

Platelet activating factor: the good and the bad in the ischemic/reperfused heart

Claudia Penna^{1,2}, Eleonora Bassino³ and Giuseppe Alloati^{2,3}

¹Dipartimento di Scienze Cliniche e Biologiche, ASO San Luigi, 10043 Orbassano (TO); ²Istituto Nazionale per la Ricerca Cardiovascolare (INRC), 40126 Bologna; ³Dipartimento di Biologia Animale e dell'Uomo, University of Torino, 10123 Torino, Italy

Corresponding author: Giuseppe Alloati, Dipartimento di Biologia Animale e dell'Uomo, Via Accademia Albertina 13, 10123 Torino, Italy. Email: giuseppe.alloati@unito.it

Abstract

The present review is focused on the dual role played by platelet-activating factor (PAF) in ischemia and reperfusion (I/R) injury of the heart. Although the involvement of PAF in the pathogenesis of myocardial reperfusion injury is well established, in the last few years it has emerged that very low concentrations of PAF exert cardioprotective effects, comparable to that afforded by ischemic preconditioning (IP). PAF is a potent phosphoglyceride involved in different pathophysiological conditions affecting the cardiovascular system, including the development of myocardial I/R injury. PAF is released from the I/R myocardium in concentrations (1–10 nmol/L) high enough to negatively modulate coronary circulation as well as electrical and contractile activities. PAF may act either directly, via generation of secondary mediators, or through the activation of inflammatory cells like platelets and polymorphonuclear neutrophils, which exacerbate postischemic myocardial injury. The effects of PAF are mediated through specific receptors (PAFRs) that belong to the superfamily of G protein-coupled receptors. Since cardiomyocytes not only produce PAF but also possess PAFRs, it is likely that PAF acts as an autocrine/paracrine mediator. Although the negative effects exerted by high concentrations of PAF are well established, several recent findings from our and other laboratories have demonstrated that very low concentrations (pmol/L) of PAF infused before ischemia induce cardioprotective effects similar to those afforded by IP, and that endogenous PAF production participates in the induction of IP itself. The IP-like action exerted by low concentrations of PAF is due to the activation/phosphorylation of kinases included in the reperfusion injury salvage kinase (RISK) pathway, such as protein kinase C, Akt/PkB and nitric oxide synthase. Together with the activation of mitochondrial K_{ATP} channels, these events may allow prevention of mitochondrial permeability transition pores opening at reperfusion. Moreover, the nitric oxide-dependent S-nitrosylation of L-type Ca²⁺ channels induced by PAF reduces intracellular Ca²⁺ overload.

Keywords: platelet-activating factor, heart, ischemia/reperfusion, ischemic preconditioning

Experimental Biology and Medicine 2011; **236**: 390–401. DOI: 10.1258/ebm.2011.010316

Introduction

Platelet-activating factor (1-O-alkyl-2-acetyl-sn-glycero-3-phosphocholine, PAF) is an autacoid with profound effects on many systems, including the muscles, vascular tissue and brain. The term PAF was coined in 1972 by Benveniste *et al.*¹ to indicate the lipid mediator released from rabbit basophils after IgE stimulation, initially described as 'a soluble factor' involved in leukocyte-dependent histamine and serotonin release from platelets (PLT).² PAF is a phospholipid with diverse and potent physiological effects that belongs to a family of biologically active, structurally related alkyl phosphoglycerides.^{3–7} While PAF is minimally expressed under normal physiological conditions, several cell types

(such as neutrophils and monocytes) release significant amounts of PAF in particular conditions, such as oxidative stress or ischemia and reperfusion (I/R). PAF mediates cell-to-cell communication, acting both as an intercellular or an intracellular messenger. Some of its actions are achieved at concentrations as low as 1–10 pmol/L and play a relevant role in the development of several pathological and physiological processes. Numerous cell types, in particular those involved in inflammatory reactions, have been shown to produce PAF upon stimulation.^{3–7} In addition, human endothelial cells were found to produce PAF after stimulation by several inflammatory mediators, including leukotrienes C₄ and D₄, histamine, bradykinin, hydrogen peroxide,

interleukin (IL)-8 and IL-1 α , or tumor necrosis factor (TNF)- α .³⁻⁷ Cardiomyocytes have also been reported to synthesize PAF under appropriate stimulation. *In vitro*, PAF promotes the aggregation, chemotaxis, granule secretion and oxygen radical generation from leukocytes and the adherence of leukocytes to the endothelium. PAF increases the permeability of the endothelial cell monolayer, stimulates the contraction of smooth muscle⁸ and exerts negative inotropic and arrhythmogenic effects on cardiac muscle.⁹⁻¹⁴ PAF decreases the viability of a variety of cell types, including human lymphocytes, corneal epithelial cells, HaCaT cells and human colon carcinoma cells.³⁻⁷

PAF is synthesized by a variety of tissues and cells through two different pathways. The *remodeling pathway* present in the inflammatory cells includes the activation of phospholipase A₂ (PLA₂) and acetyl CoA: 1-alkyl-sn-glycero-3-phosphorylcholine 2-O acetyltransferase, and the production of 2-lysophospholipids as metabolic intermediates (1-alkyl-2-lyso-glycero-3-phosphocholine, lyso-PAF).¹⁵⁻¹⁷ The other PAF biosynthetic pathway, which is mainly operative in the kidney and in the central nervous system, has been termed the *de novo pathway*.^{18,19} This pathway involves the synthesis of 1-O-alkyl-2-acetylgllycerol, which is then converted to PAF by a specific choline phosphotransferase. A PAF acetylhydrolase (PAF-AH) present in plasma²⁰ and in various tissues degrades PAF by cleaving the short acyl chain at sn-2 position and originating the biologically inactive lyso-PAF.²¹

PAF acts via specific receptors (PAFRs) present on membranes of different cell types, in particular smooth muscle cells,²² cardiomyocytes²³ and endothelial cells.²⁴ In these latter cells, PAFRs are expressed not only on the cell surface but also in the large endosomal compartment.²⁵ It has been suggested that cell surface receptor activation induces immediate effects, whereas long-term responses are mediated by intracellular receptors. In microvascular endothelial cells, intracellular PAFR stimulation modulates several biological functions, such as transcriptional regulation of major genes, namely cyclooxygenase-2 and inducible nitric oxide synthase (NOS).²⁶ Membrane PAFRs belong to the G protein-coupled receptor (GPCR) superfamily.^{27,28} As they are coupled to both G_q and G_i types of G proteins, the stimulation of PAFRs leads to activation of PLC and PLA₂. This activation causes the transient production of diacylglycerol, which activates protein kinase C (PKC), and of inositol trisphosphate, which mediates the release of internal calcium stores. Moreover, PAF has been found to stimulate the release of arachidonic acid in various cell types by different mechanisms.²⁹⁻³² The activation of PLA₂ by PAF occurs through a PKC-dependent mechanism and may be regulated by the intracellular levels of cAMP.³⁰⁻³² Arachidonic acid metabolites have been shown to mediate several biological activities of PAF in the cardiovascular system.²⁹⁻³³ In addition, it has been shown that PAF stimulates tyrosine phosphorylation of several proteins in PLT,³⁴ neutrophils³⁵ and macrophages.³⁶ Because PAFRs contain several tyrosine residues in its intracellular loops and tail, it has been suggested that tyrosine phosphorylation may be involved in receptor downregulation.³⁶ It has been shown that PAF may activate a mitogen-activated protein

kinase (MAPK)³⁷ and may induce the early tyrosine phosphorylation of focal adhesion kinase in human endothelial cells.³⁸ Moreover, in human neutrophils, PAF activates MAPK kinase-3, a known activator of p38 MAPK.³⁹ Further details regarding signal transduction pathways involved in cardiac effects of PAF will be presented in the following sections.

Role of PAF in I/R injury

Reperfusion injury is defined as the myocardial cellular dysfunction induced by the re-establishment of coronary perfusion, in contrast to myocardial damage brought about during the preceding ischemic episode. Reperfusion injury involves mechanical, extracellular and intracellular processes, which include the following key mechanisms: generation of reactive oxygen species (ROS), reduced availability of NO, Ca²⁺ overload (Figure 1) and the opening of mitochondrial permeability transition pores (mPTPs; Figure 2).⁴⁰⁻⁴³

Release of PAF from I/R heart

Several experiments investigating the production of PAF and the protective effect of PAF receptor antagonists in the I/R heart support the hypothesis that PAF plays a relevant role in these pathophysiological conditions.^{4,40} Intravascular release of PAF has been detected in the blood of patients with coronary artery disease undergoing atrial pacing.⁴³ Increased plasma concentrations of PAF during myocardial infarction have been related in part to increased production of PAF by neutrophils,⁴⁴ or to depression of plasma PAF-AH activity, which in turn may allow a prolonged half-life of newly synthesized PAF. In contrast, other studies failed to detect increased PAF concentrations in the peripheral blood after myocardial infarction in humans.^{45,46} Besides humans, elevated blood PAF concentrations were also detected in baboons following myocardial infarction⁴⁷ and in sheep after coronary occlusion and reperfusion.⁴⁸ The presence of increased levels of the PAF metabolite, lyso-PAF, in canine myocardium subjected to permanent ligation of a coronary branch gave indirect evidence of elevated PAF biosynthesis during myocardial ischemia.⁴⁹ *In vivo* findings were supported by *in vitro* experiments on isolated perfused hearts, demonstrating the release of PAF during reperfusion. In rabbits, PAF was detected in the coronary effluent during the initial reperfusion of ischemic heart.^{50,51} Although the precise cellular source of PAF was not identified in this model, likely candidates are endothelial cells and cardiomyocytes. Indeed, it has been reported that cultured endothelial cells,⁵² as well as neonatal rat myocytes, synthesize PAF after prolonged hypoxia.⁵³ In the isolated rabbit heart, in which the effects of PAF are platelet-dependent, the amounts of released PAF (in the nmol/L range) were enough to activate platelets to cause a significant worsening of left ventricular function during reperfusion.⁵⁰ A further worsening occurred when both polymorphonuclear neutrophils (PMNs) and PLT were present, as a consequence of PAF-dependent PMN-PLT cooperation.⁵⁴ The effects of

