

Resveratrol promotes neurotrophic factor release from astroglia

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Abstract

Neurotrophic factors such as glial cell line-derived neurotrophic factor (GDNF) and brain-derived neurotrophic factor (BDNF) are considered to contribute to the development, maintenance and survival of neurons, glia and oligodendrocytes. Astroglia are a major source of various neurotrophic factors. Thus, enhancement of astroglia-mediated neurotrophic factor release might hold promising potential for neurological diseases. Resveratrol, a natural non-flavonoid polyphenol found in grapes and red wine, has been recognized to be beneficial for health. Here, rat primary astroglia-enriched cultures were used to investigate the effects of resveratrol-mediated neurotrophic factor release and the related mechanisms. The cultures were treated with 25–100 μ mol/L resveratrol for 12–48 h. Results showed resveratrol increased BDNF and GDNF production in the culture medium. In addition, the production of BDNF in the supernatant of cultures was increased five-fold over control cultures 24 h after resveratrol treatment and then remained high 36 h later. Meanwhile, the production of GDNF was initially increased by up to four-fold 24 h after resveratrol treatment and continued to increase to six-fold at 36 h and remained at a high level till 48 h. Western blot analysis of BDNF and GDNF protein in astroglia at different time points after resveratrol treatment indicated similar increases. Furthermore, resveratrol significantly induced the phosphorylation of extracellular signal-regulated kinase 1/2 (ERK1/2) and cAMP responsive element-binding protein (CREB) in astroglia. Overall, resveratrol is effective in promoting astroglia-derived neurotrophic factor release, and this effect is mediated, at least in part, by the activation of ERK1/2 and CREB.

Keywords: resveratrol, neurological disorders, neurotrophic factors, BDNF, GDNF

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Introduction

Recent advances have demonstrated that glia (particularly astroglia and microglia) play an indispensable role in neurological disorders and have become the prime targets for therapy.^{1,2} Astroglia serve many housekeeping functions, such as the maintenance of the extracellular environment and the stabilization of cell-cell communications in the central nervous system (CNS).³ It is becoming increasingly evident that astroglia are a major source of various neurotrophic factors, such as glial cell line-derived neurotrophic factor (GDNF) and brain-derived neurotrophic factor (BDNF).⁴ Furthermore, neurotrophic factors are considered to contribute to the development, maintenance and survival of neurons, glia and oligodendrocytes.⁵ In contrast, lack of neurotrophic factors might lead to neuronal death and the pathogenesis of neurodegenerative diseases.^{6,7} Thus, enhancement of astroglia-mediated neurotrophic factor release might hold promising potential for the treatment of neurological diseases.

Resveratrol (3, 4', 5-trihydroxy-trans-stilbene), a natural non-flavonoid polyphenol found in grapes and red wine, has been recognized to be beneficial for health.⁸ It is known to possess a large number of pharmacological properties such as cardioprotective, anti-inflammatory, antioxidant and anti-cancer effects.⁹ Besides these beneficial actions, increasing interest has focused on its neuroprotective activities. Resveratrol could cross the blood-brain barrier¹⁰ and produce neuroprotection against ischemia, seizure and neurodegenerative diseases.¹¹ Recent studies indicated that resveratrol improved hippocampal atrophy through enhancing neurogenesis and inhibiting apoptosis of granular cells in chronic fatigue mice.¹² However, the mechanisms underlying these effects still remain unclear. Our previous studies demonstrated that resveratrol protected dopaminergic neurons from lipopolysaccharide (LPS)-induced neurodegeneration via its anti-inflammatory properties.¹³ To extend our understanding of resveratrol mediated neuroprotection, rat primary astroglia cultures

were applied to investigate the effects of resveratrol on the production of neurotrophic factors from astroglia and further elucidate the related molecular mechanisms.

Materials and methods

Animals

Female Wistar rats (200–300 g) were purchased from the Experimental Animal Centre of the Third Military Medical University (Chongqing, China; Specific-pathogen free Grade II; Certificate No. scxk 2002003). Housing and breeding of the animals were performed in strict accordance with Animal Care and Use Guidelines in China.

