

# Total Acid-Labile CO<sub>2</sub> Content in Dog Renal Cortex and Medulla<sup>1</sup> (34455)

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Information concerning the renal corticomedullary distribution of bicarbonate (HCO<sub>3</sub><sup>-</sup>) and CO<sub>2</sub> is of importance since either may affect intra- and extracellular pH (1), H<sup>+</sup> secretion rate by renal collecting duct cells (2), and in addition, certain metabolic processes (3) which may occur in the renal medulla.

We have attempted to determine whether there is a corticomedullary gradient for total CO<sub>2</sub> (CO<sub>2</sub> + HCO<sub>3</sub><sup>-</sup>) by measuring the acid-labile CO<sub>2</sub> (4) content of renal cortex and medulla. The acid-labile CO<sub>2</sub> content of tissue includes gaseous or dissolved CO<sub>2</sub>, HCO<sub>3</sub><sup>-</sup>, and carbamino-bound CO<sub>2</sub>. Thus, a corticomedullary gradient for any one of these moieties could account for any difference observed in the measured acid-labile CO<sub>2</sub> content between renal cortex and medulla. Due to the countercurrent arrangement of the tubules and blood vessels in the renal medulla, it seemed likely that trapping of the gaseous CO<sub>2</sub>, which is produced locally, could raise the medullary acid-labile CO<sub>2</sub> above that in the cortex. The following experiments were done to test this hypothesis.

**Methods.** Female mongrel dogs were anesthetized with sodium pentobarbital (30 mg/kg iv); additional doses of 10 mg/kg iv pentobarbital were given as needed. Duplicate 5-ml arterial (5) samples were obtained in oiled syringes for anaerobic pH determination using the Cambridge Research Model pH meter thermostated at 38°. Samples (5-ml) for determination of urine pH were ob-

tained through a self-retaining bladder catheter. The left kidney and its pedicle were exposed through a flank incision, the pedicle was then clamped and the kidney excised. The kidney was cut transversely into six sections (the poles were discarded) and as much medulla as possible (the position of the arcuate vessels was used to determine the corticomedullary junction) was rapidly separated from the cortex with stainless-steel cork borers of appropriate size. The kidney sections were immediately placed in liquid nitrogen, in a CO<sub>2</sub>-free, nitrogen atmosphere (6). Less than 30 sec elapsed from the time of clamping of the pedicle to the time of freezing the tissue in liquid nitrogen. Aliquots of frozen cortex and medulla were pulverized while immersed in liquid N<sub>2</sub>, in a stainless-steel mortar which was immersed in liquid nitrogen. Approximately 1-g aliquots of frozen cortical or medullary tissue were then transferred to tared flasks containing 0.1 N NaOH and 0.5% ferric fluoride (as enzyme inhibitor) (4). The flasks were immediately closed, the tissue allowed to thaw, and the wet weight of the tissue samples was taken. Determination of the total tissue CO<sub>2</sub> content was done in triplicate using the manometric Van Slyke apparatus according to the method of Danielson and Hastings (4). All handling of the frozen tissue samples was done in a nitrogen atmosphere free of CO<sub>2</sub> (6).

**Results.** The results of all five experiments done are summarized in Table I. The total acid-labile CO<sub>2</sub> content of renal medullary tissue is significantly greater than that of renal cortical tissue. The variability in the determination of total CO<sub>2</sub> content was similar in tissue samples from the same animal and in tissue samples from different animals. In the one experiment (Expt. 3) where only one

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TABLE I. Acid-Labile CO<sub>2</sub> Content in Dog Kidney.

Expt.	pH		Acid-labile CO <sub>2</sub> content ( $\mu$ moles/g wet wt) <sup>a</sup>		
	Blood	Urine	Cortex	Medulla	Corticomedullary difference <sup>b</sup>
1	—	—	10.50 $\pm$ .46 (3)	13.25 $\pm$ .19 (3)	—2.73
2	7.25	6.98	13.36 $\pm$ 1.55 (3)	16.50 $\pm$ 1.39 (3)	—3.17
3	7.32	7.01	16.25 (1)	16.22 $\pm$ 1.22 (4)	+0.03
4	7.24	7.00	14.58 $\pm$ .50 (4)	16.77 $\pm$ .31 (3)	—2.19
5	7.28	7.30	10.68 $\pm$ .60 (3)	16.17 $\pm$ 1.14 (2)	—5.49
Means $\pm$ 1 SE			13.07 $\pm$ .99	15.78 $\pm$ .57	—2.71 $\pm$ .79
No. of experiments			(5)	(5)	(5)
					<i>p</i> < .05

<sup>a</sup> Means  $\pm$  1 SE are presented. The number in parentheses indicates the number of separate CO<sub>2</sub> determinations done on a single tissue sample.

<sup>b</sup> Negative figures indicate that the CO<sub>2</sub> content is higher in medulla.

cortical sample was analyzed, the renal cortical CO<sub>2</sub> content exceeded that in the renal medulla. Comparison of the CO<sub>2</sub> content observed in this experiment with those of the other experiments, indicated that this difference is due to a relatively high cortical CO<sub>2</sub> content rather than to a decrease in medullary CO<sub>2</sub> content.