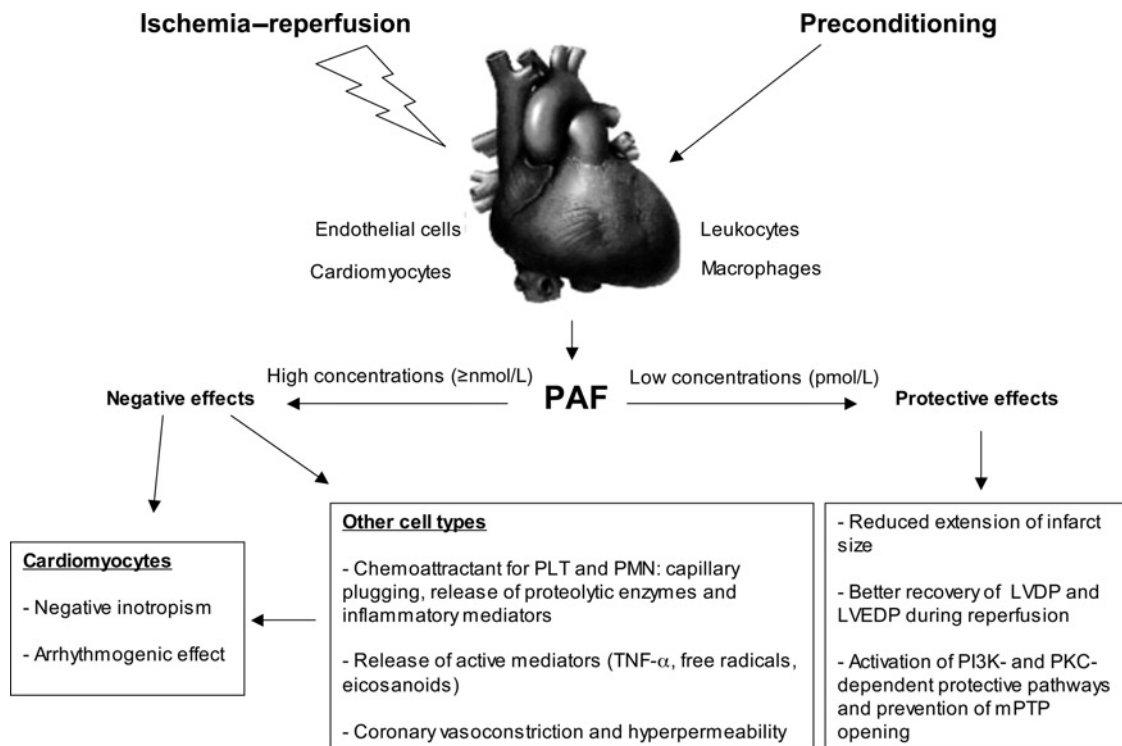


Figure 1 Dual role played by platelet-activating factor (PAF) in ischemia and reperfusion (I/R) injury of the heart. Released at high concentrations (\geq nmol/L) after prolonged ischemia followed by reperfusion, PAF participates to the pathogenesis of myocardial reperfusion injury. On the other hand, very low concentrations (pmol/L) of PAF may exert a cardioprotective effect, comparable to that afforded by the classical ischemic preconditioning. The negative inotropic and arrhythmogenic effects exerted by PAF are due both to a direct action on cardiomyocytes and to indirect effects consequent to reduced coronary flow and release of other mediators. LVDP, left ventricular developed pressure; LVEDP, left ventricular end-diastolic pressure; mPTP, mitochondrial permeability transition pores; PLT, platelets; PMN, polymorphonuclear neutrophils

PAFR antagonists were studied in different *in vitro* and *in vivo* experimental models. PAFR antagonists prevented both the PLT-dependent and -independent mechanical and electrical alterations that occurred, respectively, in rabbit,^{50,55,56} guinea pig⁵⁷ and rat isolated heart⁵⁸ after I/R. In the experimental myocardial infarction due to coronary occlusion, PAFR antagonists reduced the hemodynamic alterations as well as the size of the necrotic area and the accumulation of PLT and leukocytes observed in rabbit^{59,60} and sheep⁶¹ hearts. In rats, PAFR antagonists reduced infarct size and the occurrence of arrhythmias.^{62–64} Although some conflicting results were obtained with different PAFR antagonists, a comparable protective effect was also observed in the dog.^{65,66} To date, no clinical trials have investigated the protective effect of PAFR antagonists in cardiac alterations associated with I/R in humans.

In summary, the relevant role of PAF in cardiac dysfunctions caused by I/R is supported by several *in vitro* and *in vivo* findings, showing the release of PAF during reperfusion and the protective effect exerted by PAF receptor antagonists. The concentrations of PAF released (1–10 nmol/L) are high enough to activate PLT and PMN, causing a significant worsening of cardiac function during reperfusion.

Effects of PAF in coronary circulation

PAF infusion into the coronary circulation induced variations in the coronary tone, depending on the doses and

the animal species used. Low doses of PAF (0.03–0.3 nmol/L) induced both an increment and decrement of coronary blood flow (CBF) in the pig, in the absence of significant changes in systemic blood pressure. Higher doses (1–10 nmol/L) of PAF caused a reduction in CBF accompanied by a negative inotropic effect and electrocardiogram signs of ischemia (S-T segment depression and arrhythmias). The early increase in CBF is independent from the generation of cyclo- and lipoxygenase-derived metabolites, while the subsequent vasoconstriction is primarily due to the production of thromboxane A₂.⁶⁷ PAF also induced coronary vasoconstriction and S-T segment depression in rabbits.⁶⁸ Conflicting effects were observed in dogs, in which PAF has been reported to reduce CBF and systemic arterial pressure,⁶⁹ to produce a PLT-dependent coronary vasodilation⁷⁰ or a biphasic vasodilator/vasoconstrictor effect.⁷¹ The effects of PAF strongly depend on the integrity of endothelial cells; indeed, PAF induces a vasodilating effect when the endothelium is intact, whereas vasoconstriction is prominent in the presence of injured endothelium, as it may occur after ischemia.⁷²

The coronary vasoconstrictor action of high doses of PAF observed *in vivo* was confirmed by *ex vivo* experiments performed on isolated perfused hearts. PAF induced a dose-dependent increase in coronary vascular resistances in the isolated guinea pig heart.^{9,11,73–75} The vasoconstrictor effect of PAF was completely blocked by PAFR antagonists, thus suggesting the involvement of specific PAF receptors.⁷⁵

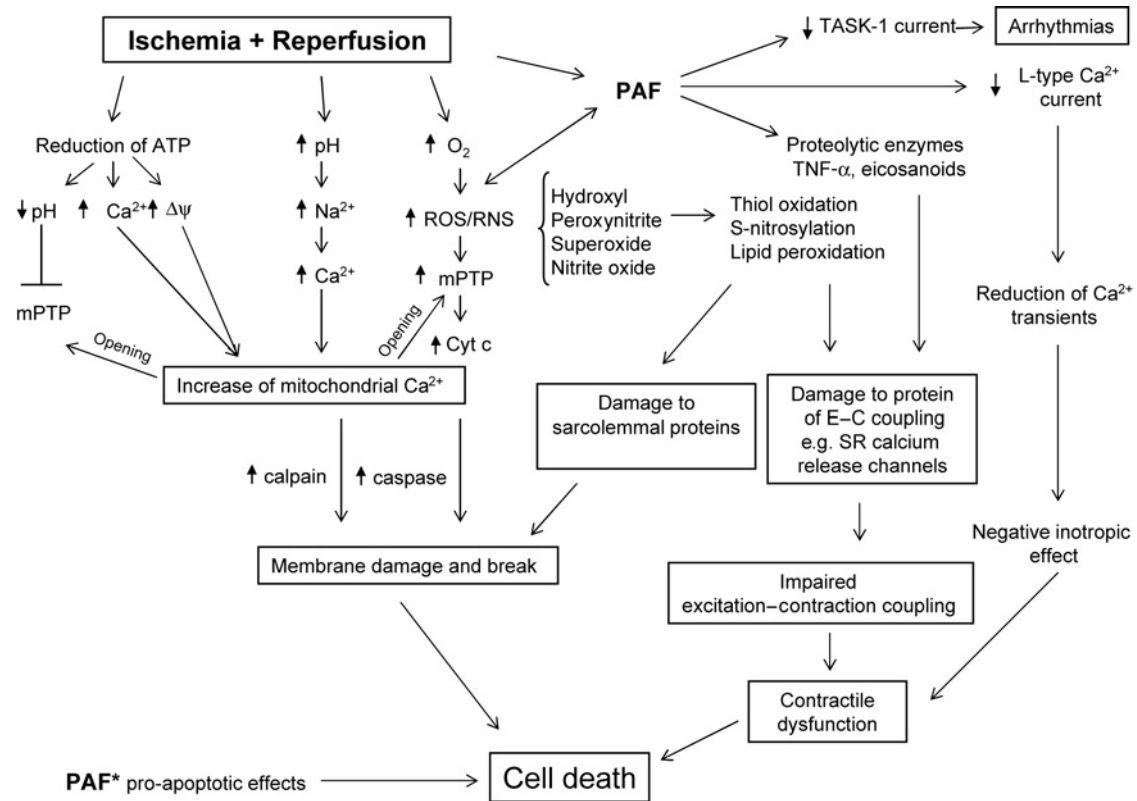


Figure 2 Cross-link between the intracellular pathways responsible for the negative effects of platelet-activating factor (PAF) and those involved in cell damage and contractile dysfunction in ischemia and reperfusion (I/R) heart. PAF* indicates the direct proapoptotic effect exerted by PAF. Cyt c, cytochrome c; Δψ, mitochondrial membrane potential; E-C, excitation-contraction; mPTP, mitochondrial permeability transition pore; RNS, reactive nitrogen species, ROS, reactive oxygen species; SR, sarcoplasmic reticulum; TASK-1, TWIK-related acid-sensitive K⁺ channels

