

Dietary wolfberry ameliorates retinal structure abnormalities in db/db mice at the early stage of diabetes

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Abstract

Hyperglycemia-linked oxidative stress and/or consequent endoplasmic reticulum (ER) stress are the causative factors of pathogenesis of diabetic retinopathy. Dietary bioactive components which mitigate oxidative stress may serve as potential chemopreventive agents to prevent or slow down the disease progression. Wolfberry is a traditional Asian fruit consumed for years to prevent aging eye diseases in Asian countries. Here we report that dietary wolfberry ameliorated mouse retinal abnormality at the early stage of type 2 diabetes in db/db mice. Male mice at six weeks of age were fed the control diet with or without 1% (kcal) wolfberry for eight weeks. Dietary wolfberry restored the thickness of the whole retina, in particular the inner nuclear layer and photoreceptor layer, and the integrity of the retinal pigment epithelia (RPE), and the ganglion cell number in db/db mice. Western blotting of whole retinal cell lysates revealed that addition of wolfberry lowered expression of ER stress biomarkers binding immunoglobulin protein (BiP), protein kinase RNA-like ER kinase (PERK), activating transcription factor 6 (ATF6) and caspase-12, and restored AMP-activated protein kinase (AMPK), thioredoxin, Mn superoxide dismutase (Mn SOD) and forkhead O transcription factor 3 α (FOXO3 α) activities. To determine if our observations were due to the high contents of zeaxanthin and lutein in wolfberry, additional studies using these carotenoids were conducted. Using the human adult diploid RPE cell line ARPE-19, we demonstrated that both zeaxanthin and lutein could mimic the wolfberry preventive effect on activation of AMPK, thioredoxin, Mn SOD, FOXO3 α activities, normalize cellular reactive oxygen species and attenuate ER stress in ARPE-19 cells exposed to a high glucose challenge. The zeaxanthin preventive effect was abolished by small interfering RNA knockdown of AMPK α . These results suggested that AMPK activation appeared to play a key role in upregulated expression of thioredoxin and Mn SOD, and mitigation of cellular oxidative stress and/or ER stress by wolfberry and zeaxanthin and/or lutein. Taken together, dietary wolfberry on retinal protection in diabetic mice is, at least partially, due to zeaxanthin and/or lutein.

Keywords: AMPK, diabetic retinopathy, ER stress, reactive oxygen species, wolfberry, zeaxanthin

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Introduction

Diabetic retinopathy is a retinal damage caused by complications of diabetic mellitus. It is the leading cause of vision impairment and blindness in working age adults. Unfortunately, no permanent cure is currently available. It is well accepted that hyperglycemia is a major causative factor of the progression of the disease.^{1,2} At the early stage of diabetes, retinal blood microvessels are still intact and no observable blood leakage occurs. However, hyperglycemia-induced cellular oxidative stress would alter cellular energy homeostasis, mitochondria biogenesis

and metabolic activity, for instance down-regulation of AMP-activated protein kinase (AMPK).^{3–5}

AMPK is a trimeric serine/threonine kinase and functions as a cellular energy sensor to maintain cellular energy homeostasis. AMPK consists of a catalytic α subunit and non-catalytic β and γ subunits. The α subunit has a NH₂-terminal catalytic kinase domain and a COOH-terminal regulatory domain where the β and γ subunits bind. Phosphorylation of the AMPK α subunit (AMPK α) on Thr172 causes activation of AMPK.³ AMPK has been proposed to mediate hyperglycemia-induced reactive oxygen species (ROS) production in the mitochondria.⁵ Prolonged diabetic inhibition

of AMPK causes mitochondria dysfunction and disruption of cellular ROS homeostasis.^{5,6} Thus, AMPK signaling could be a novel target for diabetic therapy.⁷

Hyperglycemia-linked oxidative stress disrupts protein synthesis and protein folding in the endoplasmic reticulum (ER) among diabetic patients.^{8,9} Studies have suggested that protein unfolding and/or misfolding in the ER would eventually lead to ER stress, and subsequent cell apoptosis.^{10,11} Three signaling pathways are involved in ER stress, which include three ER stress transducer proteins, inositol-requiring protein-1 (IRE1), activating transcription factor 6 (ATF6), protein kinase RNA-like ER kinase (PERK), and an ER stress sensor protein binding immunoglobulin protein (BiP).^{10,12–14} In unstressed cells, BiP binds to and inactivates IRE1, ATF6 and/or PERK. When ER stress occurs, protein expression levels of BiP, IRE1, ATF6 and PERK are elevated. BiP dissociates from all three transducers, which triggers activation of those three transducer-mediated signaling pathways, and subsequently leads to induction of C/EBP homologous protein transcription factor, and activation of c-Jun NH₂-terminal kinase and caspase, which eventually results in apoptosis in diabetes.^{12–14} Thus, ER stress has been considered as a major driven force to cellular oxidative damage in the diabetic retina.¹³

Wolfberry (*Lycium barbarum* L., Chinese name Goji berry) is a fruit which has been consumed for hundreds of years in China and Eastern Asia to prevent eye diseases. It was exported to Western countries in the last century. Fresh wolfberry fruits are bright orange-red and oblong-shaped. They can be purchased fresh or as a dried fruit, drink and/or a wine. Wolfberry contains large amounts of lutein and zeaxanthin in ester forms, which are neuroprotective in the progression of macular degeneration.^{14,15} Zeaxanthin and lutein are dominantly enriched in the retinal pigment epithelium (RPE) and other retinal layers, particularly within the central macula.¹⁴ In addition, wolfberry contains significant amount of polysaccharides and phenolics.^{14,16,17} Small molecules such as betaine, cerebroside, various vitamins and zinc are observed in wolfberry as well.^{14,17,18} According to traditional Chinese medicine literature, wolfberry can nourish liver and kidney, help re-balance the 'Yin' and 'Yan' (i.e. energy homeostasis), boost immune system and improve vision. However, the molecular mechanisms of how the bioactive constituents of wolfberry exert their influence on vision are not well understood.

This study sought to determine the nature of the preventive effect of dietary wolfberry and/or its bioactive components, zeaxanthin and lutein, on diabetic retinopathy using models of the db/db leptin receptor deficient type 2 diabetic mouse *in vivo* and the human RPE cell culture *in vitro*. The results suggested that wolfberry, zeaxanthin and/or lutein ameliorated retinal damage and this could be due to the restoration of AMPK and its downstream target proteins involving attenuation of ER stress and/or oxidative stress in the diabetic retina and in the human RPE cells exposed to a high glucose challenge. These findings could lead to the development of complementary dietary regimens for prevention or delay of the onset of retinopathy at the early stage of type 2 diabetes.

Materials and methods

Animal, wolfberry and design

Six-week-old male wild-type (C57BLKS/J, abbreviated WT) and type 2 diabetic (BKS.C g-m +/+ *Lepr*^{db}/J, abbreviated db/db) mice were purchased from Jackson Laboratory (Bar Harbor, MA, USA). Forty-two db/db and 42 WT mice were equally divided into two treatment groups. One group of mice were fed a regular control diet (CD, *n* = 21) and the other group were fed a control diet with the addition of 1% (kcal) wolfberry (Wolfberry Diet, WD, *n* = 21). The diets were formulated and purchased from the Research Diets Inc. (Table 1) (New Brunswick, NJ, USA).

Wolfberry used in this project was from the same batch of the 'Zhongning' wolfberry which was purchased from Zhongning (Ningxia, China) through a local grocery store. Macronutrient contents of wolfberry fruits were analyzed in the Analytical Service Laboratory at Kansas State University (202 Weber Hall, Manhattan, KS, USA). About 68.2% of the mass of dried wolfberry fruits was carbohydrate, 11.8% was protein and 8.6% was fat, giving a total caloric value in a 100 g serving of about 397 kcal (estimated calories = protein × 4 + carbohydrate × 4 + fat × 9). Thus, the Wolfberry diet contained 10.7 g wolfberry fruits in the CD per 4057 kcal (10.7 g/4057 kcal) (Table 1). Animals were fed their respective diets for eight weeks. Food consumption and blood glucose were tracked on a weekly basis. All experiments conformed to the Association for Research in Vision and Ophthalmology Statement for Use of Animals in Ophthalmic and Vision Research and were performed under an institutionally approved animal protocol.

Mice were group housed (three/cage) in a controlled environment with a 12-h light/dark cycle at a constant room temperature. All animals had free access to water and food throughout the study. After eight weeks, animals were sacrificed by CO₂ according to the approved protocols. Blood samples were collected for plasma insulin measurement. Note, in our preliminary study we observed that CO₂ euthanization did not cause any significant change of insulin concentrations in both animal groups,

Table 1 Diet formulas of the control diet (CD, 10 kcal% fat) and wolfberry diet (WD, control diet with 1% [kCal] wolfberry), formulated by the Research Diets Inc. (New Brunswick, NJ, USA)

Diet ingredient (g)	CD	WD
Casein	200	198.7
L-Cystine	3	3
Corn starch	315	311.4
Maltodextrin 10	35	35
Sucrose	350	346.4
Cellulose	50	48.9
Soybean oil	25	24.5
Lard	20	19.5
Mineral mix S10026	10	10
Dicalcium phosphate	13	13
Calcium carbonate	5.5	5.5
Potassium citrate, 1H ₂ O	16.5	16.5
Vitamin mix V10001	10	10
Choline bitartrate	2	2
Wolfberry	0	10.7
Total	1055	1055.1

WT and db/db, compared with that of cervical dislocation. Heart, kidney, liver and right eyes were removed, weighed and placed in liquid nitrogen immediately, then stored at -80°C until processed for protein extraction. The left eyes of mice were removed and fixed for light microscopy.

