

Evidence That Thyroxine Inhibits either Basal or TRH-Induced TSH Secretion Only after Conversion to Triiodothyronine¹ (42679)

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Abstract. When TRH was administered every 15 min for 2 hr in euthyroid rats, equivalent modestly supraphysiologic doses of either T₄ or T₃ suppressed TRH-induced TSH secretion after 45 min. Pretreatment with iopanoic acid blocked the ability of T₄ but not of T₃ to suppress TRH-induced TSH secretion 2 hr after administration of the respective thyroid hormone. Pretreatment with iopanoic acid also blocked the ability of T₄, but not of T₃, to depress the elevated basal plasma TSH concentration of hypothyroid rats within 2 hr. Propylthiouracil did not significantly inhibit the ability of T₄ to depress TRH-induced TSH secretion and only slightly depressed the ability of T₄ to reduce the elevated plasma TSH of hypothyroid rats. Our data support the concept that although equivalent physiologic doses of T₄ or T₃ inhibit basal or TRH-induced TSH secretion equally rapidly, TSH inhibition produced by T₄ is probably dependent on its rapid conversion to T₃, either within the pituitary or peripherally. T₃ thus seems to be exerting almost all the negative feedback effects on TSH secretion under the conditions of our experiments. © 1988 Society for Experimental Biology and Medicine.

Whether both T₄ and T₃ possess an innate ability to inhibit TSH secretion and exert hormonal action (2, 3) or whether T₄ must first be converted to T₃ to exert these effects has not been unequivocally settled (4-11). Although the nuclear binding affinity for T₃ is much greater than that for T₄ (12), it is not certain that all thyroid hormone action depends on nuclear binding (13-16). Both T₄ and T₃ depress basal TSH secretion and inhibit the ability of TRH to stimulate TSH secretion within 1 hr. The rapid effect of T₄ to inhibit TSH secretion has been postulated by Larsen and Silva to be due to intrapituitary conversion of T₄ to T₃ (6, 14). Iopanoic acid inhibits conversion of T₄ to T₃ (17-20) and significantly depresses the ability of T₄, but not of T₃, to inhibit basal TSH secretion several hours after administration of thyroid hormone. However, previous studies by Boado *et al.* (3) made less than 30 min after T₄ or T₃ administration have indicated that iopanoic acid does not significantly inhibit the ability of T₄ to depress TRH-induced TSH secretion. This suggests that T₄ may

have a direct inhibitory effect on this phenomenon. We therefore designed the following experiments to explore this problem further.

Materials and Methods. Adult 350- to 400-g male Simonsen rats were used in all experiments. All drugs were given in a dose adjusted per 100 g body weight. TRH was given as a 1- μ g iv bolus via atrial catheter every 15 min for 2 hr or under light ether anesthesia via jugular puncture. T₄ was given as 1 or 2.5 μ g iv; T₃ was given as 0.25 or 0.625 μ g iv. Some rats were made hypothyroid with 0.03% MMI in the drinking water for 2 weeks. Some rats were given 5 mg iopanoic acid or 2 mg PTU ip 24, 16, and 1.5 hr before beginning TRH injections. All blood samples were taken via an atrial catheter or by jugular puncture.

All experiments were performed between 0830 and 1600 hr. An indwelling intraatrial catheter was implanted through the jugular vein and fed out subcutaneously through the top of the head 18 hr before the experiment. Following implantation of the cannula, the rats were maintained in an isolated room in individual metal wastebaskets with sawdust in the bottom and with free access to food and water. The extracorporeal portion of the cannula was suspended above the rat's head

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to allow free mobility. Heparin in 0.9% saline (1 USP U/ml) was maintained in the cannula between sampling and injection, which were performed through the same cannula.

TRH administration and blood sampling. Bolus injections of TRH (Sigma) were given through the catheter every 15 min for 2 hr, using a 1-ml syringe. The TRH was diluted to give a concentration of 1 μ g in 50 μ l of saline. Blood samples of 0.25 ml were taken at each indicated time and replaced with an equal volume of 0.9% saline to minimize blood volume depletion. Samples were centrifuged at 4°C. Plasma was separated and frozen at -20°C until analyzed.

TSH assay and statistics. All hormone assays from any given experiment were performed at the same time. TSH was measured by radioimmunoassay of 0.1 ml of plasma or 1:1000 diluted pituitary homogenates, using rat TSH kits kindly supplied by the National Hormone and Pituitary Program, NIADDK. The stated potency of the reference standard supplied (NIAMDD-rTSH-RP-1, 0.22 USP U/mg) was used to calculate plasma hormone concentration. Our range of sensitivity was 15–1000 ng/tube (intraassay variation <7%, interassay variation <10%). Statistical analysis was performed by using Student's paired or unpaired *t* test and the Newman-Keuls multiple comparison test.