Comparable coronary vasoconstriction was obtained with high doses of PAF in isolated perfused rat heart.^{76–78} However, low doses induced vasodilation alone or vasodilation followed by vasoconstriction, which involves production of both prostaglandins and leukotrienes.^{76–78} In the isolated rabbit heart, which is insensitive to this mediator when perfused with physiological saline alone,^{50,79} PAF markedly reduced coronary flow when blood cells were added to the perfusion fluid.⁷⁹ Thus, the isolated rabbit heart was used as a model to study the *in vitro* interaction between PLT and PMN in the coronary circulation. In the rabbit heart perfused with PLT alone, the infusion of PAF induced a dose-dependent decrease of coronary flow, while PMN stimulated by PAF had no effect. However, as a result of cooperation of PMN with PLT, a marked reduction of coronary flow occurred when both these cells were stimulated with PAF.⁵⁴ A comparable increase in coronary resistances was observed in rabbit hearts perfused with PMN activated by *N*-formyl-L-methionyl-L-leucyl-L-phenylalanine.⁸⁰ The fact that the neutrophil-dependent coronary vasoconstriction was inhibited by a PAFR antagonist suggested a relevant role for PAF as a secondary mediator.⁸¹ The vasoactive effect of PAF and its PMN-dependent mechanism have been directly studied in isolated coronary vessels, in which PAF induced a dose-dependent decrease in the diameter of the vessels and increased the permeability coefficient of albumin.⁸² Disruption of the endothelium abolished the vasomotor

response to PAF, and perfusion with PMN significantly enhanced PAF-induced changes in vasomotor tone and permeability. Furthermore, administration of PAF caused PMN adhesion to the endothelium of coronary arterioles. Although isolated human coronary artery rings did not react to PAF challenge in basal conditions, after hypoxia they undergo a PAF-dependent biphasic contraction, characterized by an initial short phase of contraction, followed by a longer tonic shortening.⁸³

Taken together, these findings indicate that PAF exerts dose-dependent alterations of coronary tone. At concentrations comparable to those detected in the coronary effluent after I/R (1–10 nmol/L), PAF caused a reduction in coronary flow accompanied by a negative inotropic effect. The vasoconstrictor effect of PAF is due to specific PAFR activation and, depending on the animal species, it may be due to a direct action on smooth muscle cells or, at least in part, to generation of cyclo- and lipoxygenase-derived metabolites, or activation of PLT and PMN.

Electrophysiological and inotropic effects of PAF on cardiac cells

The alterations in cardiac function observed *in vivo* after infusion of PAF can result either from a direct action on the heart or from indirect effects, such as systemic changes and variations in pre- and afterload pressures. Moreover, alterations in cardiac performance may depend on the effect of PAF on

the coronary circulation, on the conduction system and on the contractile properties of myocardium.⁸⁴ Experiments performed on the isolated heart perfused at constant flow or isolated atrial and ventricular preparations stimulated at constant frequency indicated that PAF exerts a direct effect on cardiac electrical and contractile activities. The effects of PAF on the isolated heart perfused at constant pressure are quite different, depending on the animal species studied. In guinea pig isolated heart, infusion of PAF induced conduction arrhythmias and reduced both the action potential duration and the force of contraction.^{85,86} The negative inotropic effect of PAF was further confirmed by *in vitro* experiments on isolated guinea pig atrium and papillary muscle,^{10,87,88} showing a direct negative inotropic effect of PAF, paralleled by a significant decrease of action potential duration.^{10,89} In contrast, the negative effects induced by PAF in the isolated rat heart were mainly dependent on the reduction of coronary flow consequent to production and release of endogenous leukotrienes.²⁹ PAF induced a biphasic dose-dependent effect on human cardiac muscles, characterized by a transient positive effect followed by a marked prolonged negative inotropic effect and reduction of action potential duration.^{12,13} Experiments performed on isolated cardiac cells confirmed the ability of PAF to directly modulate the electrical and contractile activities of cardiac muscle. It has been shown that the decrease in twitch tension induced by PAF in isolated adult cardiomyocytes is accompanied by a reduction of intracellular systolic calcium concentration⁹⁰ and of L-type calcium current.⁹¹ On the other hand, the repolarization abnormalities and conduction arrhythmias induced by PAF in cardiac cells may be explained with the observed alterations of inwardly rectifying background potassium channels (IK1)⁹² and inhibition of TWIK-related acid-sensitive K⁺ channels (TASK-1).⁹³ In addition, PAF stimulates cardiac muscarinic potassium channels, via a PLA₂-lipoygenase dependent mechanism.³² These results are consistent with the fact that the presence of PAFRs on cardiomyocytes, previously suggested by the protective effect of PAFR antagonists, was directly demonstrated by cloning and characterization of PAF receptor gene in human cardiomyocytes.²³

The intracellular pathways involved in the myocardial response to PAF were investigated in a series of experiments performed in our laboratory. The role of PI3K γ -mediated signal transduction in the regulation of myocardial performance in response to PAF was analyzed using p110 γ knockout mice.⁹⁴ We demonstrated that the negative inotropic effect of PAF in the heart depends on PI3K γ in response to a Gi-mediated signal transduction pathway. The blocking effect induced by a NOS inhibitor suggests that PKB/Akt-mediated phosphorylation and thus activation of eNOS are the critical events. On the same experimental model, we further studied the relevance of PAF/PI3K γ signaling for the outcome of I/R in isolated hearts, demonstrating that loss of PI3K γ resulted in a significantly better postischemic recovery of contractile force during reperfusion than wild-type controls. Postischemic function in wild-type hearts could be improved by both a PAFR antagonist and a NOS inhibitor, indicating the PAF/PI3K γ /NOS pathway as an important cause of myocardial dysfunction.⁹⁴

In addition to exerting direct effects on cardiac cells, it has been shown that PAF stimulates the release of other biologically active mediators such as eicosanoids (leukotrienes and thromboxane), superoxide anions and TNF- α . These mediators can furthermore enhance myocardial dysfunction.⁴⁰ In particular, TNF- α , which like PAF acts as cardio-depressant agent released by I/R heart, may cooperate with PAF. Indeed, TNF- α induces release of PAF from several cell types, including cardiac and skeletal muscle cells, and PAF receptor antagonists have been shown to reduce the negative effects of TNF- α .^{95,96} Apoptosis has been implicated in myocardial infarction-related cell death. Besides the above-mentioned detrimental actions of PAF, it has been recently shown that high concentrations of PAF (0.2–20 μ mol/L) directly induce apoptosis in H9c2 cardiomyocytes via a calcium-dependent p38 MAPK-activated cytochrome c/caspase-3 apoptosis signaling pathway (Figure 2).⁹⁷

In summary, it has been shown that, besides altering cardiac performance by reducing CBF, PAF may also exert direct effects on the electrical and contractile activities. The shortening of the action potential duration and the negative inotropic effect induced by PAF have been related to reduction of both L-type calcium current and intracellular systolic calcium concentration. The repolarization abnormalities and conduction arrhythmias induced by PAF may be explained with the observed alterations of IK1 current and inhibition of TASK-1 channels. By using p110 γ knockout mice, it has also been shown that the PI3K γ /NOS pathway plays an important role in myocardial dysfunction induced by PAF.

What causes PAF synthesis and release during I/R?

Selected experiments were performed to investigate the mechanisms responsible for PAF biosynthesis in I/R heart. The occurrence of oxidative stress related to the generation of ROS at the beginning of reperfusion plays an important role in reperfusion-induced cell death.^{40,98} Since the generation of oxygen radicals and their release into the coronary effluent occurs in parallel with the release of PAF during the early phases of reperfusion,⁵⁰ we hypothesized that oxidative stress may trigger the synthesis of PAF. To test this hypothesis, we treated guinea-pig isolated perfused heart with dihydroxyfumaric acid (DHF), a free radical-generating compound. DHF stimulated PAF production and caused mechanical and electrical alterations that were prevented by superoxide dismutase or by the PAFR antagonist WEB 2170.⁹⁹ These results support the hypothesis that the burst of ROS released during early reperfusion may lead to PAF biosynthesis and that, on the other hand, PAF may, at least in part, act as secondary mediator of oxygen radicals in the heart. In line with our observations on multicellular cardiac preparations, it has been already shown that H₂O₂ stimulates the synthesis of PAF by primary cultures of bovine pulmonary artery and by human umbilical vein endothelial cells (HUVECs). In parallel with PAF synthesis, H₂O₂ also induced the endothelial-cell-dependent adhesion of neutrophils to HUVEC monolayers.¹⁰⁰ Besides the above-mentioned studies, in

which PAF production was induced by acute *in vitro* treatment with chemicals generating a marked oxidative stress, other observations suggested that the interaction between ROS and PAF may occur also *in vivo* in certain pathophysiological situations. Inside the cells, the toxic actions of ROS are limited by antioxidant enzyme systems that protect them against peroxidation and control concentrations of intracellular peroxides. One of these, the enzyme phospholipid hydroperoxide glutathione peroxidase, acts as a negative modulator of PAF biosynthesis through inhibition of p38 phosphorylation. Since selenium is required for optimal activity of glutathione peroxidases, it is reasonable that selenium deficiency would result in the accumulation of lipid peroxides and subsequent induction of PAF synthesis. In fact, it has been shown that selenium deficiency causes increased production of PAF in human endothelial cells with enhanced activity of PLA₂ and acetyltransferase.⁴ Moreover, it has been shown that PAF mediates the adhesion of monocytes to endothelium induced by LDL and oxidized LDL.¹⁰¹ Cigarette smoking, a factor associated with the pathogenesis of atherosclerosis, causes platelet activation, LDL oxidative changes and increases PAF concentrations.¹⁰² The latter alteration was associated with a compensatory increase of PAF-AH activity. However, *in vitro* studies demonstrated that cigarette-derived products as well as oxidative changes of LDL, which physiologically carry PAF-AH, inhibit the activity of the enzyme that catabolizes PAF.¹⁰³ Furthermore, PAF may also oxidize LDL, via stimulation of human monocytes/macrophages and neutrophils to produce superoxide anions and hydrogen peroxide.¹⁰⁴

In summary, experiments performed to investigate the mechanisms responsible for PAF biosynthesis in I/R heart indicated that the synthesis of PAF is probably triggered by the generation of ROS and the resulting oxidative stress occurring at the beginning of reperfusion.

In conclusion, several data comprise evidence for PAF as one of the major GPCR agonists depressing myocardial contractility and thus causing further distress to the ischemic organ in ischemic heart disease.^{4,40} Several *in vivo* and *in vitro* studies demonstrated that PAF is released from I/R heart, and that activation of PAFRs on smooth muscle cells of coronary vessels or on cardiac cells, results in a relay of signals causing a reduction of coronary flow or direct negative inotropic and arrhythmogenic effects. PAF acts as a potent chemoattractant for PLTs and PMNs and promotes their activation and cooperation, thus exacerbating postischemic myocardial injury by capillary plugging and via damage induced by the formation of ROS or proteolytic enzymes. Furthermore, PAF exerts a direct proapoptotic effect on myocardial cells, and interacts with other inflammatory mediators, like TNF- α and arachidonate derivatives, exerting a detrimental action in the I/R heart (Figure 2).

