

## The Inhibition by ADP of NADPH-Supported Adrenal Steroid 11 $\beta$ -Hydroxylation<sup>1</sup> (39371)

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Accumulating evidence suggests the possibility that, in the adrenal cortex, the sequence of reactions from cholesterol to corticosterone or cortisol may be functionally a highly integrated system. In addition to the primary control by the adrenocorticotrophic hormone (ACTH) which takes place at the transformation of cholesterol to pregnenolone, a series of secondary controls of the steroidogenic pathway appear to be present. Thus, a number of investigations have shown a relationship between steroid 11 $\beta$ -hydroxylation and oxidative phosphorylation (1-5) and steroid 11 $\beta$ -hydroxylation can inhibit the conversion of cholesterol to pregnenolone but the reverse situation does not occur (6). Ca<sup>2+</sup> has been found to stimulate both pregnenolone synthesis (7) and 11 $\beta$ -hydroxylase (8). Reduced NAD<sup>+</sup> inhibits the formation of progesterone from pregnenolone and this inhibition can be reversed by ascorbic acid and other substances (9). Both NAD<sup>+</sup> and NADH stimulate  $\Delta$ 5-3-keto-steroid isomerase (10).

It has been found recently that the succinate supported 11 $\beta$ -hydroxylation of deoxycorticosterone (DOC) to form corticosterone in rat adrenal mitochondria is inhibited by ADP (11). Much of the data can be explained on the basis of a competition for reducing equivalents between the hydroxylase system and the cytochrome chain. However, some observations suggested the possibility that ADP may also effect the hydroxylase system directly. In this report data is presented which indicate that ADP can indeed directly inhibit steroid 11 $\beta$ -hydroxylation.

**Materials and methods.** Mitochondria were prepared from rat adrenals as described previously (12). Following the second 0.25 M sucrose wash, the mitochondrial

pellet was suspended in a volume of 12 mM Tris, pH 7.5, equal to that of the original homogenate (90 mg wet weight of tissue per ml) and contained 2.0 to 2.5 mg of protein per ml as determined by the method of Lowry *et al.* (13).

The complete system for the 11-hydroxylation of deoxycorticosterone (DOC) to form corticosterone consisted of 0.2 ml of the mitochondrial preparation, 36 mM Tris, pH 7.5, 20 mM CaCl<sub>2</sub>, 30  $\mu$ M DOC, 0.07 ml of NADPH prepared by a preliminary incubation for 15 min at room temperature of a solution containing, per ml, 25  $\mu$ mol of NaNADP<sup>+</sup>, 70 of  $\mu$ mole Na<sub>2</sub> glucose-6-P, and about 2000 units of glucose-6-P dehydrogenase, other additions as indicated and 0.25 M sucrose to a final volume of 1.25 ml. The DOC was added in 0.01 ml of ethanol and an incubation control with 0.01 ml of ethanol without DOC was routinely carried out. The incubations were carried out at 20° in a Dubanoff shaking incubator for 6 min, at which time 0.4-ml aliquots were delivered into 3 ml of dichloromethane and, after shaking and centrifugation, 2 ml of the dichloromethane taken for corticosterone determination by a modification (12) of the fluorometric method of Glick *et al.* (14).

**Results.** The incubation conditions used permit the 11 $\beta$ -hydroxylation of deoxycorticosterone to be supported directly by exogenous NADPH. This would obviate a competition by the hydroxylase and the cytochrome chain for reducing equivalents derived from a Krebs cycle acid such as succinate. It is seen from the data in Table I that 11 $\beta$ -hydroxylase supported by NADPH as the source of electrons is inhibited by ADP and ATP. ADP is more inhibitory than ATP, as was found with intact mitochondria (11) and that a clear inhibition is seen at concentrations as low as 50  $\mu$ M.

Ca<sup>2+</sup>, which is present in these incubations, is known to induce swelling in adrenal

<sup>1</sup> This work has been supported by Grant No. AM-13361 from the United States Public Health Service.

TABLE I. THE EFFECT OF ADP AND ATP ON THE FORMATION OF CORTICOSTERONE FROM DEOXYCORTICOSTERONE.

Nucleotide concentration (mM)	ADP		ATP	
	n mole corticosterone per ml	Percentage change	n mole corticosterone per ml	Percentage change
0	13.0		13.0	
0.01	13.0	0	13.4	0
0.05	9.07	-30	12.1	-7
0.10	6.34	-51	11.5	-12
0.50	2.88	-78	7.06	-46
1.0	1.58	-88	4.90	-62

mitochondria (8, 15) and ADP and ATP have been found to inhibit this swelling (15). The inhibition of the 11-hydroxylase by ADP may result from a modification of the mitochondrial membranes so that the entry of NADPH is affected. Several experiments were carried out to explore this possibility. The data in Table II show that the presence of several different types of detergents, which would be expected to disrupt mitochondrial structure, the inhibition by ADP persists. In the presence of some detergents the sensitivity of the hydroxylase to  $\text{Ca}^{2+}$  changes and  $\text{Ca}^{2+}$  concentration has been adjusted to obtain maximal activity.