Fasting blood glucose test

A drop of blood ($\sim 5\ \mu\text{L}$) was collected from six hours fasting mice by a tail snip. Blood glucose was tested by the Precision Xtra[®] blood glucose monitoring system (Alameda, CA, USA), according to the manufacturer's instruction.

Plasma insulin measurement

Fasting (6 h) blood samples were collected into PST[™] tubes with polymer gel and lithium heparin (BD, Franklin Lakes, NJ, USA). Plasma was separated by centrifugation. Plasma insulin concentration was determined using the ultrasensitive mouse insulin ELISA kit (Crystal Chem, Downers Grove, IL, USA), following the manufacturer's instruction.

Human RPE cell culture and a high glucose challenge

The human RPE cell line ARPE-19 was purchased from the American Type Culture Collection (ATCC; Rockville, MD, USA). Cells were grown in Dulbecco's modified eagle medium (DMEM) (low glucose at 1 g/L or 5.5 mmol/L; Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum (Atlanta Biologicals, Lawrenceville, GA, USA), 50 $\mu\text{g}/\text{mL}$ gentamicin, 0.05 unit/mL penicillin, 50 $\mu\text{g}/\text{mL}$ streptomycin (Invitrogen) at 37°C in an atmosphere of 95% air and 5% CO_2 . Cells at passages 8–14 were used for experiments.

ARPE-19 cells were exposed to 36 mmol/L glucose for 48 h to mimic hyperglycemia *in vitro*. Such high glucose could maximize significant cellular hyperglycemic response. Mannitol was used as an osmotic control (addition of 30.5 mmol/L mannitol into DMEM low glucose medium to achieve 36 mmol/L).

Knockdown of endogenous AMPK α

Endogenous AMPK α was knocked down by small interfering RNA (siRNA) targeting to AMPK α DNA sequence 5' GCA TAT GCT GCA GGT AGA TAA 3'. The AMPK α siRNA was transfected into cells using HiPerFect siRNA transfection reagent according to the instructions provided (Qiagen, Valencia, CA, USA). The cells were left for six hours to knockdown AMPK α prior to experimentation. Knockdown of AMPK α was monitored by Western blot. The vehicle (transfection reagent) and negative control siRNA (AllStars Negative Control siRNA, Qiagen, catalog # 1027281) were used as negative controls.

Detection of cellular ROS

Cellular ROS level was detected by a fluorimetric assay using carboxy-H2DCFDA (C400) as the probe (Invitrogen). One hundred microliters of 4×10^4 cells/mL ARPE-19

cells were seeded into a black 96-well cell culture plate and incubated at 37°C for 12 h in a 5% CO_2 humidified incubator. Following the incubation, cells were treated with high glucose (36 mmol/L) only and/or high glucose with zeaxanthin and/or lutein (at 0.625 and 1.25 $\mu\text{mol}/\text{L}$) for 48 h. Note, zeaxanthin and/or lutein stock solution was prepared at 10 mmol/L in dimethyl sulfoxide (DMSO). In our preliminary experiments, we tested the antioxidant effect of DMSO on ARPE-19 cells, and observed that DMSO under 0.05% had no impact on oxidative stress linked gene expression, and ROS production (data not shown). Cells were then washed with prewarmed phosphate-buffered saline (PBS). One hundred microliters of carboxy-H2DCFDA at 10 $\mu\text{mol}/\text{L}$ in PBS was added in each well and cells were further incubated in the culture incubator for additional 45 min. The fluorescence was quantified using a microplate reader with a fluorescence excitation of 485 nm and emission at 538 nm. Relative fluorescence intensity was normalized to untreated ARPE-19 cells (the PBS group) and graphed.

Profiling wolfberry carotenoids by high-performance liquid chromatography

The wolfberry carotenoid was profiled by high-performance liquid chromatography (HPLC) by Craft Technologies, Inc. (Wilson, NC, USA) (<http://www.crafttechnologies.com/>). Carotenoid content was expressed as $\mu\text{g}/\text{g}$ fruits (dried fruits).

Determination of water soluble carbohydrates

Water soluble carbohydrate of wolfberry fruits was determined by the anthrone method.¹⁹ Fruits were blended into a powder. The powder was mixed with 4.5 volume of deionized water and boiled for 30 min. The mixture was filtered with four layers of cheese cloth. The aqueous phase was harvested and subjected to further centrifugation at 1500g for 30 min at 4°C (Sorvall RC-5, Thermo Scientific, Waltham, MA, USA). The supernatants were freeze-dried. The freeze-dried powder was reconstituted in anthrone solution to achieve the solution at 2 g/L anthrone in 75% H_2SO_4 . The mixture was boiled at 100°C for 15 min. After the samples were cooled down, the absorbance was read at 578 nm using a spectrophotometer. Glucose was used as a standard.

Measurement of total phenolics

Wolfberry fruits were blended into a powder. The powder was mixed with 10 volume of 95% ethanol, with stirring for 24 h at 4°C . Total phenolics were then extracted by reflux extraction three times, two hours of each at room temperature. The mixture was filtered, and aqueous phase was harvested and subjected to further evaporation to get rid of ethanol. The remaining mass was freeze-dried. The freeze-dried powder was reconstituted in PBS to achieve water soluble solution for total phenolic measurement.

The amount of total phenolics was determined according to the Folin-Ciocalteu method.²⁰ Ten microliters of wolfberry water soluble solution (at 1 mg/mL in deionized water) was mixed with 50 μL Folin-Ciocalteu's reagent and 50 μL Na_2CO_3 solution (20%, w/v), then incubated

for two hours at room temperature. After that the absorbance of samples was measured at 760 nm using a spectrophotometer. Results were expressed as milligram quercetin equivalent per gram wolfberry fruits.

Western blot

Alteration of retinal protein expression and phosphorylation was determined by Western blot as described.²¹ Whole retina was dissected under a dissecting microscope. Whole retinal cell lysates and ARPE-19 cell lysates were achieved as described previously.^{21,22} The primary and secondary antibodies were purchased from Cell Signaling Tech. (Danvers, MA, USA). Immunoreactive bands were detected by chemiluminescence (ECL; Thermo Scientific Pierce, Rockford, IL, USA) and visualized by the FluorChem 8800 advanced image system (Alpha Innotech, San Leandro, CA, USA). β -Actin was used as a equal loading control. Total pixel intensity of each protein band was normalized to β -actin, and was then used for graphing and statistic analysis.

Light microscopy

Mouse retinal microscopy was performed as previously described.²² Eye balls were collected and then fixed in a fixative containing 2% paraformaldehyde, 2.5% glutaraldehyde (Sigma-Aldrich, St Louis, MO, USA) and 0.1mol/L cacodylate, and then postfixed with osmium tetroxide (Electron Microscopy Sciences, Fort Washington, PA, USA). Dehydration was in 70% ethanol for 12 h, overnight with 100% ethanol, and 12 h with 100% acetone and finally embedding in Epon LX112 (Electron Microscopy Sciences). Sections were taken perpendicular to the optic nerve so that all of the layers of the retina could be observed. Eyes were sectioned until the optic nerve was observed, so that the depth was consistent in all of the sections. Thick sections (1 μ m) were obtained and stained with 1% (W/V) toluidine blue O (Electron Microscopy Sciences) and viewed using a Nikon Eclipse E600 light microscope (Nikon Inc., Melville, NY, USA). The thickness of central retina, the ganglion cell layer (GCL), the inner plexiform layer (IPL), the inner nuclear layer (INL), the outer plexiform layer (OPL), the outer nuclear layer (ONL), and the photoreceptor layer (including the outer segment [OS] and the inner segment [IS]) were measured at the site of 300 μ m away from the center of the optic nerves, and analyzed using analySIS software (Soft Imaging System GmbH, Munster, Germany). The ganglion cell number per 100 μ m of the GCL was counted in the central retinal regions (between 300 and 400 μ m away from the center of the optic nerves).^{23,24} Structural alteration was analyzed and photographed in the same area of the central retina (around 300 μ m away from the center of the optic nerves).

Statistical analysis

All results were expressed as mean $\pm$ SD. *n* represents the number of mice in each experimental group. Differences in measured variables corresponding to dietary treatment (control and wolfberry) and genotype (WT and db/db

mice) were tested using two-way analysis of variance (ANOVA). Data normality was tested using the D'Agostino-Pearson omnibus test. Some data were also tested by the Q-test. The ARPE-19 cell culture experiments were triplicate. The results were analyzed by one-way ANOVA with Dunnett's multiple comparison using SAS 9.1. Representative images of Western blot and light microscopy on retinal structure were shown. The level of significance was considered at $P \leq 0.05$.

Results

Wolfberry is enriched in zeaxanthin, water soluble carbohydrate and phenolics

Carotenoid HPLC analysis results showed that wolfberry fruits contained a significant amount of zeaxanthin (at 1761.3 μ g/g fruits), lutein (at 50.9 μ g/g fruits) and β -cryptoxanthin (at 25.8 μ g/g fruits) (Table 2). Analytical results showed that wolfberry fruits used in this study contained water soluble carbohydrates at $25.2 \pm 1.5\%$ (w/w), and phenolics at 7.8 ± 0.4 mg quercetin equivalent/g fruits.