Reagents

Resveratrol was obtained from Sigma Chemical Co. (St Louis, MO, USA). All the materials of cell cultures were purchased from Invitrogen (Carlsbad, CA, USA). Primary antibodies were obtained from Cell Signaling Technology (Beverly, MA, USA). The AntiVectastain avidin-biotin complex kit was obtained from Vector Laboratories (Burlingame, CA, USA). Enzyme-linked immunosorbent assay (ELISA) kits were obtained from Chemicon International Inc. (Temecula, CA, USA).

Rat primary astroglia-enriched cultures

Primary astroglia-enriched cultures were prepared from the whole brains of one-day-old rat pups as described previously.¹⁴ After a confluent monolayer of glial cells had been established, microglia were separated from astroglia by shaking the flasks at 180 rpm for 1 h. The remaining astroglia were detached with trypsin-ethylenediaminetetraacetic acid (EDTA) and seeded in the culture medium. After two or three consecutive passages, immunocytochemical analysis indicated that the cultures were composed of >98% astroglia.

Measurement of BDNF and GDNF by ELISA

The production of BDNF and GDNF in the culture supernatant was quantified using ELISA kits according to the procedures provided by the manufacturer.

Western blot analysis

The whole cells from primary astroglia-enriched cultures were washed with cold phosphate-buffered saline and lysed with radioimmunoprecipitation assay (RIPA) cell lysis buffer consisting of 50 mmol/L Tris-HCl (pH 7.4), 150 mmol/L NaCl, 1 mmol/L EDTA, 5 μ g/mL aprotinin, 1 mmol/L phenylmethylsulfonyl fluoride, 5 μ g/mL leupeptin, 1% Triton X-100, 1% sodium deoxycholate and 1 mmol/L Na₃VO₄.¹³ The lysates were incubated on ice for 30 min and then centrifuged at 12,000 $\times$ g for 25 min. Protein levels from the whole cell were quantified using a bicinchoninic acid assay. Equal amounts of total protein (30–50 μ g per lane) were separated on 4–12% Bis-Tris Nu-polyacrylamide gel

electrophoresis gel and transferred onto polyvinylidene difluoride membranes. Membranes were blocked with 5% non-fat milk and then incubated with the following primary antibodies: anti-BDNF, anti-GDNF, anti-phospho-cAMP responsive element-binding (CREB), anti-CREB, anti-phospho-ERK1/2 (p-ERK1/2), anti-ERK1/2 and anti- β -actin at 1:1000 dilution and horseradish peroxidase (HRP)-conjugated secondary antibodies at 1:2500 dilution. The blots were developed with enhanced chemiluminescence reagent.

Statistical analysis

Data are expressed as mean $\pm$ SEM from three independent experiments performed in triplicate. Statistical significance was assessed by analysis of variance (ANOVA) using GraphPad Prism software (GraphPad Software Inc., San Diego, CA, USA). When ANOVA showed significant differences, pairwise comparisons between means were tested by Bonferroni's post-test with correction. A value of $P < 0.05$ was considered statistically significant.

Results

Resveratrol increased BDNF and GDNF production in the culture medium

Rat primary astroglia-enriched cultures were treated with resveratrol (25–100 μ mol/L). After resveratrol treatment for 24 h, the release of BDNF and GDNF in the supernatant was detected by ELISA. On the basis of ANOVA with Bonferroni's test analysis, resveratrol significantly increased BDNF (six-fold) and GDNF (four-fold) production in the culture medium as shown in Figure 1.

Resveratrol time-dependently induced BDNF and GDNF release

Rat primary astroglia-enriched cultures were treated with resveratrol (100 μ mol/L). Twelve, 24, 36 and 48 h after resveratrol treatment, the release of BDNF and GDNF in the supernatant was measured by ELISA. On the basis of ANOVA with Bonferroni's test analysis, the production of BDNF was increased by up to five-fold of control cultures 24 h after resveratrol treatment and then maintained at a four-fold level 36 h later (Figure 2a). The production of GDNF was initially increased by up to four-fold 24 h after resveratrol application and continued to increase to six-fold at 36 h and was still five-fold higher 48 h later (Figure 2b).