Results. Dose and temporal characteristics of T₄ and T₃ inhibition of TRH-induced TSH secretion. The first set of experiments was designed to determine for both T₄ and T₃ the dose and time parameters necessary to obtain a clear depression of TRH-induced TSH secretion. We had previously established (21, 22) that repetitive bolus doses of 1 μ g TRH iv through an indwelling atrial catheter in freely moving unanesthetized euthyroid rats induced a biphasic rise in plasma TSH concentration with an initial sharp rise at 15 min, a nadir at 60 min, and a second rise with a slower rate of increase and lower but sustained amplitude which usually peaked at approximately 90 min, as shown in the top panel, Fig 1.

Intravenous injection of 1 μ g T₄ or 0.25 μ g T₃ at time 0 did not appreciably alter this pattern (lower panels, Fig 1.) However, increasing the dose to 2.5 μ g T₄ or 0.625 μ g T₃ caused a significant suppression of TRH-in-

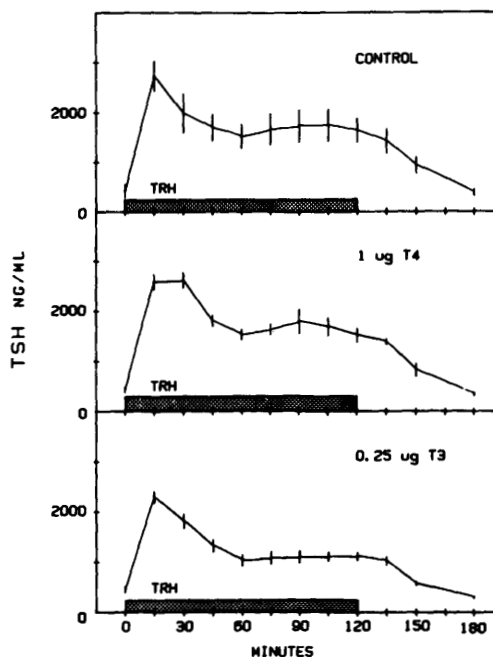


FIG. 1. Effect of 1 μ g T₄ or 0.25 μ g T₃ at 0 min on TRH-induced TSH secretion in euthyroid rats. There were four animals in each of the three groups identified by the legend in each panel. All animals had chronic indwelling cannulas. One microgram TRH per one hundred grams body weight was injected every 15 min for 2 hr as indicated by the horizontal stippled bar in each panel. The mean $\pm$ SE of plasma TSH concentration is indicated by each point and vertical line.

duced TSH secretion but only after 45 min (Fig 2).

Effect of iopanoic acid on T₄- and T₃-induced inhibition of TRH-induced TSH secretion in euthyroid rats. The above experiments indicated that a reasonable time to test the early effects of blocking intrapituitary T₄ to T₃ conversion with iopanoic acid would be 2 hr after administration of T₄. Accordingly, plasma TSH was measured immediately before and 15 min after a bolus injection of 1 μ g TRH; 2.5 μ g T₄ was given immediately after the second blood sampling. Two hours later, the TRH test was repeated. One group was pretreated with iopanoic acid, a drug which blocks conversion of T₄ to T₃ in both the pituitary and the peripheral tissues; one group was pretreated with propylthiouracil, a drug which blocks peripheral but not intrapituitary conversion of T₄ to T₃ (23–27), and

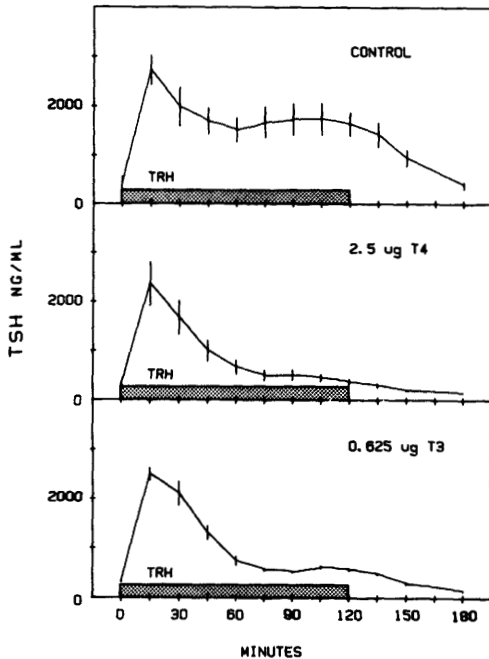


FIG. 2. Effect of 2.5 μ g T₄ or 0.625 μ g T₃ at 0 min on TRH-induced TSH secretion in euthyroid rats. The data are shown as in Fig. 1. There were four rats in each of the control and T₄ groups and three rats in the T₃ group.

a control group was not pretreated with drugs and was given saline instead of thyroid hormone at the end of the first TRH test.