Wolfberry does not lower blood glucose and insulin in db/db type 2 diabetic mice

db/db type 2 diabetic and the genetic background WT mice were fed with CD and/or control diet with addition of 1% (kcal) wolfberry (WD) for eight weeks, starting at six weeks of age. The diet formulas were shown in Table 1. The results demonstrated that db/db mice consumed larger amount of diets than that of WT mice; however, daily food consumption rates of db/db mice were not different within CD and WD diets (Figure 1a). WT mice had greater heart:body weight and kidney:body weight, but lower liver:body weight than those of db/db mice ($P < 0.01$, $n = 21$) (Table 3). Supplementation of the wolfberry at 1% (kcal) did not cause significant difference in the heart, liver, kidney and overall body weight of either db/db or WT mice (Table 3 and Figure 1b).

Fasting blood glucose and plasma insulin were further determined in those animals. As shown in Figure 1c, WT mice had regular fasting blood glucose concentrations that did not differ by diet. Fasting blood glucose concentrations were elevated in db/db mice fed CD starting at six weeks of age, indicating the development of hyperglycemia. Addition of wolfberry did not lower fasting blood glucose concentration (WD versus CD). Plasma insulin was

Table 2 Wolfberry carotenoid profile by high-performance liquid chromatography

Carotenoid	μ g/g wolfberry fruits
Zeaxanthin	1761.3
Lutein	50.9
β -Cryptoxanthin	25.8
α -Cryptoxanthin	<0.1
Lycopene	<0.1
cis-lycopene	<0.1
α -Carotene	<0.1
β -Carotene	<0.1
cis- β -carotene	<0.1

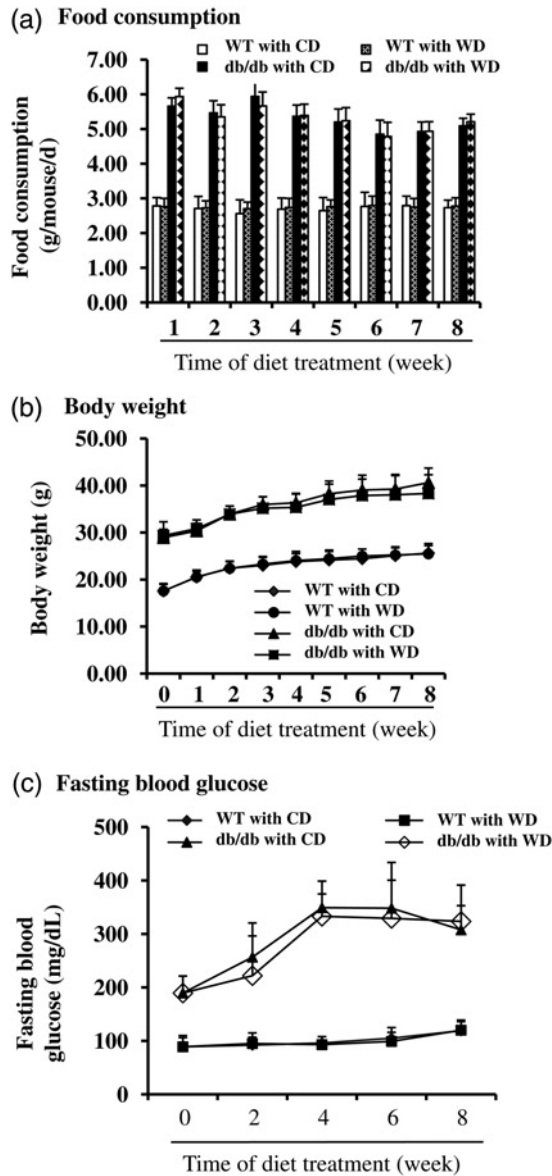


Figure 1 Food consumption, body weight and fasting blood glucose in wild-type (WT) and db/db type 2 diabetic mice. Six-week old male WT and db/db diabetic mice (db/db) were fed the control diet (CD) or the control diet with 1% (kcal) wolfberry (WD) for eight weeks. Food consumption (a) and body weight (b) were monitored weekly. Fasting blood glucose (c) was tracked biweekly. $n = 21$

measured at study termination (for 8 weeks) (Table 3). The result showed that db/db mice at 14 weeks of age had significant higher insulin concentration than that of WT mice ($P < 0.05$). No difference was observed by diet (CD versus WD), suggesting that wolfberry at 1% (kcal) did not systemically improve hyperglycemia and insulin resistance in db/db type 2 diabetic mice.

Wolfberry ameliorates retinal abnormality in db/db type 2 diabetic mice

Retinal thickness and structural alteration were determined by light microscopy. We defined that the central retina thickness is the distance from the GCL to the RPE layer in

the central area of retina which is located at around 300 μm away from the center of optic nerves. As shown in Figure 2a, the whole central retina in db/db type 2 diabetic mice was significantly thinner than that of WT mice ($P < 0.05$, $n = 21$). Application of dietary wolfberry significantly improved retina thickness in db/db mice (CD versus WD, $P < 0.05$). Overall, whole retina thickness was not altered in WT mice with or without wolfberry treatment.

Additionally, we measured the thickness of the GCL, IPL, INL, OPL, ONL and photoreceptor layer (IS + OS) and found that the INL and photoreceptor layer (IS + OS) were significant thinner in db/db mice at 14 weeks of age than that of WT mice ($P < 0.05$). However, no significant change was observed in the thickness of the GCL, IPL, OPL and ONL. Wolfberry improved thickness of the photoreceptor layer and INL in db/db mice (Figure 2a, data on OPL and GCL were not shown).

Ganglion cell number was measured in the central retinal region (Figure 2b). There were significantly fewer ganglion cells in diabetic db/db mice fed CD than that of WT mice. Application of wolfberry completely improved the loss of ganglion cells in db/db mice. Ganglion cell number was constant in WT mice with or without wolfberry treatment.

We further investigated retinal structural change in mice with or without wolfberry treatment. As shown in Figure 2c, nuclear distribution in the ONL was much disorganized in db/db mice fed CD ($n = 21$), compared with WT mice, although overall thickness of the ONL was not significantly changed (Figure 2a). We consistently observed a channel-like structure (arrowed in the inserted image) in the RPE layers of all 21 db/db mice fed CD. No such structure was found in either db/db fed WD, or WT fed CD or WD. Thus we excluded any artifact of the eye tissue processing. This structure might be a very early sign of the disruption of integrity of the RPE layer, although no similar structure has been reported in human diabetic retinopathy. Improvement of those retinal abnormalities was apparently observed in db/db mice fed dietary wolfberry (Figure 2c, the far right panel). Wolfberry caused no structural alteration in WT mice.

Wolfberry attenuates ER stress in retina of db/db type 2 diabetic mice

Next, we determined the mechanism of wolfberry on restoration of retinal structure in db/db mice. After dietary wolfberry treatment for eight weeks, the retinal cell lysates of WT and db/db mice were analyzed by Western blot to monitor alteration of ER stress protein expression. Results in Figures 3a–e demonstrated that elevated protein expression of ER stress biomarkers, BiP, PERK, ATF6, and active caspase-12 occurred in db/db mice fed the CD, compared with those of WT mice fed the CD, indicating that db/db mice developed retinal ER stress at 14 weeks of age. Addition of 1 kcal% dietary wolfberry significantly mitigated retinal ER stress as demonstrated by lowered expression of ER stress biomarkers in db/db mice, and in some cases the levels did not differ by diet (CD versus WD) in WT mice ($P < 0.05$) (Figures 3a–e). Caspase-3

Table 3 Dietary wolfberry does not alter mouse organ weight and fasting plasma insulin level (mean $\pm$ SD, $n = 21$)

Animal	Diet	Kidney/BW (g/g)	Liver/BW (g/g)	Heart/BW (g/g)	Fasting plasma insulin (ng/mL)
WT	CD	0.016 $\pm$ 0.0013	0.038 $\pm$ 0.0018	0.0061 $\pm$ 0.0005	0.71 $\pm$ 0.329
	WD	0.017 $\pm$ 0.0012	0.039 $\pm$ 0.0021	0.0062 $\pm$ 0.0004	0.53 $\pm$ 0.302
db/db	CD	0.011 $\pm$ 0.0009*	0.065 $\pm$ 0.0035 [†]	0.0028 $\pm$ 0.0003 [‡]	3.69 $\pm$ 1.094 [§]
	WD	0.011 $\pm$ 0.0007*	0.061 $\pm$ 0.0029 [†]	0.0031 $\pm$ 0.0003 [‡]	4.42 $\pm$ 1.863 [§]