The observed greater total CO<sub>2</sub> content of renal medullary tissue might arise from a difference in water content in the two areas of the kidney (7). However, in dog, the medullary water content exceeds that of the cortex (tissue water is 81%  $\pm$  0.3% of wet weight in the medulla vs 78%  $\pm$  0.3% in the cortex (7)). In Table II are summarized the calculated total acid-labile CO<sub>2</sub> contents of dog medullary and cortical tissues expressed per gram of wet or dry weight, and also per gram of tissue water. The medullary CO<sub>2</sub> content per gram wet weight is higher by 17%

than that of the cortex and higher by 39% when expressed per gram dry weight. Thus, the acid-labile CO<sub>2</sub> concentration in medullary tissue H<sub>2</sub>O is greater than in cortical tissue water.

*Discussion.* Our observations do not permit us to identify whether it is the carbamino-CO<sub>2</sub>, the gaseous CO<sub>2</sub>, or the [HCO<sub>3</sub><sup>-</sup>] which accounts for the high medullary acid-labile CO<sub>2</sub> content.

Any difference in carbamino-CO<sub>2</sub> content between renal cortex and medulla would be due to a difference in the concentration of terminal amino groups of proteins between the two regions (1). Although there is a corticomedullary gradient for albumin (8), it is doubtful whether the increased albumin content could form enough carbamino-CO<sub>2</sub> to account for the observed difference in CO<sub>2</sub> content. Further, the concentration of hemoprotein in medulla is lower than in cortex

TABLE II. Renal Acid-Labile CO<sub>2</sub> Content on Wet- and Dry-Weight Basis.

	Total acid-labile CO <sub>2</sub> content <sup>a</sup>		
	Cortex	Medulla	Cortex—medulla
$\mu$ moles/g wet wt	13.07 $\pm$ 0.99	15.78 $\pm$ 0.57	—2.71 $\pm$ 0.79
$\mu$ moles/g dry wt	59.40 $\pm$ 4.50	83.10 $\pm$ 3.0	—23.70 $\pm$ 5.44
$\mu$ moles/g tissue H <sub>2</sub> O	16.75 $\pm$ 1.27	19.45 $\pm$ 0.71	—2.70 $\pm$ 1.48

<sup>a</sup> Means  $\pm$  1 standard error of the mean are presented. The standard errors for the derived data (i.e., per gram dry weight and per gram of tissue water) were calculated as the geometric mean of the standard error for the CO<sub>2</sub> content (per gram wet weight) and for the renal tissue water content (9).

(9). Thus, it is unlikely that carbamino-CO<sub>2</sub> is the basis for the high CO<sub>2</sub> content of medulla.

During alkalosis the PCO<sub>2</sub> of the urine and presumably of the medulla may be as high as 200 mm Hg. (10). Assuming a solubility coefficient for CO<sub>2</sub> in renal medullary tissue water of 0.2834  $\mu$ moles/ml per mm Hg (which is similar to that in whole blood (11)), and assuming a tissue water content of 81% of wet tissue weight (7), the concentration of dissolved CO<sub>2</sub> at PCO<sub>2</sub> of 200 mm Hg is 4.58  $\mu$ moles/g wet weight. Thus, in alkalosis, if the PCO<sub>2</sub> in the medulla and in the urine were the same, the medullary PCO<sub>2</sub> alone could account for acid-labile CO<sub>2</sub> content of renal medulla which is higher than in cortical tissue.

If we now assume that the higher medullary acid-labile CO<sub>2</sub> content observed in our experiments is due entirely to a difference in PCO<sub>2</sub> this would require that the medullary PCO<sub>2</sub> be 95 mm Hg greater than the cortical PCO<sub>2</sub>. If cortical PCO<sub>2</sub> is 45 mm Hg (similar to venous PCO<sub>2</sub>) a medullary CO<sub>2</sub> tension of 140 mm Hg would be necessary to account for our experimental observations as being due entirely to a high medullary PCO<sub>2</sub>. However, the reported urine PCO<sub>2</sub> under experimental circumstances similar to ours is not above 100 mm Hg (12). Further, Ulich *et al.* (13) have recently reported that the PCO<sub>2</sub> in vasa recta blood of normal rats (during NaCl infusion) is only  $\sim$  10 mm Hg greater than that of arterial blood. If the PCO<sub>2</sub> of vasa recta blood is of similar magnitude in dog, the contribution of the gaseous CO<sub>2</sub> moiety to the total CO<sub>2</sub> of medulla is minimal. Thus, the high medullary total CO<sub>2</sub> content reported here is best accounted for by an accumulation of HCO<sub>3</sub><sup>-</sup> in the medulla. Further, since Ulich *et al.* (13) observed no increase in HCO<sub>3</sub><sup>-</sup> of vasa recta blood above that

present in arterial blood, most of the increased HCO<sub>3</sub><sup>-</sup> content is located in the extravascular compartment of the medullary tissue, and most probably in the intracellular compartment.

**Summary.** Total acid-labile CO<sub>2</sub> was measured in aliquots of dog renal cortex and medulla frozen in liquid nitrogen. A higher (15.87  $\mu$ moles/g wet weight vs 13.55  $\mu$ moles/g wet weight,  $p < .05$ ) acid-labile CO<sub>2</sub> content was found in the renal medulla. It is suggested that the higher medullary total acid-labile CO<sub>2</sub> content is primarily due to a high [HCO<sub>3</sub><sup>-</sup>] in the medullary intracellular fluid rather than to a high medullary PCO<sub>2</sub>.

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