:479–81
- 211. Koenig A, Norgard-Sumnicht K, Linhardt R, Varki A. Differential interactions of heparin and heparan sulfate glycosaminoglycans with the selectins. Implications for the use of unfractionated and low molecular weight heparins as therapeutic agents. *J Clin Invest* 1998;**101**:877–89
- 212. Telen MJ, Batchvarova M, Shan S, Bovee-Geurts PH, Zennadi R, Leitgeb A, Brock R, Lindgren M. Sevuparin binds to multiple adhesive ligands and reduces sickle red blood cell-induced vaso-occlusion. Br J Haematol 2016;175:935–48
- 213. Cheung ATW, Chan MS, Ramanujam S, Rangaswami A, Curl K, Franklin P, Wun T. Effects of poloxamer 188 treatment on sickle cell vaso-occlusive crisis: computer-assisted intravital microscopy study. J Investig Med 2004;52:402–06
- Okpala I. Investigational selectin-targeted therapy of sickle cell disease. Expert Opin Investig Drugs 2015;24:229–38
- 215. Manwani D, Chen G, Carullo V, Serban S, Olowokure O, Jang J, Huggins M, Cohen HW, Billett H, Atweh GF, Frenette PS, Shi PA. Single-dose intravenous gammaglobulin can stabilize neutrophil Mac-1 activation in sickle cell pain crisis. *Am J Hematol* 2015;90:381–85
- 216. Belcher JD, Chen C, Nguyen J, Abdulla F, Zhang P, Nguyen H, Nguyen P, Killeen T, Miescher SM, Brinkman N, Nath KA, Steer CJ, Vercellotti GM. Haptoglobin and hemopexin inhibit vaso-occlusion and inflammation in murine sickle cell disease: role of heme oxygenase-1 induction. *PLoS One* 2018;13:e0196455
- 217. Ferreira WA, Jr, Chweih H, Lanaro C, Almeida CB, Brito PL, Gotardo EMF, Torres L, Miguel LI, Franco-Penteado CF, Leonardo FC, Garcia F, Saad STO, Frenette PS, Brockschnieder D, Costa FF, Stasch JP, Sandner P, Conran N. Beneficial effects of soluble guanylyl cyclase stimulation and activation in sickle cell disease are amplified by hydroxyurea: in vitro and in vivo studies. J Pharmacol Exp Ther 2020;**374**:469–78
- 218. Conran N, Torres L. cGMP modulation therapeutics for sickle cell disease. *Exp Biol Med* 2019;**244**:132–46
- 219. Zimmer DP, Shea CM, Tobin JV, Tchernychev B, Germano P, Sykes K, Banijamali AR, Jacobson S, Bernier SG, Sarno R, Carvalho A, Chien YT, Graul R, Buys ES, Jones JE, Wakefield JD, Price GM, Chickering JG, Milne GT, Currie MG, Masferrer JL. Olinciguat, an oral sGC stimulator, exhibits diverse pharmacology across preclinical models of cardiovascular, metabolic, renal, and inflammatory disease. *Front Pharmacol* 2020;**11**:419
- 220. Almeida CB, Traina F, Lanaro C, Canalli AA, Saad ST, Costa FF, Conran N. High expression of the cGMP-specific phosphodiesterase, PDE9A, in sickle cell disease (SCD) and the effects of its inhibition in erythroid cells and SCD neutrophils. *Br J Haematol* 2008;**142**:836–44
- 221. Charnigo RJ, Beidler D, Rybin D, Pittman DD, Tan B, Howard J, Michelson AD, Frelinger Al, Iii Clarke N. PF-04447943, a phosphodiesterase 9A inhibitor, in stable sickle cell disease patients: a phase Ib randomized, placebo-controlled study. *Clin Transl Sci* 2019;**12**:180–88
- 222. McArthur JG, Svenstrup N, Chen C, Fricot A, Carvalho C, Nguyen J, Nguyen P, Parachikova A, Abdulla F, Vercellotti GM, Hermine O, Edwards D, Ribeil JA, Belcher JD, Maciel TT. A novel, highly potent and selective phosphodiesterase-9 inhibitor for the treatment of sickle cell disease. *Haematologica* 2020;**105**:623–31
- 223. Strader MB, Jana S, Meng F, Heaven MR, Shet AS, Thein SL, Alayash AI. Post-translational modification as a response to cellular stress induced by hemoglobin oxidation in sickle cell disease. *Sci Rep* 2020;**10**:14218
- 224. Niihara Y, Zerez CR, Akiyama DS, Tanaka KR. Oral L-glutamine therapy for sickle cell anemia: I. Subjective clinical improvement and