BW, body weight; CD, control diet; WD, wolfberry diet; WT, wild-type mouse

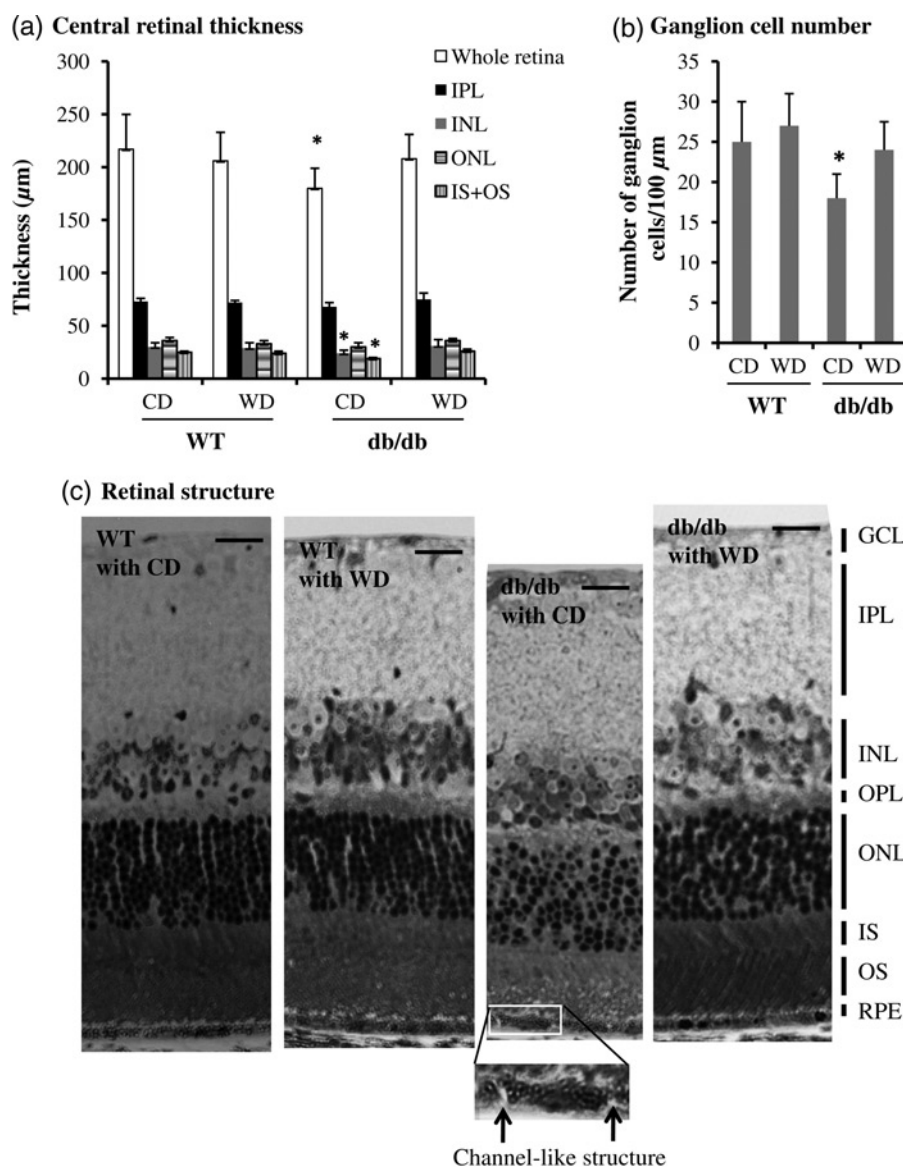
* $P < 0.01$ versus WT group with CD[†] $P < 0.01$ versus WT group with CD[‡] $P < 0.01$ versus WT group with CD[§] $P < 0.05$ versus WT group with CD

Figure 2 Wolfberry improves retinal abnormality in db/db type 2 diabetic mice. After dietary treatments mouse eye balls were fixed, and thick retina sections were prepared as described in the Materials and methods section. (a) The central retinal thickness, and the inner plexiform layer, inner nuclear layer, outer nuclear layer and photoreceptor layer (IS + OS) were measured and graphed ($n = 21$). * $P < 0.05$ versus WT with CD. (b) Ganglion cells were counted and graphed as described in the Materials and methods section. Loss of ganglion cells in the db/db mouse fed CD was significant. * $P < 0.05$ versus WT with CD. (c) Retinal structure of WT and db/db mice after dietary treatments. The inserted picture showing a channel-like structure in the RPE cell layer of db/db type 2 diabetic mice (arrowed) ($n = 21$). GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; ONL, outer nuclear layer; IS, inner segment layer; OS, outer segment layer; RPE, retinal pigment epithelium layer; WT, wild-type mice; db/db, db/db type 2 diabetic mice; CD, control diet; WD, control diet with 1% (kcal) wolfberry. Scale bar, 10 μ m

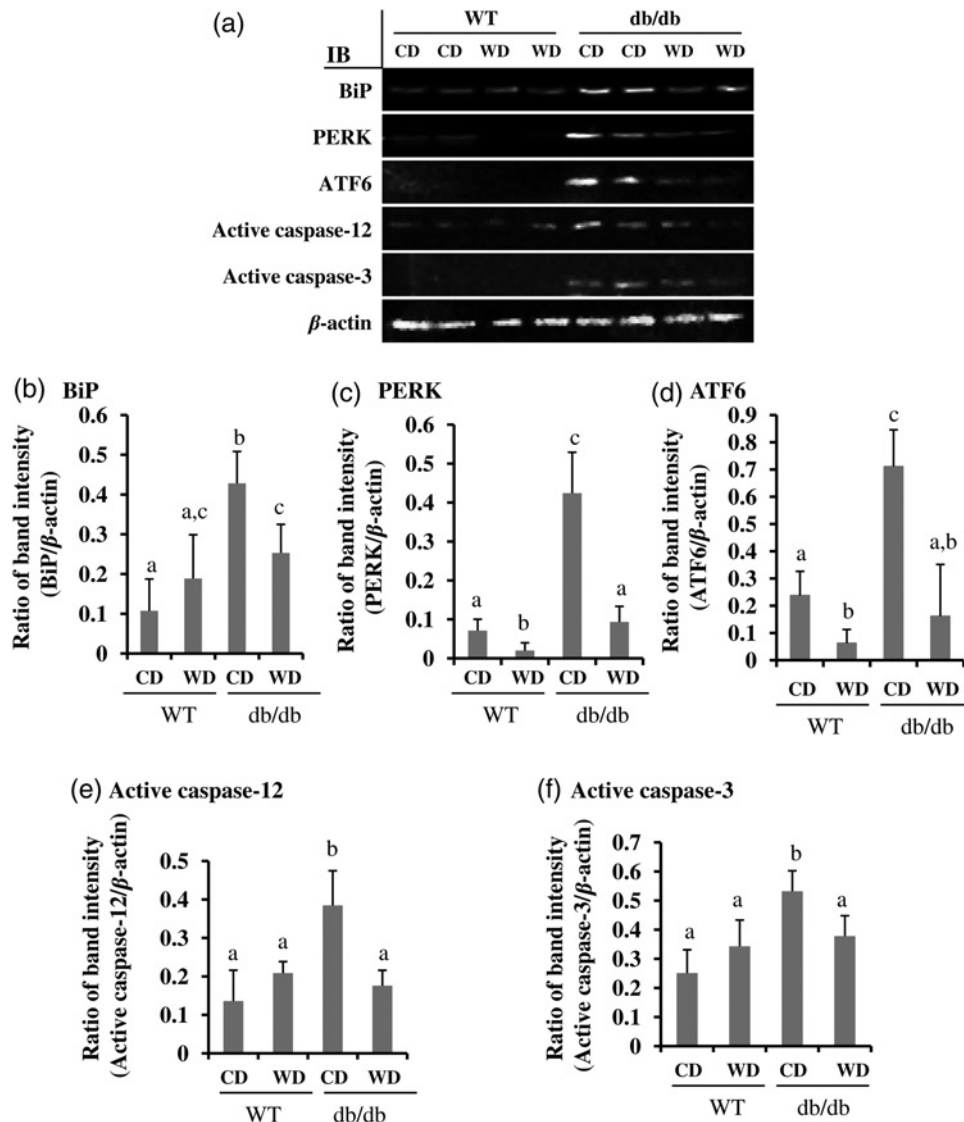


Figure 3 Attenuation of endoplasmic reticulum (ER) stress by dietary wolfberry in db/db type 2 diabetic mice. After dietary treatments, animals were euthanized. The whole retina was isolated and subjected to Western blot. Representative images were shown (a). β -Actin was used as an equal loading control to normalize each protein expression. Relative band intensity of each protein to β -actin was used for graphing and statistic analysis. $n = 21$. In all cases, the statistic significance at $P \leq 0.05$ was marked by different letters a, b and/or c. (a) Representative images; (b) BiP; (c) PERK; (d) ATF6; (e) active caspase-12; (f) active caspase-3. CD, control diet; WD, control diet with 1% (kcal) wolfberry; BiP, binding immunoglobulin protein (also named as glucose regulated protein 78, GRP78); PERK, protein kinase RNA-like ER kinase; ATF6, activating transcription factor-6; IB, immunoblot; WT, wild type

activation in retina of db/db mice was significantly inhibited by wolfberry ($P < 0.05$) (Figures 3a and f).

Wolfberry induces protein expression of AMPK, FOXO3 α and antioxidant enzymes thioredoxin and Mn superoxide dismutase in retina of db/db type 2 diabetic mice

As shown in Figures 4a–c, AMPK α and phospho-AMPK α Thr172 proteins were significantly down-regulated in retina of db/db mice fed CD, which was normalized by feeding WD ($P < 0.05$). Phospho-AMPK α Thr172, but not total AMPK α proteins, was elevated significantly in the wolfberry treated retina of WT mice. The results suggested that addition of wolfberry restored AMPK activity in diabetic retina.

Thioredoxin and Mn superoxide dismutase (Mn SOD) are major antioxidant enzymes in scavenging cellular ROS in the cytoplasm and mitochondria, respectively.^{25–27} As shown in Figures 4a, d and e, the protein expression of thioredoxin and Mn SOD was inhibited in the retina of db/db mice fed CD ($P < 0.05$). Wolfberry treatment in db/db mice caused elevated expression of thioredoxin and Mn SOD which were comparable to WT mice.