To further investigate the effects of resveratrol on BDNF and GDNF protein levels, total cell protein was harvested 12, 24, 36 and 48 h after resveratrol (100 μ mol/L) treatment, respectively. Western blot analysis was applied to detect the protein expressions. The densitometry values of BDNF, GDNF and β -actin were analyzed and normalized to each respective control group. On the basis of ANOVA with Bonferroni's test analysis, consistent with the BDNF and GDNF production in the culture medium, astroglia-derived BDNF protein levels increased by up to 170% of control

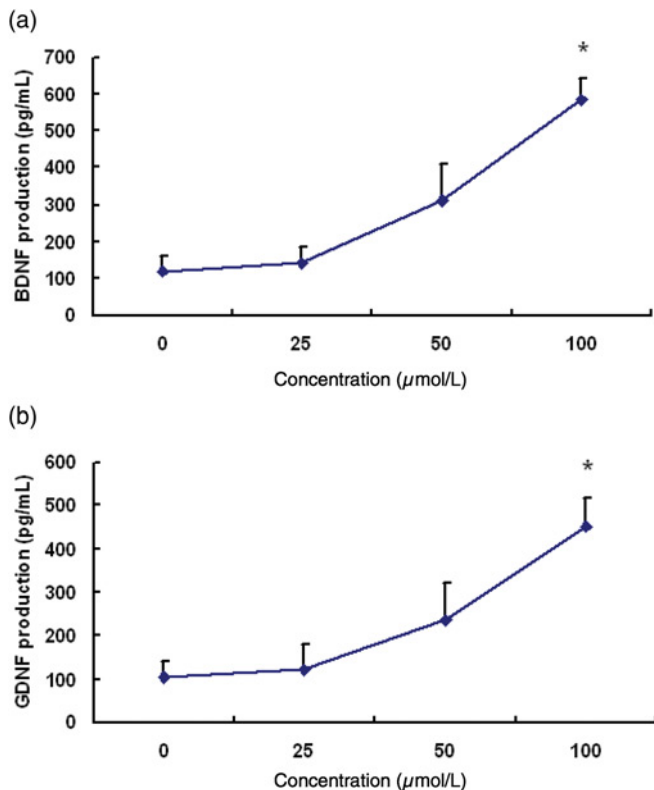


Figure 1 Resveratrol increased brain-derived neurotrophic factor (BDNF) and glial cell line-derived neurotrophic factor (GDNF) production in the culture medium. The release of BDNF (a) and GDNF (b) in the culture medium 24 h after treatment with resveratrol was detected by enzyme-linked immunosorbent assay. Data are expressed as a percentage of the control cultures and are the mean $\pm$ SEM from three independent experiments performed in triplicate. * $P < 0.05$ compared with the control cultures (analysis of variance with Bonferroni's test). (A color version of this figure is available in the online journal)

culture levels 24 h after resveratrol treatment and remained at 140% of control culture levels 36 h later (Figure 3a). The GDNF protein expression was increased by up to 140% of control cultures after resveratrol treatment for 24 h and peaked to 160% at 36 h and maintained at 130% of control cultures 48 h later (Figure 3b).

Induction of CREB and ERK1/2 activation by resveratrol

Rat primary astroglia-enriched cultures were treated with resveratrol (25–100 μmol) for 60 min. The whole-cell lysates were analyzed by Western blotting assay. The ratio of densitometry values of phosphorylated CREB and ERK1/2 (p-ERK1/2) compared with total CREB and ERK1/2 was analyzed and normalized to each respective control group. On the basis of ANOVA with Bonferroni's test analysis, resveratrol induced the phosphorylation of CREB and ERK1/2 by up to 150% and 180% of control cultures, respectively, as shown in Figure 4.