PAF induces ischemic preconditioning

Ischemic preconditioning

In 1986, Murry et al.¹⁰⁵ introduced the term *ischemic preconditioning* (IP) to describe the reduction of infarct size obtained in

canine hearts after applying four 5-minute alternate episodes of circumflex artery occlusion followed by four days of myocardial reperfusion. The efficacy of IP in limiting the severity of I/R injury was confirmed by repeating the IP protocol in several experimental models. IP not only reduces the extent of the infarct size but also the number and severity of conduction arrhythmias and contractile dysfunction. Three aspects can be distinguished in the process of IP: the initial trigger included in the short periods of I/R activates signaling pathways, which in turn act upon an end-effector, which then induces the delay of lethal ischemic damage during sustained ischemia.^{106–108} At first, preconditioning protection lasts a few hours (2–3 h) immediately after the preconditioning procedure. This *early classical preconditioning*, or *first window of protection*, is followed by a period without protection lasting about 12–24 h. Then, the protection reappears and lasts 24–72 h in the so-called *second window of protection* (*late preconditioning*). The mechanistic components underlying the protection afforded by IP have been conventionally classified as *triggers* (factors that act before the index ischemic episode and activate downstream signaling mechanisms) and *mediators/ effectors* (factors that act during the index ischemic episode and mediate the protective effect). This separation is not rigid, because certain signaling components have been demonstrated to act both as triggers and as mediators/ effectors. The stimulation of cell-surface receptors, specifically GPCR, by endogenous ligands, such as adenosine, opioids and bradykinine, generated by the brief ischemic episodes during the preconditioning phase, provide the initial trigger for the signal transduction pathway of protection.¹⁰⁹ Besides these classical agents, other mediators such as TNF- α ,¹¹⁰ chromogranin A-derived peptides,¹¹¹ growth hormone-releasing hormone (GHRH)¹¹² and GHRH-related peptides¹¹³ have been shown to induce cardioprotection in a dose- and time-dependent manner.

The various surface receptor agonist-mediated signal transduction pathways involved in cell protection include at least the following kinases (singly, or in combination, depending on circumstances): PI3K/Akt, PKC ϵ , PKG, p70S6K, extracellular signal-regulated kinase (ERK) 1/2, MAPK and finally phosphorylation of the residual Ser9, that induces inactivation of the 'mitochondria-associated' GSK-3 β pool. These kinases, which are activated at the time of myocardial reperfusion, are termed *reperfusion injury salvage kinases* (RISK). Another important element involved in the IP pathway is mitochondrial ATP-sensitive K⁺ channels (mitoK_{ATP}). Pharmacological activation of mitoK_{ATP} was shown to induce a protective state sensitive to inhibition by mitoK_{ATP} blockers. In addition, the activation of the RISK pathway inhibits the formation of mPTPs during the reperfusion phase following the infarcting ischemia. In fact, the opening of mPTPs completely disrupts mitochondrial function and invariably leads to cell death by either necrosis or by apoptosis. It is likely that a large number of cells are killed by this mechanism during reperfusion (Figure 3).^{109,114}

Low concentrations of PAF induce cardioprotection

Until five years ago, studies performed on the cardiac effects of PAF were mainly devoted to investigating the effects

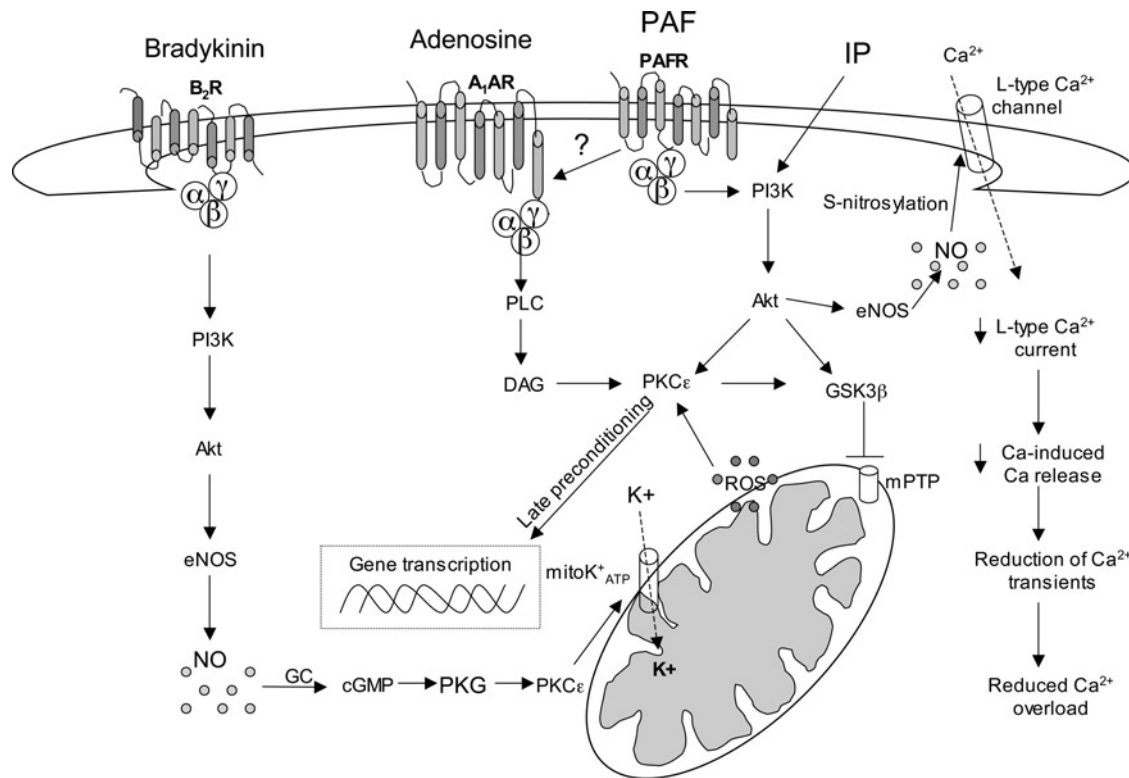


Figure 3 Intracellular signaling pathways involved in the protective effect of classical ischemic preconditioning (IP), bradykinin, adenosine and platelet-activating factor (PAF). PAF-induced protection partially involves adenosine A_1 receptors. A_1AR , A_1 adenosine receptor; Akt, serine/threonine protein kinase; $\alpha\beta\gamma$, G protein subunits; B_2R , B_2 bradykinine receptor; DAG, diacylglycerol; eNOS, endothelial nitric oxide synthase; GC, guanylyl cyclase; GSK3 β , glycogen synthase kinase β ; mPTP, mitochondrial permeability transition pores; mitoK $_{ATP}$, mitochondrial ATP-dependent K $^{+}$ channels; NO, nitric oxide; PAFR, PAF receptor; PI3K, phosphatidylinositol 3-kinase; PKC, protein kinase C; PKG, protein kinase G; PLC, phospholipase C; ROS, reactive oxygen species

exerted by high concentrations of this mediator, comparable to those released in severe pathophysiological conditions, such as after a long-lasting period of ischemia,^{4,40} or acute anaphylactic shock.^{11,115} As described in the above section, cardiac tissue undergoing prolonged I/R releases amounts of PAF high enough to activate inflammatory blood cells and to induce severe myocardial dysfunction, including profound electrophysiological alterations and a negative inotropic effect.^{4,40} We cannot exclude, however, that brief periods of I/R induce the release of very small quantities of PAF, unable to alter myocardial contractility *per se*, but enough to activate kinases involved in IP, such as PKC and PI3K. In fact, PAF is thought to be a mediator of cell-to-cell communication, and some of its above-mentioned actions are reported to be achieved at concentrations as low as 1 pmol/L. However, in many species, such a concentration does not appreciably affect heart function.³⁻⁷ We thus hypothesized that the administration of a very low dose of PAF (in the pmol/L range) could be able to induce protective effects akin to IP and that the activation of PAF receptors might play a role in triggering IP. Several findings indeed suggest that, depending on its concentration, PAF may exert biphasic or, in some cases, opposite effects. For instance, while low doses of PAF (pmol/L range) are able to induce both increment and decrement of CBF, it causes a marked vasoconstriction when infused at higher doses (up to 10 nmol/L).⁷¹⁻⁷⁹ In the central nervous system, although PAF is required for neuronal

survival,¹¹⁶ it can induce apoptosis when administered at elevated concentrations (μ mol/L range).¹¹⁷ The biphasic or even opposite, both beneficial or detrimental, effects exerted by PAF according to its concentration, somehow resemble the behavior of NO, the main end-effector of PAF action on cardiac muscle.¹¹⁸ To verify whether PAF is a possible endogenous agent involved in IP, we performed experiments on isolated rat hearts undergoing 30 min of ischemia followed by two hours of reperfusion. A short treatment of the heart before ischemia with a very low concentration of PAF (20 pmol/L), while unable *per se* to alter cardiac performance, did reduce the extension of infarct size and improved the recovery of left ventricular developed pressure during reperfusion. The cardioprotective effect of PAF was comparable to that observed in hearts in which IP was induced by the classical protocol (brief periods of ischemia separated by reperfusion intervals).¹¹⁹ The fact that the PAF receptor antagonist WEB-2170 abrogated the cardioprotective effect induced by both PAF and IP not only suggested that the action of PAF involves a specific receptor-mediated mechanism but also that endogenous PAF is involved in triggering IP. The protective effect of PAF against postischemic injury has been recently confirmed by Leary *et al.*¹²⁰ by studying postischemic functional recovery in isolated hearts from wild-type and PAF receptor-knockout mice. Postischemic performance was reduced in hearts with targeted deletion of the PAF receptor and in wild-type hearts treated with a PAF receptor