Forkhead O transcription factor 3 α (FOXO3 α) dually functions as a proapoptotic and/or antiapoptotic transcription factor in response to stress.^{28,29} Activation of FOXO3 α protects cells from oxidative stress in diabetes.^{30,31} The results in Figures 4a, f and g demonstrated that protein concentrations of phospho-FOXO3 α Ser253 and total FOXO3 α decreased significantly in the retina of db/db diabetic mice fed CD, which were restored by wolfberry ($P < 0.05$).

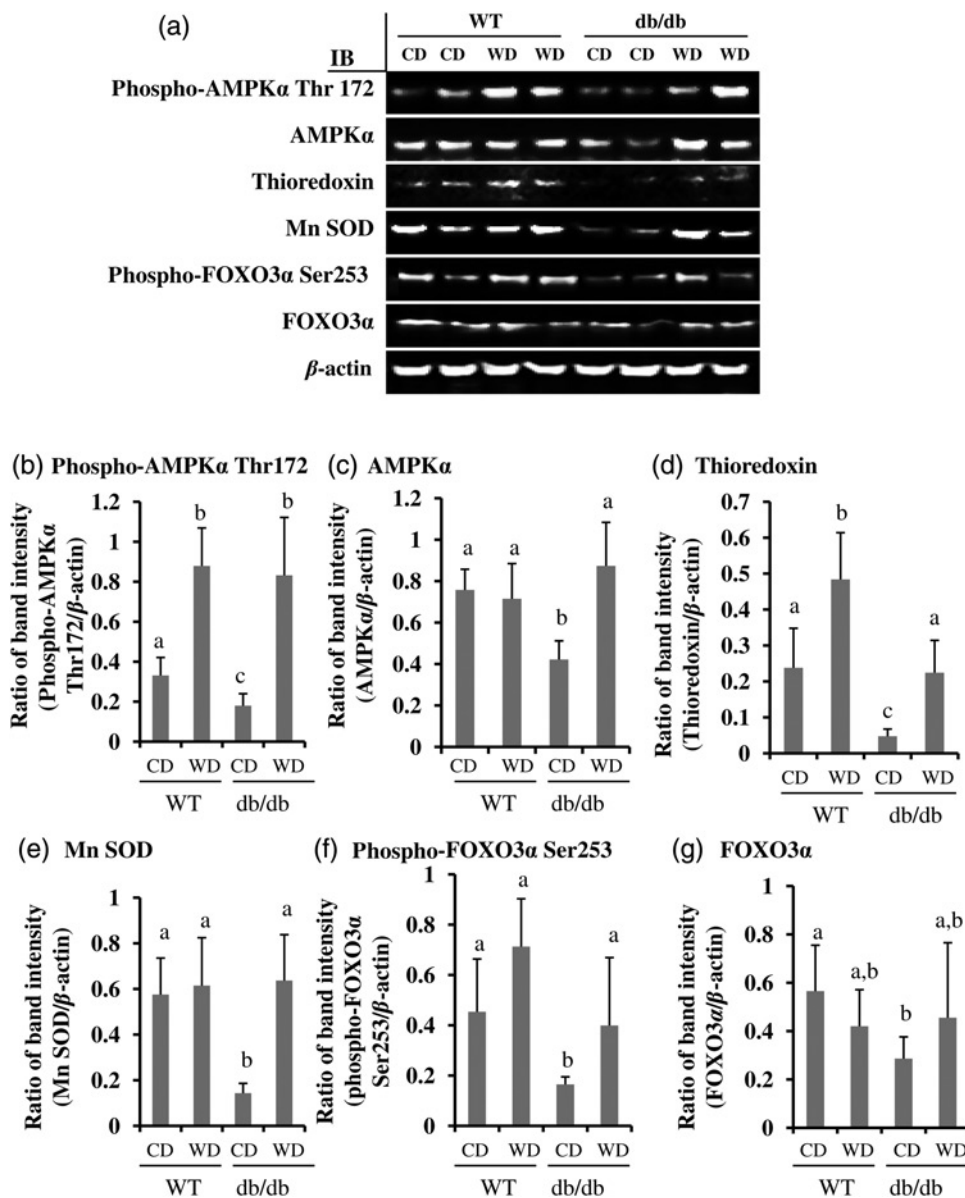


Figure 4 Wolfberry restores AMPK, FOXO3α and antioxidant enzymes thioredoxin and Mn SOD in the retina of db/db type 2 diabetic mice. Whole retina was isolated and subjected to Western blot at study termination. Representative images were shown (a). β-Actin was used as an equal loading control to normalize each protein expression. Relative band intensity of each protein to β-actin was used for graphing and statistic analysis. $n = 21$. The statistic significance at $P \leq 0.05$ was marked by different letters a, b and/or c. (b) phospho-AMPKα Thr172; (c) AMPKα; (d) thioredoxin; (e) Mn SOD; (f) phospho-FOXO3α Ser253; and (g) FOXO3α. CD, control diet; WD, control diet with 1% (kcal) wolfberry; WT, wild type; AMPKα, AMP activated protein kinase α; Mn SOD, Mn superoxide dismutase; FOXO3α, forkhead O transcription factor 3α

Wolfberry did not alter FOXO3α expression and phosphorylation on Ser253 in the retina of WT mice.

Activation of AMPKα by zeaxanthin and/or lutein protects human RPE cell culture against a high glucose challenge

Finally, we wished to determine what potential bioactive constituents of wolfberry exert the influence on retinal protection in db/db mice. Zeaxanthin and lutein, which are abundant in wolfberry, have been documented to be specifically transported and accumulated in the RPE and other retinal layers, and act as a potential antioxidant in the prevention of macular degeneration.^{32,33} Here we used a

human adult diploid RPE cell line ARPE-19 to determine their protective effects on a high glucose challenge. In the preliminary study we investigated the glucose dose response in APRE-19 cells and found that exposure to 36 mmol/L glucose could achieve significant hyperglycemic effects in cells, which was similar to human diabetic retina, for instance, production of ROS, activation of ER stress and apoptosis. Thus, in this report we used 36 mmol/L glucose to mimic hyperglycemia condition *in vitro*. Mannitol was used as an osmotic control. Zeaxanthin and/or lutein were simultaneously added into the culture medium for 48 h. Then whole-cell lysates were subjected to Western blot. Data showed that the expression levels of proteins (e.g. phospho-AMPKα Thr172, AMPKα, FOXO3α, thioredoxin,

Mn SOD) in high glucose challenged cells were significantly lower than those in PBS and/or mannitol groups. PERK protein levels were significantly elevated in the cells exposed to high glucose (Figure 5a, $P < 0.05$, $n = 3$). Both zeaxanthin and/or lutein could mimic the wolfberry preventive effect on activation of AMPK α , FOXO3 α , Mn SOD, thioredoxin and inhibition of PERK (Figure 5a, $P < 0.05$, $n = 3$, only AMPK and PERK mean data shown in lower graphs). Further, we determined that high glucose induced generation of cellular ROS in ARPE-19 cells. Zeaxanthin and/or lutein mitigated ROS generation dose dependently in ARPE-19 cells exposed to a high glucose challenge (Figure 5b, $P < 0.05$, $n = 3$, gray columns).

To confirm that AMPK α activation is essential to diminished cellular ROS generation by a high glucose challenge, we knocked down AMPK α by siRNA in the zeaxanthin-treated (1.25 $\mu\text{mol/L}$) ARPE-19 cells under a high glucose challenge (36 mmol/L glucose). Vehicle only (HiPerfect transfection reagent) and the negative control siRNA were used as negative controls. Results clearly demonstrated that knockdown of AMPK α significantly abolished the zeaxanthin effect on normalization of cellular ROS level induced by high glucose ($P < 0.05$) (Figure 5b, far right three black columns: AMPK α siRNA versus Vehicle or Negative siRNA; Figures 6a and b). Western blot results further showed that knockdown of AMPK α caused significant decreases in protein expression of FOXO3 α , Mn SOD and thioredoxin, but increases in PERK in high glucose challenged ARPE-19 cells treated with zeaxanthin at 1.25 $\mu\text{mol/L}$ ($P < 0.05$) (Figures 6a and b, each protein expression was normalized to β -actin). Data suggested that AMPK α modulated downstream activities of FOXO3 α , thioredoxin and Mn SOD. Zeaxanthin diminished high glucose-induced cellular ROS production and/or ER stress via restoration of AMPK α .

Discussion

In the current study, we investigated the preventive effects of wolfberry on hyperglycemia-induced retinal damage by application of dietary wolfberry at 1% (kcal) to the db/db mouse, starting at six weeks of age while the fasting blood glucose concentration had been elevated to 189.4 ± 31.9 mg/dL. The results have demonstrated for the first time that RPE integrity and retinal structure were altered at the early stage of diabetes in the db/db mouse model, and this abnormality could be ameliorated by dietary wolfberry.

The genetically programmed diabetic db/db mouse is an adequate model for studies on the pathogenesis of retinopathy; no retinal disorder occurs in the new born mouse.³⁴ It has been reported that the db/db mouse starts to develop hyperglycemia at eight weeks of age,³⁵ which was very similar to our observation. Breakdown of blood-retinal barrier and leakage of retinal microvessels occurs at 18 weeks of age. About seven weeks later, severe microvessel leakage and retinopathy were observed.^{36–40} In the current study we observed that fasting blood glucose concentration was elevated to >300 mg/dL at 10 weeks of

age in db/db fed CD but not WT mice, indicating that the db/db mouse developed hyperglycemia and/or hyperinsulinemia (Figure 1c) at the similar age as previously reported.^{34,35} According to publications, retinal blood microvessels may still be intact; no blood leakage occurs at this early stage of diabetes in db/db mice.^{36–40} Thus, the young db/db mouse (<18 weeks of age) would be an excellent type 2 diabetic model for the study in prevention of the development of diabetic retinopathy.