Discussion

This study indicated that resveratrol promoted the release of BDNF and GDNF neurotrophic factors from astroglia,

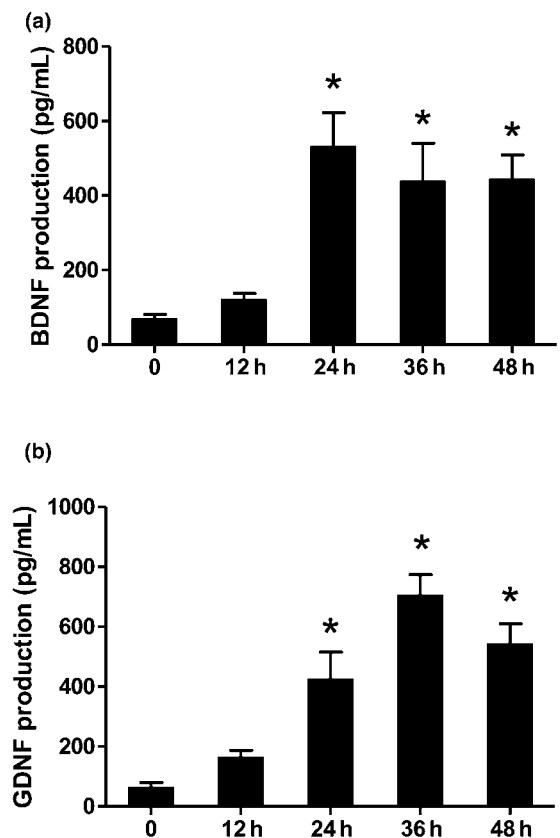


Figure 2 Resveratrol time-dependently induced brain-derived neurotrophic factor (BDNF) and glial cell line-derived neurotrophic factor (GDNF) release. The release of BDNF (a) and GDNF (b) in the supernatant after treatment of 100 μmol/L resveratrol was measured by enzyme-linked immunosorbent assay. Data are expressed as a percentage of the control cultures and are the mean $\pm$ SEM from three independent experiments performed in triplicate. * $P < 0.05$ compared with the control cultures (analysis of variance with Bonferroni's test)

as evidenced by ELISA of the culture medium and by Western blot analysis of astroglia protein. Astroglia and microglia are becoming the targets of neuroprotection.^{1,7,9} Our previous studies have shown that microglia are involved in protection against LPS-induced neuroinflammation and dopaminergic neuronal damage by resveratrol^{9,13} and other neuroprotective compounds.^{15,16} Here, we further demonstrate that the neuroprotective effects of resveratrol also involve the activation of astroglia, with increased production of neurotrophic factors, probably mediated through the activation of the CREB and ERK1/2 signaling pathways.

A growing body of evidence indicates that astroglia play a critical role in the regulation of neurogenesis, synaptogenesis and neurotransmitter environments of the CNS.¹⁷ By releasing various neurotrophic factors, astroglia also provide neuroprotection against neuronal damage.¹⁸ BDNF is up-regulated in astroglia in the vicinity of injury sites in humans and experimental animals.¹⁹ Upon activation, astroglia release BDNF to rescue neuronal cells from neuritic degeneration and neuronal death.^{20,21} Several lines of evidence indicate that BDNF has beneficial effects on cognition, learning and memory formation by modulating synaptic plasticity.²² Studies also show that