antagonist. Moreover, perfusion with pmol/L concentrations of PAF improved postischemic function in hearts from wild-type mice. To investigate the pathways involved in the preconditioning effect of PAF, the role of several kinases was studied by using pharmacological agents and by Western blot analysis in the isolated rat heart.^{119,121} In a first series of experiments, we observed that the cardioprotective effect of PAF was reduced by both PKC and PI3K inhibitors. In agreement with these findings, Western blot analysis revealed that PAF infusion enhanced the phosphorylation of PKC ϵ and Akt (the downstream target of PI3K) to levels higher than those measured in control hearts, and comparable to those observed after IP treatment.¹¹⁹ However, since the actual protection of IP occurs in the reperfusion phase, an additional series of experiments was performed to study whether the activation of kinases involved in PAF-induced preconditioning persisted also in this latter phase.¹²¹ To test this hypothesis, we studied whether PKC and PI3K inhibitors are able to block the protective effect of PAF, even when the treatment occurs during the initial reperfusion, as well as phosphorylation of PKC ϵ , PKB/Akt, GSK-3 β and ERK1/2 at the beginning of reperfusion. The reduction of infarct size and contractile dysfunctions induced by PAF treatment was abolished by postischemic infusion of chelerythrine or LY294002. A comparable effect was observed in the heart pretreated with PAF and infused with the mPTP opener atracyloside.¹²² During reperfusion, phosphorylation/activation of PKC ϵ , PKB/Akt and the phosphorylation/inactivation of GSK-3 β were enhanced in PAF-treated hearts. We thus concluded that the cardioprotective effect exerted by PAF pretreatment involves activation of PKC and PI3K also in postischemic phases and might be mediated by the prevention of mPTP opening in reperfusion via GSK-3 β inactivation. The role of mitoK_{ATP} channels and ROS in PAF-induced cardioprotection was studied by using N-acetyl-L-cysteine or 5-hydroxydecanoate to scavenge ROS or to block mitoK_{ATP} channels, respectively.¹²² Both these substances abolished the effects of PAF, suggesting that the cardioprotective effect exerted by PAF-pretreatment involves activation of mitoK_{ATP} channels and redox signaling in preischemic phase. We hypothesized that treatment with PAF before ischemia somehow (i.e. with the involvement of mitoK_{ATP}, ROS, PKB/Akt and PKC ϵ) activates a memory function that results in the re-activation of PI3K, PKB/Akt and PKC ϵ at reperfusion. This reactivation represents a pivotal step of the RISK pathway and may allow prevention of mPTP opening at reperfusion.^{108,109} It has indeed been suggested that in the reperfusion phase of preconditioned hearts, PKB/Akt may be upstream to PKC ϵ , which is, in turn, upstream to GSK-3 β . Activation of the PI3K/Akt/NOS signaling pathway was also demonstrated to provide cardioprotection against various stressors by preserving mitochondrial integrity and function. This pathway signaling may converge on the prevention of mPTP opening, leading to cardioprotection.¹²³ In addition, phospho-Akt may also stimulate NOS, the activity of which has been also shown to play an important role in PAF-induced cardioprotection.¹²⁰ In a series of experiments aimed to elucidate the mechanism of the cardioprotective effect induced by

PAF, Leary *et al.* indeed demonstrated that PAF exerts a beneficial effect also on isolated cardiomyocytes (i.e. in the absence of endothelium), via a NO-dependent, S-nitrosylation mechanism. PAF receptor activation attenuates the time-dependent loss of shortening and increase in intracellular Ca²⁺ transients in Ca²⁺-overloaded non-ischemic ventricular myocytes. These protective effects of PAF depend on nitric oxide production, but not on the activation of guanylyl cyclase and production of cGMP.¹²⁰ The fact that reversible S-nitrosylation of myocardial proteins must occur after PAF stimulation is consistent with the hypothesis that low-level PAFR activation initiates NO-induced S-nitrosylation of Ca²⁺ handling proteins, such as L-type Ca²⁺ channels, leading to an attenuation of Ca²⁺ overload¹²⁴ in the heart undergoing I/R (Figure 3). Taken together, these latter results demonstrate that, in addition to stimulating the classical RISK pathways converging on preserving mitochondrial integrity and function, at low concentrations PAF may also provide a protective effect via additional pathways, such as protein S-nitrosylation at the plasmalemmal concentration. The ability of PAF to attenuate Ca²⁺ overload in cardiac cells is consistent with our results obtained in the isolated heart, in which the beneficial effects exerted by PAF involve both myocardial contractility and relaxation (i.e. the reduction in contracture development during reperfusion).^{119,121} An additional important finding of the study of Leary and co-workers is the demonstration that PAF exerts direct protective effects on cardiomyocytes, although in the isolated heart we used as an experimental model, endothelial cells activated by PAF might have participated in PAF-induced protection. Finally, we remind here of the preliminary experiments performed in our laboratory, which suggested that besides the above-mentioned mechanisms, the protective effects of PAF may also involve cross-talk between PAFR and membrane receptors for other mediators known to induce IP. We did indeed find that PAF-induced protection was partially blunted by the adenosine A₁ receptor antagonist 8-cyclopentyl-1,3-dipropylxanthine (Figure 3).¹¹⁹ These results suggest that the protection afforded by PAF not only shares the same cascade of kinases but it may also involve the receptors themselves for other mediators inducing preconditioning of the heart. Although further studies are required to elucidate this interaction, we think this is another interesting piece of a complicated, as yet undiscovered puzzle.

In summary, recent findings demonstrated that brief periods of I/R induce the release of very small quantities (pmol/L range) of PAF, which are unable to alter myocardial contractility *per se*, but enough to activate kinases included in the RISK pathway, such as PKC ϵ , PKB/Akt, GSK-3 β and ERK1/2. Together with the activation of mitochondrial K_{ATP} channels, these events may allow prevention of mPTPs opening at reperfusion, leading to a reduction of infarct size and postischemic contractile dysfunctions, comparable to that afforded by the IP. In parallel to activation of RISK pathways, PAF-dependent NO production and reduction of intracellular Ca²⁺ overload due to NO-dependent S-nitrosylation of L-type Ca²⁺ channels may participate towards the protective effect of PAF.

Conclusions: physiological significance of PAF in cardioprotection

In conclusion, like NO and ROS, PAF may too act as a 'double edged sword' agent on the I/R heart, being able to exert a cardioprotective activity when released at very low concentrations, as occurs after brief ischemic periods. Indeed, low concentrations of exogenous PAF are able to trigger preconditioning-like effects without evident alteration of cardiac function. The signaling events downstream of PAFR stimulation involve the activation of several kinases included in the RISK pathway, thus preventing mPTP opening at reperfusion. Moreover, endogenous synthesis and release of low quantities of PAF, induced by brief I/R periods, might play a crucial role in the triggering of IP. Evidence for preconditioning in humans derives from *in vitro* studies¹²⁵ and observations made during coronary angioplasty and cardiac surgery.^{126–128} It is known that exercise can mimic the protective effect of IP.^{129,130} Indeed, it has been recently demonstrated that exercise delays the appearance of ST segment depression during a subsequent effort in the early and late periods of protection after exercise-induced ischemia in patients with stable angina.¹³¹ Since a very low increase in PAF concentrations occurs during exercise in normoalbuminuric diabetic patients,¹³² it may be argued that low levels of PAF participate in exercise-induced cardioprotection. The fact that low levels of PAF may be released in certain conditions, such as in non-infarcting ischemia¹¹⁹ and atrial pacing,⁴³ or during exercise,¹³² and the fact that these low levels of PAF participate in IP, may be taken into account when strategies of myocardial protection are considered. In particular, these data emphasize the potential importance of a moderate release of PAF, such as that occurring during exercise, as an attempt by the cardiovascular system to protect itself against I/R damages. On the other hand, since inhibition of the PAF protective pathway reduces myocardial postischemic function, the fact that clinical therapies for inflammatory diseases that lead to complete blockade of PAFR may eliminate a significant, endogenous cardioprotective pathway must be taken into account.

Author contributions: GA conceptualized and organized the Minireview, CP and EB co-wrote the first draft of the paper, GA amended the drafts and finalized the figures. All authors reviewed the final draft of the manuscript before submission.

ACKNOWLEDGEMENTS

The authors were supported by funding from Regione Piemonte (CP and GA), the National Institute for Cardiovascular Research (INRC; CP and GA), and from the Italian Ministry of the University and Research (MIUR) and Progetti di Rilevanza Nazionale (PRIN, 2008) (CP).

REFERENCES

- 1 Benveniste J, Henson PM, Cochrane CH. Leukocyte-dependent histamine release from rabbits platelets. The role of IgE, basophils, and a platelet activating factor. *J Exp Med* 1972;**136**:1356–77