Hyperglycemia induces biochemical abnormality in diabetic cells and tissues, which leads to retinal degeneration. Diabetic retinal degeneration is associated with ganglion cell loss, and reduction of retinal thickness including INL, IPL and/or ONL.^{23,41–43} The db/db leptin receptor deficient mouse is a spontaneous type 2 diabetic mouse model. Our findings in loss of ganglion cells was consistent with the previous report.⁴¹ We also found for the first time that INL and photoreceptor layers were abnormal compared with WT mice (C57BLKS/J).

Wolfberry fruits have been most popularly consumed as a part of Chinese cuisine although it is also considered as an herbal medicine. Wolfberries are being sold as herbal supplements in Western countries as well.¹⁴ Consumption of wolfberries is beneficial to general health, including vision.^{14–18} In this study we applied a low-dose (1% (kCal)) wolfberry to both WT and db/db mice to mimic what humans would eat on a practical basis. Apparently both types of animals fed the wolfberry supplemented diets survived as well as those fed the CD, as shown in gain of body mass and overall organ weight, suggestive of no obvious side-effects at the dose range. However, wolfberry at 1% (kcal) did not systematically affect the levels of fasting blood glucose and insulin in the diabetic mouse at 14 weeks of age. This may be due to the retinal specific accumulation of its bioactive components zeaxanthin and lutein in mice as discussed below.

The bioactive constituents of wolfberry have been well characterized. The major groups of substance in the wolfberry are carotenoids, polysaccharides and flavonoids.¹⁴ Zeaxanthin and lutein dipalmitates are the most dominant and unique constituents of wolfberry carotenoid fractions.^{14,15} Zeaxanthin and lutein are specifically transported to and preferentially taken up, and accumulated in retinal cells by sharing the transportation pathway of HDL cholesterol.^{32,33} It has been suggested that in the retina, zeaxanthin and/or lutein bind to antioxidant enzymes, which in turn stabilize the proteins under chronic oxidative stress in the case of macular degeneration.^{32,40–46} Recently, Gierhart and colleagues⁴⁷ reported that zeaxanthin administration at 0.02% or 0.1% (diet by weight) significantly inhibited diabetes-induced retinal oxidative damage, but no effect was achieved on the severity of hyperglycemia in rats at the early stage of diabetes. In the current study we used wolfberry fruits and pure zeaxanthin and/or lutein to treat diabetic mice and human RPE cells, respectively. The results suggested that wolfberry protected retinal cells against a high glucose challenge locally but not systemically, at least partially, if not all, through zeaxanthin and/or lutein, though there may be some potential synergistic interactions of

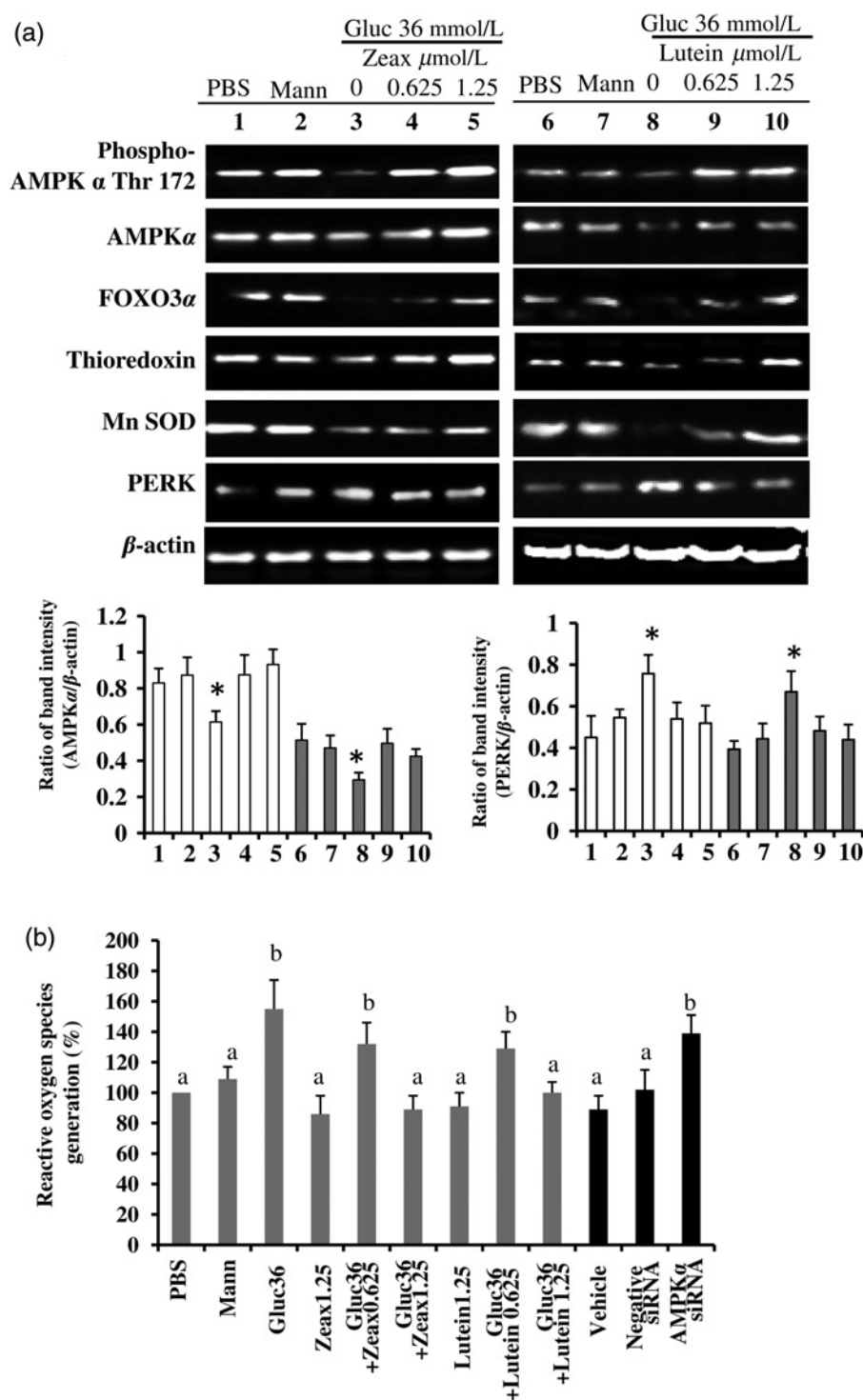


Figure 5 Zeaxanthin and/or lutein activation of AMPK protects human retinal pigment epithelium (RPE) cell culture against a high glucose challenge. Human adult diploid RPE cell line ARPE-19 cells were treated with 36 mmol/L glucose for 48 h to mimic hyperglycemia condition in db/db mice. Zeaxanthin and/or lutein were simultaneously added into the culture medium. Mannitol was used as an osmotic control. The whole-cell lysates were subjected to Western blot (a). β -Actin was used as an equal loading control to normalize each protein expression. Relative band intensity of each protein to β -actin was used for graphing and statistic analysis. (* $P < 0.05$, $n = 3$, mean data on AMPK and PERK shown in lower graphs). Other mean data not shown.) Zeaxanthin group samples #1 to #5: PBS, mannitol, glucose only, glucose with zeaxanthin 0.625 μ mol/L and glucose with zeaxanthin 1.25 μ mol/L, * $P < 0.05$ versus the PBS (sample #1). Lutein group samples #6 to #10: PBS, mannitol, glucose only, glucose with lutein 0.625 μ mol/L and glucose with lutein 1.25 μ mol/L, * $P < 0.05$ versus the PBS (sample #6). The cellular reactive oxygen species (ROS) level was determined (b). AMPK α was knocked down by its specific siRNA in the zeaxanthin (1.25 μ mol/L) treated ARPE-19 cells under a high glucose challenge. AMPK α siRNA knockdown-altered cellular ROS level was determined and expressed in the far right three black columns of graph b ($P < 0.05$ versus the Vehicle). PBS, phosphate-buffered saline; Mann, mannitol; Gluc, glucose; Zeax, zeaxanthin; AMPK α , AMP activated protein kinase α ; Mn SOD, Mn superoxide dismutase; PERK, protein kinase RNA-like ER kinase

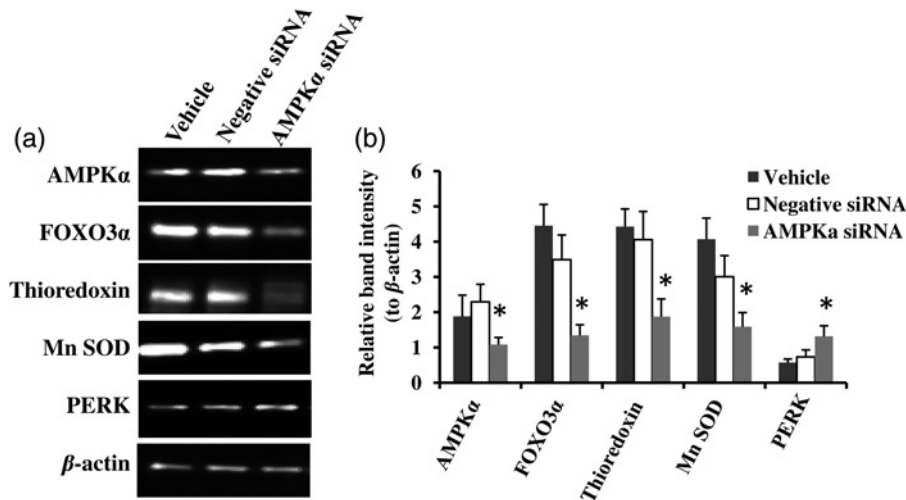


Figure 6 Small interfering RNA (siRNA) knockdown of AMP-activated protein kinase α (AMPK α) causes significant decreases in protein expression of FOXO3 α , Mn SOD and thioredoxin, but increases in PERK in high glucose challenged ARPE-19 cells. AMPK α was knocked down by its specific siRNA in the zeaxanthin (1.25 μ M/L) treated ARPE-19 cells under a high glucose challenge. Alteration of protein expression was monitored by Western blot. Representative images were shown (a). β -Actin was used as an equal loading control to normalize each protein expression. Relative band intensity of each protein to β -actin was used for graphing and statistic analysis (b). * $P \leq 0.05$ versus the Vehicle groups. PERK, protein kinase RNA-like ER kinase; Mn SOD, Mn superoxide dismutase

other bioactive components in wolfberry, such as phenolics and polysaccharides.