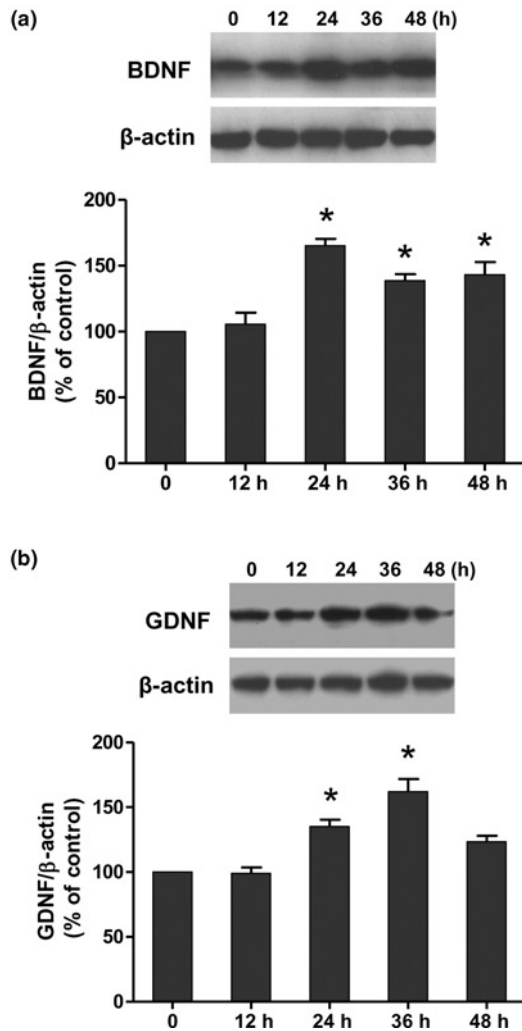


Figure 3 Resveratrol increased brain-derived neurotrophic factor (BDNF) and glial cell line-derived neurotrophic factor (GDNF) protein in astroglia cultures. The levels of BDNF (a) and GDNF (b) protein were determined by Western blot analysis after 100 $\mu\text{mol/L}$ resveratrol treatment for different times. The densitometry values of target proteins (BDNF or GDNF) were normalized to β -actin of the same sample. Data are expressed as a percentage of the control cultures and are the mean $\pm$ SEM from three independent experiments performed in triplicate. * $P < 0.05$ compared with the control cultures (analysis of variance with Bonferroni's test)

the decreased level of BDNF was discerned in neurodegenerative diseases such as Alzheimer's disease, Huntington's disease, Parkinson's disease, depression, epilepsy and chronic pain.²³ In addition, GDNF has been recognized to play a crucial role in maintaining the physiological functions of the brain and pathological features of the CNS.²⁴ GDNF produced neuroprotection against ischemia, drug abuse and neuropathic pain.²⁵ Moreover, astroglia were reported to protect neurons through secreting various neurotrophic factors such as GDNF.²⁶ Besides BDNF, increasing evidence demonstrated that the alterations of the GDNF levels were detected in the peripheral blood and postmortem brain tissue of patients with mood disorders.^{27,28} Therefore, promotion of BDNF and GDNF release is expected to have therapeutic potential for neurological disorders. This study is among the first to reveal that resveratrol significantly induced BDNF and GDNF production in astroglia cultures.