- 2 Kravis TC, Henson PM. IgE-induced release of a platelet activating factor from rabbit lung. *J Immunol* 1975;**115**:1677–81
- 3 Chao W, Olson MS. Platelet-activating factor: receptors and signal transduction. *Biochem J* 1993;**292**:617–29
- 4 Montrucchio G, Alloati G, Camussi G. Role of platelet-activating factor in cardiovascular pathophysiology. *Physiol Rev* 2000;**80**:1669–99
- 5 Prescott SM, Zimmerman GA, Stafforini DM, McIntyre TM. Platelet-activating factor and related lipid mediators. *Annu Rev Biochem* 2000;**69**:419–45
- 6 Stafforini DM, McIntyre TM, Zimmerman GA, Prescott SM. Platelet-activating factor, a pleiotropic mediator of physiological and pathological processes. *Crit Rev Clin Lab Sci* 2003;**40**:643–72
- 7 Kasperska-Zaiac A, Brzoza Z, Rogala B. Platelet-activating factor (PAF): a review of its role in asthma and clinical efficacy of PAF antagonists in the disease therapy. *Recent Pat Inflamm Allergy Drug Discov* 2008;**2**:72–6
- 8 Kester M, Ledvora RF, Barany M. The potentiation of arterial contraction with platelet activating factor. *Pflügers Arch* 1984;**400**:200–2
- 9 Benveniste J, Boullet C, Brink C, Labat C. The actions of PAF-acether (platelet-activating factor) on guinea-pig isolated heart preparations. *Br J Pharmacol* 1983;**80**:81–3
- 10 Camussi G, Alloati G, Montrucchio G, Meda M, Emanuelli G. Effect of platelet activating factor on guinea-pig papillary muscle. *Experientia* 1984;**40**:697–9
- 11 Levi R, Burke JA, Guo ZG, Hattori Y, Hoppens CM, McManus LM, Hanahan DDJ, Pinkard RN. Acetyl glyceryl ether phosphorylcholine (AGEPC), a putative mediator of cardiac anaphylaxis in the guinea pig. *Circ Res* 1984;**54**:117–24
- 12 Alloati G, Montrucchio G, Mariano F, Tetta C, De Paulis R, Morea M, Emanuelli G, Camussi G. Effect of platelet-activating factor (PAF) on human cardiac muscle. *Int Arch Allergy Appl Immunol* 1986;**79**:108–12
- 13 Robertson DA, Genovese A, Levi R. Negative inotropic effect on platelet-activating factor on human myocardium: a pharmacological study. *J Pharmacol Exp Ther* 1987;**243**:834–9
- 14 Robertson DA, Wang DY, Lee CO, Levi R. Negative inotropic effect of platelet-activating factor: association with a decrease in intracellular sodium activity. *J Pharmacol Exp Ther* 1988;**245**:124–8
- 15 Alonso F, Gill MG, Sanchez-Crespo M, Mato JM. Activation of 1-alkyl-2-lyso-glycero-3-phosphocholine-acetyl-CoA transferase during phagocytosis in human polymorphonuclear leukocytes. *J Biol Chem* 1982;**257**:3376–8
- 16 Lee TC, Lenihan DJ, Malone B, Roddy LL, Wasserman SI. Increased biosynthesis of platelet activating factor in activated human eosinophils. *J Biol Chem* 1984;**259**:5526–30
- 17 Ninio E, Mencia-Huerta JM, Heymans F, Benveniste J. Biosynthesis of platelet activating factor. 1. Evidence for an acetyl-transferase activity in murine macrophages. *Biochim Biophys Acta* 1982;**710**:23–31
- 18 Bussolino F, Gremo F, Tetta C, Pescarmona GP, Camussi G. Production of platelet activating factor by chick retina. *J Biol Chem* 1986;**261**:16502–8
- 19 Snyder F. Enzymatic pathways for platelet activating factor, related alkyl glycerolipids and their precursors. In: Snyder F, ed. *Platelet-Activating Factor and Related Lipid Mediators*. Chapter 7. New York: Plenum, 1987
- 20 Farr RS, Cox CP, Wardlow ML, Jorgensen R. Preliminary studies of an acid-labile factor (ALF) in human sera that inactivates platelet-activating factor (PAF). *Clin Immunol Immunopathol* 1980;**15**:318–30
- 21 Stafforini DM. Biology of platelet-activating factor acetylhydrolase (PAF-AH, lipoprotein associated phospholipase A2). *Cardiovasc Drugs Ther* 2009;**23**:73–83
- 22 Hwang SB, Ching-Shin CL, Cheah MJ, Shen TY. Specific receptor sites for 1-O-alkyl-2-O-acetyl-sn-glycero-3-phosphocholine (platelet activating factor) on rabbit platelet and guinea pig smooth muscle membranes. *Biochemistry* 1983;**22**:4756–63
- 23 Sugimoto T, Tsuchinochi H, McGregor CG, Mutoh H, Shimizu T, Kurachi DY. Molecular cloning and characterization of the platelet activating factor receptor gene expressed in the human heart. *Biochem Biophys Res Commun* 1992;**189**:617–24
- 24 Korth R, Hirafuji M, Lalau-Keraly C, Delautier I, Bidault J, Benveniste J. Interaction of the Paf antagonist WEB 2086 and its tetrazepine analogue with human platelets and endothelial cells. *Biochem Pharmacol* 1989;**44**:223–9

- 25 Ihida K, Predescu D, Czekay RP, Palade GE. Platelet activating factor receptor is found in a large endosomal compartment in human umbilical vein endothelial cells. *J Cell Sci* 1999;**112**:285–95
- 26 Marrache AM, Gobeil F, Zhu T, Chemtob S. Intracellular signaling of lipid mediators via cognate nuclear G protein-coupled receptors. *Endothelium* 2005;**12**:63–72
- 27 Bito H, Shmizu T. Molecular characterization and physiological functions of PAF receptors. *Adv Exp Med Biol* 1997;**400**:215–21
- 28 Honda Z, Ishii S, Shimizu T. Platelet-activating factor receptor. *J Biochem* 2002;**131**:773–9
- 29 Piper PJ, Stewart AG. Coronary vasoconstriction in the rat isolated perfused heart induced by platelet activating factor is mediated by leukotriene C₄. *Br J Pharmacol* 1986;**88**:595–605
- 30 McIntyre TM, Reinhold SL, Prescott SM, Zimmerman GA. Protein kinase C activity appears to be required for the synthesis of platelet-activating factor and leukotriene B₄ by human neutrophils. *J Biol Chem* 1987;**262**:15370–6
- 31 O'Flaherty JT, Nishihira J. Arachidonate metabolites, platelet activating factor, and the mobilization of protein kinase C in human polymorphonuclear neutrophils. *J Immunol* 1987;**138**:1889–95
- 32 Nakajima T, Sugimoto T, Kurachi Y. Platelet activating factor activates cardiac GK via arachidonic acid metabolites. *FEBS Lett* 1991;**289**:239–43
- 33 Piper PJ, Stewart AJ. Antagonism of vasoconstriction induced by platelet-activating factor in guinea-pig perfused hearts by selective platelet-activating factor antagonists. *Br J Pharmacol* 1987;**90**:771–83
- 34 Dhar A, Paul AK, Shukla SD. Platelet-activating factor stimulation of tyrosine kinase and its relationship to phospholipase C in rabbit platelets: studies with genistein and monoclonal antibody phosphotyrosine. *Mol Pharmacol* 1990;**37**:519–25
- 35 Gomez-Cambronero J, Wang J, Johnson G, Huang CK, Sha'afi RI. Platelet activating factor induces tyrosine phosphorylation in human neutrophils. *J Biol Chem* 1991;**266**:6240–5
- 36 Chao J, Liu H, Hanahan DJ, Olson MS. Platelet activating factor-stimulated protein tyrosine phosphorylation and eicosanoid synthesis in rat Kupffer cells. Evidence for calcium-dependent and protein-kinase C-dependent and C-independent pathways. *J Biol Chem* 1992;**267**:6725–35
- 37 Shimizu T, Mori M, Bito H, Sakanaka C, Tabuchi S, Aihara M, Kume K. Platelet-activating factor and somatostatin activate mitogen-activated protein kinase (MAP kinase) and arachidonate release. *J Lipid Mediat Cell Signal* 1996;**14**:103–8
- 38 Soldi R, Sanavio F, Aglietta M, Primo L, Defilippi P, Marchisio PC, Bussolino F. Platelet-activating factor (PAF) induces the early tyrosine phosphorylation of focal adhesion kinase (p125^{FAK}) in human endothelial cells. *Oncogene* 1996;**13**:515–25
- 39 Nick JA, Avdi NJ, Young SK, Knall C, Gerwins P, Johnson GL, Worthen GS. Common and distinct intracellular signaling pathways in human neutrophils utilized by platelet-activating factor and FMLP. *J Clin Invest* 1997;**99**:975–86
- 40 Stangl V, Baumann G, Stangl K, Felix SB. Negative inotropic mediators released from the heart after myocardial ischemia–reperfusion. *Cardiovasc Res* 2002;**53**:12–30
- 41 Piper HM, Meuter K, Schäfer C. Cellular mechanisms of ischemia–reperfusion injury. *Ann Thorac Surg* 2003;**75**:644–8
- 42 Hoffman JW Jr, Gilbert TB, Poston RS, Silldorff EP. Myocardial reperfusion injury: etiology, mechanisms and therapies. *J Extra Corpor Technol* 2004;**36**:391–411
- 43 Montrucchio G, Camussi G, Tetta C, Emanuelli G, Orzan F, Libero L, Brusca A. Intravascular release of platelet-activating factor during atrial pacing. *Lancet* 1986;**8501**:293
- 44 Stephens CJ, Graham RM, Sturm MJ, Richardson M, Taylor RR. Variation in plasma platelet-activating factor degradation and serum lipids after acute myocardial infarction. *Coron Artery Dis* 1993;**4**:187–93
- 45 Graham RM, Strahan ME, Norman KW, Watkins DN, Sturm MJ, Taylor RR. Platelet and plasma platelet-activating factor in sepsis and myocardial infarction. *J Lipid Mediat Cell Signal* 1994;**9**:167–82
- 46 Montrucchio G, Bergerone S, Bussolino F, Alloatti G, Silvestro L, Lupia E, Cavetto A, Di Leo M, Emanuelli G, Camusi G. Streptokinase induces intravascular release of platelet-activating factor in patients with acute myocardial infarction and stimulates its synthesis by cultured human endothelial cells. *Circulation* 1993;**88**:1476–83
- 47 Annable CR, McManus LM, Carey KD, Pinckard RN. Isolation of platelet-activating factor (PAF) from ischemic baboon myocardium (Abstract). *Fed Proc* 1985;**44**:1271
- 48 Ko W, Hawes AS, Douglas Lazenby W, Calvano SE, Shin YT, Zelano JA, Antonacci AC, Wayne Isom O, Krieger KH. Myocardial reperfusion injury: platelet activating factor stimulates polymorphonuclear leukocyte hydrogen peroxide production during myocardial reperfusion. *J Thorac Cardiovasc Surg* 1991;**102**:297–308
- 49 Leong LLL, Taylor R. The lyso-precursor of platelet-activating factor (lyso-PAF) in ischemic myocardium. *J Lipid Mediat* 1991;**4**:277–87
- 50 Montrucchio G, Alloatti G, Tetta C, De Luca R, Saunders RN, Emanuelli G, Camussi G. Release of platelet-activating factor from ischemic-reperfused rabbit heart. *Am J Physiol Heart Circ Physiol* 1989;**256**:1236–46
- 51 Berti F, Magni F, Rossoni G, De Angelis L, Galli G. Production and biologic interactions of prostacyclin and platelet activating factor in acute myocardial ischemia in the perfused rabbit heart. *J Cardiovasc Pharmacol* 1990;**16**:727–32
- 52 Arnould T, Michiels C, Remacle J. Increased PMN adherence on endothelial cells after hypoxia: involvement of PAF, CD 18/CD 11b, and ICAM-1. *Am J Physiol Cell Physiol* 1993;**264**:1102–10
- 53 Burghardt C, Janero DR. The anoxic rat-heart myocyte produces and release platelet-activating factor (PAF) as a component of its ischemia-like pathology. *J Mol Cell Cardiol* 1987;**19** (Suppl. IV):pS69
- 54 Alloatti G, Montrucchio G, Emanuelli G, Camussi G. Platelet-activating factor induced platelet/neutrophil co-operation during myocardial reperfusion. *J Moll Cell Cardiol* 1992;**24**:163–71
- 55 Chakrabarty S, Fluck DS, Flores NA, Sheridan DJ. Effects of the PAF antagonist BN 50726 and BN 50739 on arrhythmogenesis and extent of necrosis during myocardial ischemia/reperfusion in rabbits. *Br J Pharmacol* 1992;**107**:705–9
- 56 Katoh S, Toyama J, Kodama I, Koike A, Abe T. Role of platelet activating factor in ischemia–reperfusion injury of isolated rabbit hearts: protective effect of a specific platelet activating factor antagonist, TCV-309. *Cardiovasc Res* 1993;**27**:1430–4
- 57 Flores NA, Sheridan DJ. Electrophysiological and arrhythmogenic effects of platelet activating factor during normal perfusion, myocardial ischemia and reperfusion in the guinea-pig. *Br J Pharmacol* 1990;**101**:734–8
- 58 Koltay M, Tosaki A, Hosford D, Esanu A, Braquet P. Effect of BN 50739, a new platelet activating factor antagonist, on ischemia induced ventricular arrhythmias in isolated working rat hearts. *Cardiol Res* 1991;**25**:391–7
- 59 Montrucchio G, Alloatti G, Mariano F, De Paulis R, Comino A, Emanuelli G, Camussi G. Role of platelet-activating factor in the reperfusion injury of rabbit ischemic heart. *Am J Pathol* 1990;**137**:71–83
- 60 Montrucchio G, Alloatti G, Mariano F, Comino A, Cacace G, Polloni R, De Filippi PG, Emanuelli G, Camussi G. Role of platelet-activating factor in polymorphonuclear neutrophil recruitment in reperfused ischemic rabbit heart. *Am J Pathol* 1993;**142**:471–80
- 61 Ko W, Lang D, Hawes AS, Zelano JA, Isom OW, Krieger KH. Platelet activating factor antagonism attenuates platelet and neutrophil activation and reduces myocardial injury during coronary reperfusion. *J Surg Res* 1993;**55**:504–15
- 62 Fontaliran F, Guillon JM, Koltai M, Braquet P. Reduction of infarct size by ginkgolide B (BN 52021) in coronary artery ligated rats. In: Braquet P, ed. *Ginkgolides Chemistry, Biology, Pharmacology and Clinical Perspectives, Barcelona. Chapter 2*. Barcelona: Prous Science, 1989
- 63 Ma XL, Weyrich AS, Krantz S, Lefer AM. Mechanisms of the cardioprotective actions of WEB 2170, bapafant, a platelet activating factor antagonist, in myocardial ischemia and reperfusion. *J Pharmacol Exp Ther* 1992;**260**:1229–36
- 64 Man RY, Kinnaird AA. Similar coronary vascular effects in the rat perfused heart of platelet activating factor structural analogues with agonist and antagonist properties. *Br J Pharmacol* 1995;**116**:2359–64
- 65 Gross GJ, Maruyama M, Vercellotti GM, Jacob HS, Christensen CW. Effect of the PAF antagonist, BN 52021, on myocardial infarct size in dogs. In: Braquet P, ed. *Ginkgolides Chemistry, Biology, Pharmacology and Clinical Perspectives*. Barcelona: Prous Science, 1989;**2**:421–5
- 66 Tamura K, Kimura Y, Tamura T, Kitashiro S, Tzuoka T, Tsuji H, Twasaka T, Inada M. The effect of platelet activating factor antagonist