AMPK is a cellular energy sensor in mammals. It is down-regulated in type 2 diabetic patients.^{3–5} Targeting AMPK has been suggested as a most successful pharmaceutical strategy for type 2 diabetes. For instance, metformin, an

oral antidiabetic drug, at least partially works through activation of AMPK.^{6,48} Wolfberry induced AMPK activation in WT mice and increased AMPK protein concentration in db/db mice (Figures 4a–c). Retinal AMPK activation may be due to retinal specific accumulation of zeaxanthin and/or lutein. Further, wolfberry restored FOXO3 α (Figures 4f and g). Data suggested that wolfberry protection of retina from caspase-3 dependent apoptosis may be through regulation of protein expression, centered by AMPK activation.

FOXO3 α is regulated by AKT/PKB, extracellular signal-regulated kinases and AMPK. It is essential to cell survival in diabetes.^{31,49,50} FOXO3 α is being clinically shown to achieve the therapeutic effects.^{30,31} Down-regulation of FOXO3 α results in disruption of cellular ubiquitination and inhibition of antioxidant activity. AMPK up-regulation of FOXO3 α has been suggested to account for increased expression of antioxidant enzymes thioredoxin and Mn SOD in diabetic endothelial cells.⁵⁰ Application of wolfberry caused up-regulation of retinal FOXO3 α and thioredoxin and Mn SOD (Figure 4), which may lead to diminishing cellular ROS generation. Increased thioredoxin and Mn SOD by dietary wolfberry may be due to up-regulation of gene expression and/or stabilization of existing proteins by zeaxanthin and/or lutein under hyperglycemia. The cell culture study further suggested that wolfberry restoration of thioredoxin and Mn SOD would help re-balance cellular redox homeostasis through normalization of cellular ROS status.

ER stress has been exclusively documented to be a major driven force to cellular oxidative retinal damage at the late stage of diabetes.^{10,51} We have demonstrated in this study that hyperglycemia induced elevated protein expression of ER stress biomarkers in early diabetic retina *in vivo*, indicating that hyperglycemia accounts for retinal ER stress, inhibition of AMPK α , thioredoxin, Mn SOD, FOXO3 α , and retinal abnormality. This was further confirmed by *in vitro* studies on human retinal ARPE-19 cells when they were exposed to a high glucose challenge (Figure 5a). This was

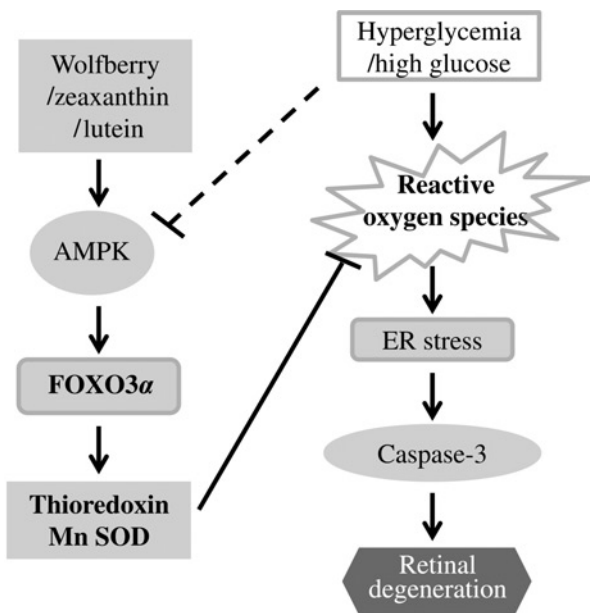


Figure 7 Schematic diagram showing wolfberry and/or zeaxanthin and lutein modulated retinal protection in type 2 diabetes. Dietary wolfberry restores AMP-activated protein kinase (AMPK), at least partially, through zeaxanthin and/or lutein. Restoration of AMPK and FOXO3 α signaling leads to enhanced expression of thioredoxin and Mn SOD which in turn results in normalization of cellular reactive oxygen species and attenuation of endoplasmic reticulum stress. This could lead to prevention of retinal apoptosis and restoration of retinal structure in the type 2 diabetic mouse. Dash line represents multiple steps. Mn SOD, Mn superoxide dismutase; FOXO3 α , forkhead O transcription factor 3 α

not due to osmotic pressure as mannitol alone did not differ from PBS controls. Prolonged hyperglycemia (or high glucose) distinctly induces generation of cellular ROS (Figure 5b), which was diminished by administration of whole wolfberries and/or the bioactive fractions zeaxanthin and lutein. AMPK α siRNA knockdown results further suggested that zeaxanthin diminished high glucose-induced cellular ROS levels via restoration of AMPK α . AMPK α is the upstream modulator of FOXO3 α and thioredoxin and Mn SOD.

Taken together, dietary wolfberry and/or its bioactive constituents, zeaxanthin and lutein, functioned as modulators of cell survival/death signaling pathways, through targeting pathways in AMPK and FOXO3 α signaling, resulting in normalization of cellular ROS and subsequent attenuation of ER stress. This could lead to prevention of retinal apoptosis and restoration of retinal structure in the type 2 diabetic mouse (Figure 7). Dietary treatments using individual fractions isolated from the wolfberry would be necessary for further mechanistic studies *in vivo*. To our knowledge, this is the first report that wolfberry bioactive constituents prevented or delayed the onset of the disease of diabetic retinopathy in an animal mode.

Author contributions: LT, YZ, YJ, LW, EO and LW conducted research; DM and DL designed research; LT, YZ, YJ, LW, DM and DL analyzed data; DM and DL wrote the manuscript; DL had primary responsibility for final content. All authors read and approved the final manuscript. LT, YZ and YJ contributed equally to this study.