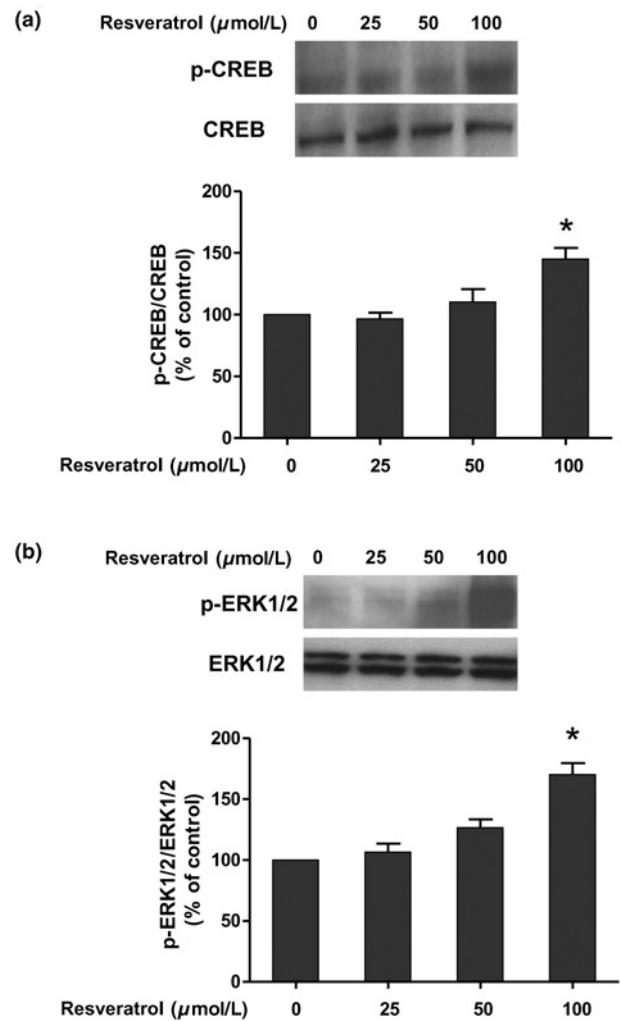


Figure 4 Resveratrol induction of cAMP responsive element-binding protein (CREB) and extracellular signal-regulated kinase 1/2 (ERK1/2) in astroglia cultures. The CREB (a) and ERK1/2 (b) protein expressions were detected by Western blot assay 60 min after 100 $\mu\text{mol/L}$ resveratrol treatment at different concentrations. Data are expressed as a percentage of the control cultures and are the mean $\pm$ SEM from three independent experiments performed in triplicate. * $P < 0.05$ compared with the control cultures (analysis of variance with Bonferroni's test)

This finding was consistent with a previous study in which resveratrol increased BDNF gene expression in the hippocampus of rat brain,^{19,29} and extended this finding to astroglia, an important brain cell type in neuroprotection.

As rapid activation of mitogen-activated protein kinase (MEK)/ERK/CREB pathways is critically involved in neurotrophic factor production in C6 glioma cells,^{30,31} we next examined the role of ERK1/2 and CREB-signaling pathways in resveratrol-mediated BDNF and GDNF release. It has been reported that the ERK1/2 signaling pathway participates in the phosphorylation of CREB in cultured astrocytes.³² Activation of CREB is observed to promote GDNF production in cultured astrocyte cells through CREB located in the promoter sequence of the GDNF.^{33,34} In addition, CREB siRNA can block the GDNF expression in astrocytes.³⁵ Therefore, the CREB signaling pathway represents a potential target responsible for inducing neurotrophic factor release.

Data from earlier clinical experiments indicate that GDNF was tested for its potential therapeutic treatment of Parkinson's disease by direct infusion into the brain of patients. However, this trial was terminated due to the occurrence of severe side-effects.^{35–38} Thus, this study might provide an alternative approach for the treatment of neurological diseases by application of natural substances such as resveratrol to promote neurotrophic factors release.

Overall, this study clearly demonstrates that resveratrol is effective in promoting the release of neurotrophic factors BDNF and GDNF in rat astroglia cultures and that the ERK1/2 and CREB pathways might participate in resveratrol-induced neurotrophic effects.

Author contributions: All authors participated in the design, interpretation of the studies, analysis of the data, review of the manuscript and conducted the experiments. FZ wrote the manuscript.