favorable change in red cell NAD redox potential. Am J Hematol 1998;58:117-21

- 225. Niihara Y, Koh HA, Tran L, Razon RL, Macan H, Stark C, Wun T, Adams-Graves PA. Phase 3 study of L–glutamine therapy for sickle cell anemia and sickle ß0-thalassemia. *Blood* 2014;**124**:86
- 226. Niihara Y, Miller ST, Kanter J, Lanzkron S, Smith WR, Hsu LL, Gordeuk VR, Viswanathan K, Sarnaik S, Osunkwo I, Guillaume E, Sadanandan S, Sieger L, Lasky JL, Panosyan EH, Blake OA, New TN, Bellevue R, Tran LT, Razon RL, Stark CW, Neumayr LD. Vichinsky EP and investigators of the phase 3 trial of l-glutamine in sickle cell D. A phase 3 trial of l-glutamine in sickle cell disease. N Engl J Med 2018;379:226-35
- 227. Ogu UO, Thomas M, Chan F, Vattappally L, Sebastian G, Crouch A, You S, Minniti CP. L-glutamine use in adults with sickle cell disease: clinical trials where success meets reality. *Am J Hematol* 2020;
- Thamilarasan M, Estupinan R, Batinic-Haberle I, Zennadi R. Mn porphyrins as a novel treatment targeting sickle cell NOXs to reverse and prevent acute vaso-occlusion in vivo. *Blood Adv* 2020;4:2372–86
- 229. Rab MAE, van Oirschot BA, Kosinski PA, Hixon J, Johnson K, Chubukov V, Dang L, Pasterkamp G, van Straaten S, van Solinge WW, van Beers EJ, Kung C, van Wijk R. AG-348 (Mitapivat), an allosteric activator of red blood cell pyruvate kinase, increases enzymatic activity, protein stability, and ATP levels over a broad range of PKLR genotypes. *Haematologica* 2020.
- 230. Brown RC, Cruz K, Kalfa TA, Kuypers FA, Saraf SL, Estepp JH, Smart LR, Malik P, Lerman ML, Mayer R, Ribadeneira MD, Forsyth S, Schroeder P, Wu E, Kelly P, Telen MJ. FT-4202, an allosteric activator of pyruvate Kinase-R, demonstrates proof of mechanism and proof of concept after a single dose and after multiple daily doses in a phase 1 study of patients with sickle cell disease. *Blood* 2020;**136**: 19–20

231. Wood KW, Geib J, Wu E, Berlin J, Webster I, Ataga KI, Howard J, Estepp JH, Telen MJ, Brevard J. An adaptive, randomized, placebocontrolled, double-blind, multi-center study of oral FT-4202, a pyruvate kinase activator in patients with sickle cell disease (PRAISE). *Blood* 2020;**136**:19–20

- 232. Liang X, Xiu C, Liu M, Lin C, Chen H, Bao R, Yang S, Yu J. Plateletneutrophil interaction aggravates vascular inflammation and promotes the progression of atherosclerosis by activating the TLR4/ NF-kappaB pathway. J Cell Biochem 2019;120:5612–19
- 233. Geng X, Mihaila R, Yuan Y, Strutt S, Benz J, Tang T, Mayer C, Oksenberg D. Inclacumab, a fully human anti-p-selectin antibody, directly binds to PSGL-1 binding region and demonstrates robust and durable inhibition of cell adhesion. *Blood* 2020;**136**:10–11
- 234. van Beers EJ, van Tuijn CF, Mac Gillavry MR, van der Giessen A, Schnog JJ, Biemond BJ. Group Cs Sickle cell disease-related organ damage occurs irrespective of pain rate: implications for clinical practice. *Haematologica* 2008;93:757–60
- 235. Hebbel RP. Adhesion of sickle red cells to endothelium: myths and future directions. *Transfus Clin Biol* 2008;**15**:14–18
- Joneckis CC, Ackley RL, Orringer EP, Wayner EA, Parise LV. Integrin alpha 4 beta 1 and glycoprotein IV (CD36) are expressed on circulating reticulocytes in sickle cell anemia. *Blood* 1993;82:3548–55
- 237. Lanaro C, Franco-Penteado CF, Albuqueque DM, Saad ST, Conran N, Costa FF. Altered levels of cytokines and inflammatory mediators in plasma and leukocytes of sickle cell anemia patients and effects of hydroxyurea therapy. J Leukoc Biol 2009;85:235–42
- Vercellotti GM, Dalmasso AP, Schaid TR, Jr, Nguyen J, Chen C, Ericson ME, Abdulla F, Killeen T, Lindorfer MA, Taylor RP, Belcher JD. Critical role of C5a in sickle cell disease. *Am J Hematol* 2019;94:327–37
- 239. Kutlar A, Embury SH. Cellular Adhesion and the Endothelium: P-Selectin. *Hematol Oncol Clin North Am* 2014;28:323-39