- TCV-309 on arrhythmias and functional recovery during myocardial reperfusion. *Coronary Artery Dis* 1994;5:267–73
- 67 Feuerstein G, Boyd LM, Ezra D, Goldstein RE. Effect of platelet-activating factor on coronary circulation of the domestic pig. *Am J Physiol Heart Circ Physiol* 1984;246:466–71
 - 68 Montrucchio G, Alloatti G, Mariano F, Meda E, Tetta C, Emanuelli G, Camussi G. The pattern of cardiovascular alterations induced by infusion of platelet activating factor in rabbit is modified by pretreatment with H₁-H₂ receptor antagonists but not by cyclooxygenase inhibition. *Agents Actions* 1987;21:72–8
 - 69 Sybertz EJ, Watkins RW, Baum T, Pula K, Rivelli M. Cardiac, coronary and peripheral vascular effects of acetyl glyceryl ether phosphorylcholine in the anesthetized dog. *J Pharmacol Exp Ther* 1985;232:156–62
 - 70 Jackson CV, Schumacher WA, Kunkel SL, Driscoll EM, Lucchesi BR. Platelet activating factor and the release of platelet-derived coronary artery vasodilator substance in the canine. *Circ Res* 1986;58:218–29
 - 71 Metha J, Wargovich T, Nichols W. Biphasic effects of platelet-activating factor on coronary blood flow in anesthetized dog. *Prostaglandins Leukotrienes Med* 1986;21:87–95
 - 72 Kim YD, Danchak RM, Heim KF, Lees DE, Myers AK. Constriction of canine coronary arteries by platelet activating factor after brief ischemia. *Prostaglandins* 1993;46:69–76
 - 73 Goldstein RE, Ezra D, Laurindo FRM, Feuerstein GZ. Coronary and pulmonary vascular effects of leukotrienes and PAF-acether. *Pharmacol Res Commun* 1986;18:151–62
 - 74 Stahl GL, Lefer DJ, Lefer AM. PAF-acether induced cardiac dysfunction in the isolated perfused guinea pig-heart. *Naunyn-Schmiedeberg's Arch Pharmacol* 1987;336:459–63
 - 75 Viossat I, Chapelat M, Chabrier PE, Braquet P. Effects of platelet-activating factor (PAF) and its receptor antagonist BN 52021 on isolated perfused guinea-pig heart. *Prostaglandins Leukotrienes Essent Fatty Acids* 1989;38:189–94
 - 76 Stahl GL, Lefer AM. Mechanism of platelet-activating factor-induced cardiac depression in the isolated perfused rat heart. *Circ Shock* 1987;23:165–77
 - 77 Hu W, Choy PC, Man RY. Characterization of the coronary vascular responses to platelet-activating factor in the isolated perfused heart. *Lipids* 1991;26:700–4
 - 78 Hu W, Kinnaird AAA, Man RYK. Mechanisms of the coronary vascular effects of platelet-activating factor in the rat perfused heart. *Br J Pharmacol* 1991;103:1097–102
 - 79 Kenzora JL, Perez JE, Bergmann SR, Lange LG. Effects of acetyl glyceryl ether phosphorylcholine (platelet activating factor) on ventricular preload, afterload and contractility in dogs. *J Clin Invest* 1984;74:1193–203
 - 80 Mickelson JK, Simpson PJ, Lucchesi BR. Myocardial dysfunction and coronary vasoconstriction induced by platelet-activating factor in the post-infarcted rabbit isolated heart. *J Mol Cell Cardiol* 1988;20:547–61
 - 81 Tippins JR, Antoniow JW, Alison MR, Garvey B, Maseri A. WEB 2086 inhibits neutrophil dependent increases in coronary resistance in blood perfused rabbit heart. *Cardiovasc Res* 1992;26:162–9
 - 82 Huang Q, Wu M, Meininger C, Kelly K, Yuan Y. Neutrophil-dependent augmentation of PAF-induced vasoconstriction and albumin flux in coronary arterioles. *Am J Physiol Heart Circ Physiol* 1998;275:1138–47
 - 83 Soloviev AE, Braquet P. The role of PAF-acether in the mechanism of isolated coronary artery spasm under hypoxia and its inhibition by BN 52021. In: Braquet P, ed. *Ginkgolides Chemistry, Biology, Pharmacology and Clinical, Barcelona*. Chapter 2. Barcelona: Prous Science, 1990
 - 84 Goldstein RE, Feuerstein GZ, Bradley LM, Stambouly JJ, Laurindo FRM, Davenport NJ. Cardiovascular effects of platelet activating factor. *Lipids* 1991;26:1250–6
 - 85 Alloatti G, Montrucchio G, Mariano F, Tetta C, Emanuelli G, Camussi G. Protective effect of verapamil on the cardiac and circulatory alterations induced by platelet-activating factor. *J Cardiovasc Pharmacol* 1987;9:181–6
 - 86 Riedel A, Mest HJ. The effect of PAF (platelet-activating factor) on experimental cardiac arrhythmias and its inhibition by substances influencing arachidonic acid metabolites. *Adv Prostaglandins Leukotrienes Med* 1987;28:103–9
 - 87 Cervoni P, Herzlinger HE, Lai FM, Tanikella TK. Aortic vascular and atrial response to 1-O-octadecyl-2-acetyl-glycerol-3-phosphocholine. *Br J Pharmacol* 1983;79:667–71
 - 88 Diez J, Delpon E, Tamargo J. Effects of platelet activating factor on contractile force and Ca fluxes in guinea-pig isolated atria. *Br J Pharmacol* 1990;100:305–11
 - 89 Tamargo J, Tejerina T, Delgado C, Barrigon S. Electrophysiological effects of platelet-activating factor (PAF-acether) in guinea pig-papillary muscles. *Eur J Pharmacol* 1985;109:219–27
 - 90 Pietsch P, Hunger T, Braun M, Roediger A, Baumann G, Felix SB. Effect of platelet-activating factor on intracellular Ca²⁺ concentration and contractility in isolated cardiomyocytes. *J Cardiovasc Pharmacol* 1998;31:758–63
 - 91 Gollasch M, Ignatieva V, Kobrinsky E, Vornovitsky E, Zaborovskaya L. Electrophysiological mechanisms responsible for the action of PAF in guinea-pig myocardium. Relation to the putative membrane signalling processes of PAF. *J Lipid Mediat* 1991;3:139–15
 - 92 Wahler GM, Coyle DE, Sperelakis N. Effects of platelet-activating factor on single potassium channel currents in guinea pig ventricular myocytes. *Mol Cell Biochem* 1990;93:69–76
 - 93 Besana A, Barbuti A, Tateyama MA, Symes AJ, Robinson RB, Feinmark SJ. Activation of protein kinase C epsilon inhibits the two-pore domain K⁺ channel, TASK-1, inducing repolarization abnormalities in cardiac ventricular myocytes. *J Biol Chem* 2004;279:33154–60
 - 94 Alloatti G, Levi R, Malan D, Del Sorbo L, Bosco O, Barberis L, Marcantoni A, Bedendi I, Penna C, Azzolino O, Altruda F, Wymann M, Hirsch E, Montrucchio G. phosphoinositide 3-kinase gamma-deficient hearts are protected from the PAF-dependent depression of cardiac contractility. *Cardiovasc Res* 2003;60:242–9
 - 95 Alloatti G, Penna C, De Martino A, Montrucchio G, Camussi G. Role of nitric oxide and platelet-activating factor in cardiac alterations induced by tumor necrosis factor-alpha in the guinea-pig papillary muscle. *Cardiovasc Res* 1999;41:611–9
 - 96 Alloatti G, Penna C, Mariano F, Camussi G. Role of NO and PAF in the impairment of skeletal muscle contractility induced by TNF-alpha. *Am J Physiol Regul Integr Comp Physiol* 2000;279:2156–63
 - 97 Zhao D, Chu WF, Wu L, Li J, Liu QM, Lu YJ, Qiao GF, Wang ZG, Zhang ZR, Yang BF. PAF exerts a direct apoptotic effect on the rat H9c2 cardiomyocytes in Ca²⁺-dependent manner. *Int J Cardiol* 2010;143:86–93
 - 98 Zhao ZQ. Oxidative stress-elicited myocardial apoptosis during reperfusion. *Curr Opin Pharmacol* 2004;4:159–65
 - 99 Alloatti G, Montrucchio G, Camussi G. Role of platelet-activating factor (PAF) in oxygen radical-induced cardiac dysfunction. *J Pharmacol Exp Ther* 1994;269:766–7
 - 100 Lewis MS, Whatley RE, Cain P, McIntyre TM, Prescott TM, Zimmerman GA. Hydrogen peroxide stimulates the synthesis of platelet activating factor by endothelium and induces endothelial cell-dependent neutrophil adhesion. *J Clin Invest* 1988;82:2045–55
 - 101 Lehr HA, Seemuller J, Hubner C, Menger MD, Messmer K. Oxidized LDL-induced leukocyte/endothelium interaction in vivo involves the receptor for platelet activating factor. *Arterioscl Thromb* 1993;13:1013–8
 - 102 Miyaura S, Eguchi H, Johnston M. Effect of a cigarette smoke extract on the metabolism of the proinflammatory autacoid platelet activating factor. *Circ Res* 1992;70:341–7
 - 103 Rouis M, Nigon F, Chapman MJ. Platelet activating factor is a potent stimulant of the production of active oxygen species by human monocyte-derived macrophages. *Biochem Biophys Res Commun* 1988;56:1293–8
 - 104 Tokumura A, Toujima M, Yoshioka Y, Fukuzawa K. Lipid peroxidation in low density lipoproteins from human plasma and egg yolk promotes accumulation of 1-acyl analogues of platelet-activating factor-like lipids. *Lipids* 1996;31:1251–8
 - 105 Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* 1986;74:1124–36
 - 106 Reimer KA, Murry CE, Yamasawa I, Hill ML, Jennings RB. Four brief periods of myocardial ischemia cause no cumulative ATP loss or necrosis. *Am J Physiol* 1986;251:1306–15
 - 107 Dekker LR. Toward the heart of ischemic preconditioning. *Cardiovasc Res* 1998;37:14–20
 - 108 Hausenloy DJ, Yellon DM. Preconditioning and postconditioning: united at reperfusion. *Pharmacol Ther* 2007;116:173–91
 - 109 Hausenloy DJ, Mocanu MM, Yellon DM. Cross-talk between the survival kinases during early reperfusion: its contribution to ischemic preconditioning. *Cardiovasc Res* 2004;63:305–12