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REFERENCES

- Busik JV, Mohr S, Grant MB. Hyperglycemia-induced reactive oxygen species toxicity to endothelial cells is dependent on paracrine mediators. *Diabetes* 2008;57:1952–65
- Nathan DM, Buse JB, Davidson MB, Heine RJ, Holman RR, Sherwin R, Zinman B. Management of hyperglycemia in type 2 diabetes: a consensus algorithm for the initiation and adjustment of therapy: a consensus statement from the American Diabetes Association and the European Association for the Study of Diabetes. *Diabetes Care* 2006;29:1963–72
- Sanz P. AMP-activated protein kinase: structure and regulation. *Curr Protein Pept Sci* 2008;9:478–92
- Zhang BB, Zhou G, Li C. AMPK: an emerging drug target for diabetes and the metabolic syndrome. *Cell Metab* 2009;9:407–16
- Kukidome D, Nishikawa T, Sonoda K, Imoto K, Fujisawa K, Yano M, Motoshima H, Taguchi T, Matsumura T, Araki E. Activation of AMP-activated protein kinase reduces hyperglycemia-induced mitochondrial reactive oxygen species production and promotes mitochondrial biogenesis in human umbilical vein endothelial cells. *Diabetes* 2006;55:120–7
- Leverve XM, Guigas B, Demaille D, Batandier C, Koceir EA, Chauvin C, Fontaine E, Wiernsperger NF. Mitochondrial metabolism and type-2 diabetes: a specific target of metformin. *Diabetes Metab* 2003;29:S88–94
- Rutter GA, Leclerc I. The AMP-regulated kinase family: enigmatic targets for diabetes therapy. *Mol Cell Endocrinol* 2009;297:41–9
- Kim I, Xu W, Reed JC. Cell death and endoplasmic reticulum stress: disease relevance and therapeutic opportunities. *Nat Rev Drug Discov* 2008;7:1013–30
- Malhotra JD, Kaufman RJ. Endoplasmic reticulum stress and oxidative stress: a vicious cycle or a double-edged sword? *Antioxid Redox Signal* 2007;9:2277–93
- Oshitari T, Hata N, Yamamoto S. Endoplasmic reticulum stress and diabetic retinopathy. *Vasc Health Risk Manag* 2008;4:115–22
- Potterat O, Goji (Lycium barbarum and L. chinense): phytochemistry, pharmacology and safety in the perspective of traditional uses and recent popularity. *Planta Med* 2010;76:7–19
- Ron D, Walter P. Signal integration in the endoplasmic reticulum unfolded protein response. *Nat Rev Mol Cell Biol* 2007;8:519–29
- Görlach A, Klappa P, Kietzmann T. The endoplasmic reticulum: folding, calcium homeostasis, signaling, and redox control. *Antioxid Redox Signal* 2006;8:1391–418
- Faitova J, Krekac D, Hrstka R, Vojtesek B. Endoplasmic reticulum stress and apoptosis. *Cell Mol Biol Lett* 2006;11:488–505
- Inbaraj BS, Lu H, Hung CF, Wu WB, Lin CL, Chen BH. Determination of carotenoids and their esters in fruits of *Lycium barbarum* Linnaeus by HPLC-DAD-APCI-MS. *J Pharm Biomed Anal* 2008;47:812–8
- Inbaraj BS, Lu H, Kao TH, Chen BH. Simultaneous determination of phenolic acids and flavonoids in *Lycium barbarum* Linnaeus by HPLC-DAD-ESI-MS. *J Pharm Biomed Anal* 2010;51:549–56
- Chang RC, So KF. Use of anti-aging herbal medicine, *Lycium barbarum*, against aging-associated diseases. What do we know so far? *Cell Mol Neurobiol* 2008;28:643–52
- Ho YS, So KF, Chang RC. Anti-aging herbal medicine – how and why can they be used in aging-associated neurodegenerative diseases? *Ageing Res Rev* 2010;9:354–62
- Yemm EW, Willis AJ. The estimation of carbohydrates in plant extracts by anthrone. *Biochem J* 1954;57:508–14
- Peri C, Pompei C. An assay of different phenolic fractions in wines. *Am J Enol Vitic* 1971;22:55–8
- Lin D, Takemoto DJ. Oxidative activation of protein kinase C γ through the C1 domain. Effects on gap junctions. *J Biol Chem* 2005;280:13682–93
- Yevseyenkov VV, Das S, Lin D, Willard L, Davidson H, Sitaramayya A, Giblin FJ, Dang L, Takemoto DJ. Loss of protein kinase C γ in knockout mice and increased retinal sensitivity to hyperbaric oxygen. *Arch Ophthalmol* 2009;127:500–6
- Arnal E, Miranda M, Johnsen-Nörling R, Díaz-Llopis M, Araiz J, Cervera E, Bosch-Morell F, Romero FJ. Beneficial effect of docosahexanoic acid and lutein on retinal structural, metabolic, and functional abnormalities in diabetic rats. *Curr Eye Res* 2009;34:928–38
- Béby F, Housset M, Fossat N, Le Greneur C, Flamant F, Godement P, Lamonerie T. Otx2 gene deletion in adult mouse retina induces rapid RPE dystrophy and slow photoreceptor degeneration. *PLoS One* 2010;5:e11673
- Jones DP. Redox sensing: orthogonal control in cell cycle and apoptosis signalling. *J Intern Med* 2010;268:432–48
- Nishikawa T, Araki E. Impact of mitochondrial ROS production in the pathogenesis of diabetes mellitus and its complications. *Antioxid Redox Signal* 2007;9:343–53
- Kim A, Joseph S, Khan A, Epstein CJ, Sobel R, Huang TT. Enhanced expression of mitochondrial superoxide dismutase leads to prolonged *in vivo* cell cycle progression and up-regulation of mitochondrial thioredoxin. *Free Radic Biol Med* 2010;48:1501–12
- Tremblay ML, Giguère V. Phosphatases at the heart of FoxO metabolic control. *Cell Metab* 2008;7:101–3
- Maiese K, Chong ZZ, Hou J, Shang YC. The 'O' class: crafting clinical care with FoxO transcription factors. *Adv Exp Med Biol* 2009;665:242–60
- Kops GJ, Dansen TB, Polderman PE, Saarloos I, Wirtz KW, Coffey PJ, Huang TT, Bos JL, Medema RH, Burgering BM. Forkhead transcription factor FOXO3a protects quiescent cells from oxidative stress. *Nature* 2002;419:316–21
- Li XN, Song J, Zhang L, LeMaire SA, Hou X, Zhang C, Coselli JS, Chen L, Wang XL, Zhang Y, Shen YH. Activation of the AMPK–FOXO3 pathway

- reduces fatty acid-induced increase in intracellular reactive oxygen species by upregulating thioredoxin. *Diabetes* 2009;**58**:2246–57
- 32 Li B, Vachali P, Bernstein PS. Human ocular carotenoid-binding proteins. *Photochem Photobiol Sci* 2010;**9**:1418–25
 - 33 During A, Doraiswamy S, Harrison EH. Xanthophylls are preferentially taken up compared with beta-carotene by retinal cells via a SRBI-dependent mechanism. *J Lipid Res* 2008;**49**:1715–24
 - 34 Midena E, Segato T, Radin S, di Giorgio G, Meneghini F, Piermarocchi S, Belloni AS. Studies on the retina of the diabetic db/db mouse. I. Endothelial cell–pericyte ratio. *Ophthalmic Res* 1989;**21**:106–11
 - 35 Tadayoni R, Paques M, Gaudric A, Vicaute E. Erythrocyte and leukocyte dynamics in the retinal capillaries of diabetic mice. *Exp Eye Res* 2003;**77**:497–504
 - 36 Clements RS Jr, Robison WG Jr, Cohen MP. Anti-glycated albumin therapy ameliorates early retinal microvascular pathology in db/db mice. *J Diabetes Complications* 1998;**12**:28–33
 - 37 Barile GR, Pachydaki SI, Tari SR, Lee SE, Donmoyer CM, Ma W, Rong LL, Buciarelli LG, Wendt T, Horig H, Hudson BI, Qu W, Weinberg AD, Yan SF, Schmidt AM. The RAGE axis in early diabetic retinopathy. *Invest Ophthalmol Vis Sci* 2005;**46**:2916–24
 - 38 Cheung AK, Fung MK, Lo AC, Lam TT, So KF, Chung SS, Chung SK. Aldose reductase deficiency prevents diabetes-induced blood–retinal barrier breakdown, apoptosis, and glial reactivation in the retina of db/db mice. *Diabetes* 2005;**54**:3119–25
 - 39 Li J, Wang JJ, Chen D, Mott R, Yu Q, Ma JX, Zhang SX. Systemic administration of HMG-CoA reductase inhibitor protects the blood–retinal barrier and ameliorates retinal inflammation in type 2 diabetes. *Exp Eye Res* 2009;**89**:71–8
 - 40 Li J, Wang JJ, Yu Q, Chen K, Mahadev K, Zhang SX. Inhibition of reactive oxygen species by Lovastatin downregulates vascular endothelial growth factor expression and ameliorates blood–retinal barrier breakdown in db/db mice: role of NADPH oxidase 4. *Diabetes* 2010;**59**:1528–38
 - 41 Sohn EJ, Kim YS, Kim CS, Lee YM, Kim JS. KIOM-79 prevents apoptotic cell death and AGEs accumulation in retinas of diabetic db/db mice. *J Ethnopharmacol* 2009;**121**:171–4
 - 42 Yeung CM, Lo AC, Cheung AK, Chung SS, Wong D, Chung SK. More severe type 2 diabetes-associated ischemic stroke injury is alleviated in aldose reductase-deficient mice. *J Neurosci Res* 2010;**88**:2026–34
 - 43 Sasaki M, Ozawa Y, Kurihara T, Kubota S, Yuki K, Noda K, Kobayashi S, Ishida S, Tsubota K. Neurodegenerative influence of oxidative stress in the retina of a murine model of diabetes. *Diabetologia* 2010;**53**:971–9
 - 44 Bernstein PS, Balashov NA, Tsong ED, Rando RR. Retinal tubulin binds macular carotenoids. *Invest Ophthalmol Vis Sci* 1997;**38**:167–75
 - 45 Crabtree DV, Ojima I, Geng X, Adler AJ. Tubulins in the primate retina: evidence that xanthophylls may be endogenous ligands for the paclitaxel-binding site. *Bioorg Med Chem* 2001;**9**:1967–76
 - 46 Bhosale AJL, Frederick J, Southwick K, Thulin CD, Bernstein PS. Identification and characterization of a pi isoform of glutathione S-transferase as a zeaxanthin-binding protein in the macula of the human eye. *J Biol Chem* 2004;**279**:49447–54
 - 47 Kowluru RA, Menon B, Gierhart DL. Beneficial effect of zeaxanthin on retinal metabolic abnormalities in diabetic rats. *Invest Ophthalmol Vis Sci* 2008;**49**:1645–51
 - 48 Hou X, Song J, Li XN, Zhang L, Wang X, Chen L, Shen YH. Metformin reduces intracellular reactive oxygen species levels by upregulating expression of the antioxidant thioredoxin via the AMPK–FOXO3 pathway. *Biochem Biophys Res Commun* 2010;**396**:199–205
 - 49 Katoh M, Katoh M. Human FOX gene family (Review). *Int J Oncol* 2004;**25**:1495–500
 - 50 Kukidome D, Nishikawa T, Sonoda K, Imoto K, Fujisawa K, Yano M, Motoshima H, Taguchi T, Matsumura T, Araki E. Activation of AMP-activated protein kinase reduces hyperglycemia-induced mitochondrial reactive oxygen species production and promotes mitochondrial biogenesis in human umbilical vein endothelial cells. *Diabetes* 2006;**55**:120–7
 - 51 Li J, Wang JJ, Yu Q, Wang M, Zhang SX. Endoplasmic reticulum stress is implicated in retinal inflammation and diabetic retinopathy. *FEBS Lett* 2009;**583**:1521–7

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