In the experiments presented in Table III the effects on the ADP inhibition of preincubation of the mitochondria under various conditions has been determined. It is seen that the ADP inhibition is still present after a 10-min preincubation in the presence of  $\text{Ca}^{2+}$ , NADPH, and ADP or in the presence of  $\text{Ca}^{2+}$  and NADPH. There is a small decrease in the inhibition under the latter preincubation condition.

**Discussion.** A number of observations indicates that the ADP effect is due to an action directly upon the hydroxylase system. The ADP inhibition persists in the presence of various detergents as well as when the mitochondria are preincubated with  $\text{Ca}^{2+}$  and NADPH where the possibility of an ADP induced impairment of NADPH entry into the mitochondrial membranes is not present. In support of this conclusion is the observation that while ADP is more inhibitory than ATP, ATP is more effective than ADP in preventing the  $\text{Ca}^{2+}$  induced swelling of adrenal mitochondria (15). The ADP inhibition of 11 $\beta$ -hydroxylase activity is

TABLE II. THE INHIBITION OF 11-HYDROXYLATION BY ADP IN THE PRESENCE OF VARIOUS DETERGENTS.<sup>a</sup>

Addition	ADP		
	n mole corticosterone/ml	Percentage change	
Lubrol	18.0	1.8	-90
Triton X-100	13.8	1.5	-89
Deoxycholate	18.1	5.8	-68

<sup>a</sup> ADP, when present, was 2 mM. The concentrations of the detergents, per milliliter of incubation medium, were Lubrol, 40  $\mu\text{g}$ ; Triton X-100, 80  $\mu\text{g}$ ; Na deoxycholate, 80  $\mu\text{M}$ .  $\text{Ca}^{2+}$  concentrations in incubations with Lubrol was 20 mM, and in those with Triton X-100 and deoxycholate were 8 mM.

TABLE III. THE EFFECT OF PREINCUBATION IN THE ABSENCE AND PRESENCE OF ADP ON THE INHIBITION OF 11-Hydroxylase by ADP.<sup>a</sup>

Preincubation	ADP		
	n mole corticosterone/ml	Percentage change	
A	17.2	1.5	-91
B	22.2	5.2	-77
C	21.3	2.3	-89

<sup>a</sup> Preincubation: A, none. B, 10 min with the complete system except for DOC. ADP, when present, was added in the final incubation. The reaction was started with DOC. C, the preincubation was the same as in B except that ADP, when added, was present during the preincubation. The reaction was started with DOC. ADP, when present, was 2 mM.

somewhat less when the mitochondria are preincubated with  $\text{Ca}^{2+}$  and NADPH (77%) than when ADP is also present during this preincubation (89%). This may indicate a possible minor effect of ADP upon NADPH entry into the mitochondrial membranes. Further work will be required to clarify the mechanism of the ADP inhibition as well as the  $\text{Ca}^{2+}$  stimulation of this enzyme system.

**Summary.** The 11 $\beta$ -hydroxylation of deoxycorticosterone to form corticosterone in adrenal mitochondria has been found to be inhibited by ADP and ATP, with ADP being the more inhibitory of the two. The evidence suggests that the ADP directly affects the enzyme system.

1. Brownie, A. C., and Grant, J. K., *Biochem. J.* **57**, 255 (1954).
2. Harding, B. W., Wilson, L. D., Wong, S. H., and Nelson, D. H., *Steroids*, Supplement II, p. 51 (1965).
3. Crammer, W., and Estabrook, R. W., *Arch. Bio-*

- chem. Biophys. **122**, 721 (1967).
4. Sauer, L. A., and Mulrow, P., Arch. Biochem. Biophys. **134**, 486 (1969).
  5. Sauer, L. A., Arch. Biochem. Biophys. **139**, 340 (1970).
  6. Young, D. G., and Hall, P. F., Biochemistry **10**, 1496 (1971).
  7. Koritz, S. B., Biochim. Biophys. Acta **56**, 63 (1962).
  8. Péron, F. G., Guerra, F., and McCarthy, J. L., Biochim. Biophys. Acta **110**, 277 (1965).
  9. Koritz, S. B., Arch. Biochem. Biophys. **100**, 349 (1963).
  10. Oleinick, N. L., and Koritz, S. B., Biochemistry **5**, 715 (1966).
  11. Koritz, S. B., and Moustafa, A. M., J. Steroid Biochem., in press (1976).
  12. Koritz, S. B., and Kumar, A. M., J. Biol. Chem. **245**, 152 (1970).
  13. Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. J., J. Biol. Chem. **193**, 265 (1951).
  14. Glick, D., Redlich, D. V., and Levine, S., Endocrinology **74**, 653 (1964).
  15. Hirshfield, I. N., and Koritz, S. B., Biochemistry **3**, 1994 (1964).
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Received January 23, 1976. P.S.E.B.M. 1976, Vol. 152.