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REFERENCES

- Block ML, Zecca L, Hong JS. Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nat Rev Neurosci* 2007;**8**:57–69
- Ralay Ranaivo H, Craft JM, Hu W, Guo L, Wing LK, Van Eldik LJ, Watterson DM. Glia as a therapeutic target: selective suppression of human amyloid-beta-induced upregulation of brain proinflammatory cytokine production attenuates neurodegeneration. *J Neurosci* 2006;**26**:662–70
- Maragakis NJ, Rothstein JD. Mechanisms of disease: astrocytes in neurodegenerative disease. *Nat Clin Pract Neurol* 2006;**2**:679–89
- Chen PS, Peng GS, Li G, Yang S, Wu X, Wang CC, Wilson B, Lu RB, Gean PW, Chuang DM, Hong JS. Valproate protects dopaminergic neurons in midbrain neuron/glia cultures by stimulating the release of neurotrophic factors from astrocytes. *Mol Psychiatry* 2006;**11**:1116–25
- Huang Y, Cheung L, Rowe D, Halliday G. Genetic contributions to Parkinson's disease. *Brain Res Brain Res Rev* 2004;**46**:44–70
- Phillips HS, Hains JM, Armanini M, Laramie GR, Johnson SA, Winslow JW. BDNF mRNA is decreased in the hippocampus of individuals with Alzheimer's disease. *Neuron* 1991;**7**:695–702
- Schindowski K, Belarbi K, Buee L. Neurotrophic factors in Alzheimer's disease: role of axonal transport. *Genes Brain Behav* 2008;**7**(Suppl 1):43–56
- Saiko P, Szakmary A, Jaeger W, Szekeres T. Resveratrol and its analogs: defense against cancer, coronary disease and neurodegenerative maladies or just a fad? *Mutat Res* 2008;**658**:68–94
- Zhang F, Liu J, Shi JS. Anti-inflammatory activities of resveratrol in the brain: role of resveratrol in microglial activation. *Eur J Pharmacol* 2010;**636**:1–7
- Wang Q, Xu J, Rottinghaus GE, Simonyi A, Lubahn D, Sun GY, Sun AY. Resveratrol protects against global cerebral ischemic injury in gerbils. *Brain Res* 2002;**958**:439–47
- Markus MA, Morris BJ. Resveratrol in prevention and treatment of common clinical conditions of aging. *Clin Interv Aging* 2008;**3**:331–9
- Moriya J, Chen R, Yamakawa J, Sasaki K, Ishigaki Y, Takahashi T. Resveratrol improves hippocampal atrophy in chronic fatigue mice by enhancing neurogenesis and inhibiting apoptosis of granular cells. *Biol Pharm Bull* 2011;**34**:354–9
- Zhang F, Shi JS, Zhou H, Wilson B, Hong JS, Gao HM. Resveratrol protects dopamine neurons against LPS-induced neurotoxicity through its anti-inflammatory actions. *Mol Pharmacol* 2010;**78**:466–77
- Zhang W, Shin EJ, Wang T, Lee PH, Pang H, Wie MB, Kim WK, Kim SJ, Huang WH, Wang Y, Zhang W, Hong JS, Kim HC. 3-Hydroxymorphinan, a metabolite of dextromethorphan, protects nigrostriatal pathway against MPTP elicited damage both *in vivo* and *in vitro*. *FASEB J* 2006;**20**:2496–511
- Zhang F, Zhou H, Wilson BC, Shi JS, Hong JS, Gao HM. Fluoxetine protects neurons against microglial activation-mediated neurotoxicity. *Parkinsonism Relat Disord* 2012;**18**:S213–7
- Zhang F, Qian L, Flood PM, Shi JS, Hong JS, Gao HM. Inhibition of IkappaB kinase-beta protects dopamine neurons against lipopolysaccharide-induced neurotoxicity. *J Pharmacol Exp Ther* 2010;**333**:822–33
- Ullian EM, Christopherson KS, Barres BA. Role for glia in synaptogenesis. *Glia* 2004;**47**:209–16
- Darlington CL. Astrocytes as targets for neuroprotective drugs. *Curr Opin Investig Drug* 2005;**6**:700–3
- Wei R, Lin CM, Tu YY. Strain-specific BDNF expression of rat primary astrocytes. *J Neuroimmunol* 2010;**220**:90–8
- Mizuno T, Kuno R, Nitta A, Nabeshima T, Zhang G, Kawanokuchi J, Wang J, Jin S, Takeuchi H, Suzumura A. Protective effects of nicergoline against neuronal cell death induced by activated microglia and astrocytes. *Brain Res* 2005;**1066**:78–85
- Kimura N, Takahashi M, Tashiro T, Terao K. Amyloid beta up-regulates brain-derived neurotrophic factor production from astrocytes: rescue from amyloid beta-related neuritic degeneration. *J Neurosci Res* 2006;**84**:782–9
- Pardon MC. Role of neurotrophic factors in behavioral processes: implications for the treatment of psychiatric and neurodegenerative disorders. *Vitam Horm* 2010;**82**:185–200
- Pezet S, Malcangio M. Brain-derived neurotrophic factor as a drug target for CNS disorders. *Expert Opin Ther Targets* 2004;**8**:391–9
- Akerud P, Canals JM, Snyder EY, Arenas E. Neuroprotection through delivery of glial cell line-derived neurotrophic factor by neural stem cells in a mouse model of Parkinson's disease. *J Neurosci* 2001;**21**:8108–18
- Linker R, Gold R, Luhder F. Function of neurotrophic factors beyond the nervous system: inflammation and autoimmune demyelination. *Crit Rev Immunol* 2009;**29**:43–68
- Villegas SN, Poletta FA, Carri NG. GLIA: a reassessment based on novel data on the developing and mature central nervous system. *Cell Biol Int* 2003;**27**:599–609
- Michel TM, Frangou S, Camara S, Thiemeyer D, Jecel J, Tatschner T, Zochling R, Grünblatt E. Altered glial cell line-derived neurotrophic factor (GDNF) concentrations in the brain of patients with depressive disorder: a comparative post-mortem study. *Eur Psychiatry* 2008;**23**:413–20
- Takebayashi M, Hisaoka K, Nishida A, Tsuchioka M, Miyoshi I, Kozuru T, Hikasa S, Okamoto Y, Shinno H, Morinobu S, Yamawaki S. Decreased levels of whole blood glial cell line-derived neurotrophic factor (GDNF) in remitted patients with mood disorders. *Int J Neuropsychopharmacol* 2006;**9**:607–12
- Rahvar M, Nikseresh M, Shafiee SM, Naghibalhossaini F, Rasti M, Panjehshahin MR, Owji AA. Effect of oral resveratrol on the BDNF gene expression in the hippocampus of the rat brain. *Neurochem Res* 2011;**36**:761–5
- Hisaoka K, Takebayashi M, Tsuchioka M, Maeda N, Nakata Y, Yamawaki S. Antidepressants increase glial cell line-derived neurotrophic factor production through monoamine-independent activation of protein tyrosine kinase and extracellular signal-regulated kinase in glial cells. *J Pharmacol Exp Ther* 2007;**321**:148–57
- Tsuchioka M, Takebayashi M, Hisaoka K, Maeda N, Nakata Y. Serotonin (5-HT) induces glial cell line-derived neurotrophic factor (GDNF) mRNA expression via the transactivation of fibroblast growth factor receptor 2 (FGFR2) in rat C6 glioma cells. *J Neurochem* 2008;**106**:244–57
- Schinelli S, Zanassi P, Paolillo M, Wang H, Feliciello A, Gallo V. Stimulation of endothelin B receptors in astrocytes induces cAMP response element-binding protein phosphorylation and c-fos expression via multiple mitogen-activated protein kinase signaling pathways. *J Neurosci* 2001;**21**:8842–53