- 110 Leucor S, Suleman N, Deuchar GA, Somers S, Lacerda L, Huisamen B, Opie LH. Pharmacological preconditioning with tumor necrosis factor- α activates signal transducer and activator of transcription-3 at reperfusion without involving classic prosurvival kinases (Akt and extracellular signal-regulated kinase). *Circulation* 2005;**112**:3911–8
- 111 Cappello S, Angelone T, Tota B, Pagliaro P, Penna C, Rastaldo R, Corti A, Losano G, Cerra MC. Human recombinant chromogranin A-derived vasostatin-1 mimics preconditioning via an adenosine/nitric oxide signaling mechanism. *Am J Physiol Heart Circ Physiol* 2007;**293**:719–27
- 112 Granata R, Trovato L, Gallo MP, Destefanis S, Settanni F, Scarlatti F, Brero A, Ramella R, Volante M, Isgaard J, Levi R, Papotti M, Alloatti G, Ghigo E. Growth hormone-releasing hormone promotes survival of cardiac myocytes *in vitro* and protects against ischemia–reperfusion injury in rat heart. *Cardiovasc Res* 2009;**83**:303–12
- 113 Alloatti G, Arnoletti E, Bassino E, Penna C, Perrelli MG, Ghé C, Muccioli G. Obestatin affords cardioprotection to the ischemic–reperfused isolated rat heart and inhibits apoptosis in cultures of similarly stressed cardiomyocytes. *Am J Physiol Heart Circ Physiol* 2010;**299**:470–81
- 114 Juhaszova M, Zorov DB, Yaniv Y, Nuss HB, Wang S, Sollott SJ. Role of glycogen synthase kinase-3 β in cardioprotection. *Circ Res* 2009;**104**:1240–52
- 115 Darius H, Lefer DJ, Smith JB, Lefer AM. Role of platelet activating factor-acether in mediating guinea pig anaphylaxis. *Science* 1986;**232**:58–60
- 116 Bozlu G, Atici A, Turhan AH, Polat A, Nayci A, Okuyaz C, Taskinlar H. Platelet-activating factor antagonist (ABT-491) decreases neuronal apoptosis in neonatal rat model of hypoxic ischemic brain injury. *Brain Res* 2007;**1143**:193–8
- 117 Ryan SD, Harris CS, Mo F, Lee H, Hou ST, Bazan NG, Haddad PS, Arnason JT, Bennet SA. Platelet activating factor-induced neuronal apoptosis is initiated independently of its G-protein coupled PAF receptor and is inhibited by the benzoate orsellinic acid. *J Neurochem* 2007;**103**:88–97
- 118 Kelly RA, Balligand JL, Smith TW. Nitric oxide and cardiac function. *Circ Res* 1996;**79**:363–80
- 119 Penna C, Alloatti G, Cappello S, Gattullo D, Berta G, Mognetti B, Losano G, Pagliaro P. Platelet-activating factor induces cardioprotection in isolated rat heart akin to ischemic preconditioning: role of phosphoinositide 3-kinase and protein kinase C activation. *Am J Physiol Heart Circ Physiol* 2005;**288**:2512–20
- 120 Leary PJ, Rajasekaran S, Morrison RR, Tuomanen EI, Chin TK, Hofmann PA. A cardioprotective role for platelet activating factor through NOS dependent S-nitrosylation. *Am J Physiol Heart Circ Physiol* 2008;**294**:2775–84
- 121 Penna C, Mognetti B, Tullio F, Gattullo D, Mancardi D, Moro F, Pagliaro P, Alloatti G. Post-ischemic activation of kinases in the pre-conditioning-like cardioprotective effect of the platelet-activating factor. *Acta Physiol* 2009;**197**:175–85
- 122 Penna C, Mognetti B, Tullio F, Gattullo D, Mancardi D, Pagliaro P, Alloatti G. The platelet activating factor triggers preconditioning-like cardioprotective effect via mitochondrial K-ATP channels and redox-sensible signaling. *J Physiol Pharmacol* 2008;**59**:47–54
- 123 Gustafsson AB, Gottlieb RA. Heart mitochondria: gates of life and death. *Cardiovasc Res* 2008;**77**:334–43
- 124 Sun J, Picht E, Ginsburg KS, Bers DM, Steenbergen C, Murphy E. Hypercontractile female hearts exhibit increased S-nitrosylation of the L-type Ca²⁺ channel α_1 subunit and reduced ischemia/reperfusion injury. *Circ Res* 2006;**98**:403–11
- 125 Speechly-Dick ME, Grover GJ, Yellon DM. Does ischemic preconditioning in the human involve protein kinase C and the ATP-dependent K⁺ channel? Studies of contractile function after simulated ischemia in an atrial in vitro model. *Circ Res* 1995;**77**:1030–5
- 126 Deutsch E, Berger M, Kussmaul WG, Hirshfeld JW Jr, Herrmann HC, Laskey WK. Adaptation to ischemia during percutaneous transluminal coronary angioplasty clinical, hemodynamic, and metabolic features. *Circulation* 1990;**82**:2044–51
- 127 Laskey WK. Beneficial impact of preconditioning during PTCA on creatine kinase release. *Circulation* 1999;**99**:2085–9
- 128 Yellon DM, Alkhulaifi AM, Pugsley WB. Preconditioning the human myocardium. *Lancet* 1993;**342**:276–7
- 129 Crisafulli A, Melis F, Tocco F, Santoboni UM, Lai C, Angioy G, Lorrai L, Pittau G, Concu A, Pagliaro P. Exercise-induced and nitroglycerin-induced myocardial preconditioning improves hemodynamics in patients with angina. *Am J Physiol Heart Circ Physiol* 2004;**287**:235–42
- 130 Lambiase PD, Edwards RJ, Cusack MR, Bucknall CA, Redwood SR, Marber MS. Exercise-induced ischemia initiates the second window of protection in humans independent of collateral recruitment. *J Am Coll Cardiol* 2003;**41**:1174–82
- 131 Michaelides AP, Andrikopoulos GK, Oikonomou EV, Psomadaki ZD, Richter DJ, Dilaveris PE, Exadaktylos NI, Stefanadis CI, Toutouzas PK. Improved myocardial performance during repetitive exercise testing: the role of extracellular superoxide dismutase activity in a model of exercise-induced myocardial preconditioning. *Am Heart J* 2003;**146**:160–7
- 132 Cavallo-Perin P, Lupia E, Gruden G, Olivetti C, De Martino A, Cassader M, Furlani D, Servillo L, Quagliuolo L, Iorio E, Boccellino MR, Montrucchio G, Camussi G. Increased blood levels of platelet activating factor in insulin-dependent diabetic patients with microalbuminuria. *Nephrol Dial Transpl* 2000;**15**:994–9