- 33 Koyama Y, Egawa H, Osakada M, Baba A, Matsuda T. Increase by FK960, a novel cognitive enhancer, in glial cell line-derived neurotrophic factor production in cultured rat astrocytes. *Biochem Pharmacol* 2004;**68**:275–82
- 34 Cen X, Nitta A, Ohya S, Zhao Y, Ozawa N, Mouri A, Ibi D, Wang L, Suzuki M, Saito K, Ito Y, Kawagoe T, Noda Y, Ito Y, Furukawa S, Nabeshima T. An analog of a dipeptide-like structure of FK506 increases glial cell line-derived neurotrophic factor expression through cAMP response element-binding protein activated by heat shock protein 90/Akt signaling pathway. *J Neurosci* 2006;**26**:3335–44
- 35 Yan M, Dai H, Ding T, Dai A, Zhang F, Yu L, Chen G, Chen Z. Effects of dexmedetomidine on the release of glial cell line-derived neurotrophic factor from rat astrocyte cells. *Neurochem Int* 2011;**58**:549–57
- 36 Kordower JH, Palfi S, Chen EY, Ma SY, Sendera T, Cochran EJ, Cochran EJ, Mufson EJ, Penn R, Goetz CG, Comella CD. Clinicopathological findings following intraventricular glial-derived neurotrophic factor treatment in a patient with Parkinson's disease. *Ann Neurol* 1999;**46**:419–24
- 37 Nutt JG, Burchiel KJ, Comella CL, Jankovic J, Lang AE, Laws ER Jr, Lozano AM, Penn RD, Simpson RK Jr, Stacy M, Wooten GF. Randomized, double-blind trial of glial cell linederived neurotrophic factor (GDNF) in PD. *Neurology* 2003;**60**:69–73
- 38 Peterson AL, Nutt JG. Treatment of Parkinson's disease with trophic factors. *Neurotherapeutics* 2008;**5**:270–80

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