There was no significant difference in the response to TRH between the first and second tests in the control rats injected with saline (Fig. 3). Treatment with 2.5 μ g T₄ at the end of the first TRH test nearly completely blocked the rise in plasma TSH from the second TRH test 2 hr later. The blocking effect of T₄ was prevented by pretreatment with iopanoic acid but not by pretreatment with propylthiouracil.

The iopanoic acid study was repeated using 0.625 μ g T₃ instead of the equivalent dose of 2.5 μ g T₄ (Fig. 4). Iopanoic acid had no effect on the ability of T₃ to block TRH-induced TSH secretion.

Studies in hypothyroid rats. Similar studies were also made in hypothyroid rats. We have previously reported that the biphasic TSH response pattern seen with repetitive or continuous TRH administration in euthyroid rats is not seen in hypothyroid rats (22). Instead, there is only an initial stimulation of secretion, followed by a return of plasma TSH concentration to or slightly below the

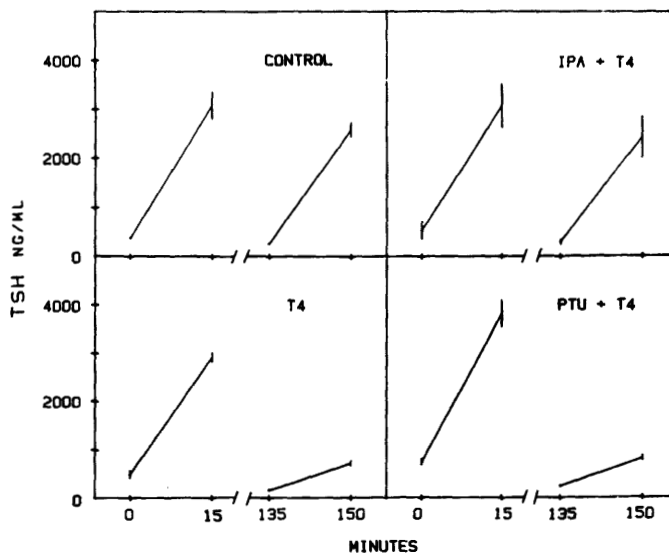


FIG. 3. Effect of iopanoic acid or propylthiouracil on 2.5 μ g T₄ inhibition of TRH-induced TSH secretion. There were four of five rats in each group. Plasma TSH is shown as mean $\pm$ SE. An intrajugular bolus injection of 1 μ g TRH was given at 0 and 135 min. Jugular blood was sampled just before and 15 min after TRH injections for plasma TSH concentration. T₄ (2.5 μ g) was given in a bolus iv injection immediately after the 15 min blood sampling. Pretreatment with iopanoic acid (IPA) and propylthiouracil (PTU) was as indicated in the text.

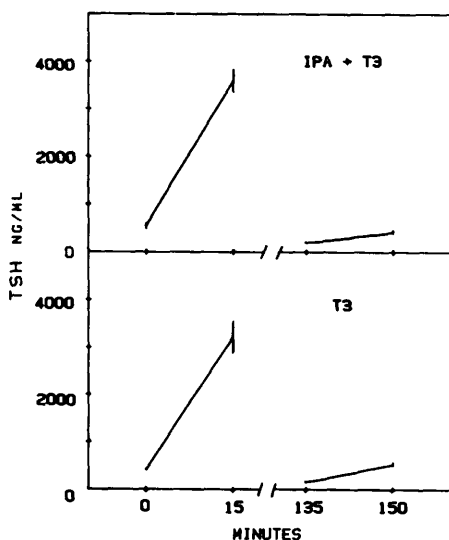


FIG. 4. Effect of iopanoic acid on 0.625 µg T₃ inhibition of TRH-induced TSH secretion. There were four animals in each group. The experiment was analogous and the data are plotted as in Fig. 3.

initial basal levels. There was no significant effect of either 2.5 µg T₄ or 0.625 µg T₃ given at 0 time on TRH-induced TSH secretion in hypothyroid rats (Fig. 5), in contrast to the clear effect of both thyroid hormones given in these concentrations in the euthyroid rats. However, plasma TSH was significantly lower in both the T₄- and T₃-treated groups than in the controls at 180 min ($P < 0.05$).

The spontaneous refractoriness of the hypothyroid rat to repetitive TRH precluded an exact repetition of the euthyroid studies in hypothyroid animals. Therefore, we examined the ability of iopanoic acid and propylthiouracil to block the respective abilities of T₄ or T₃ to depress basal plasma TSH secretion in hypothyroid rats 2 hr after their administration. Similar studies have been reported by Larsen examining the plasma TSH at the slightly later interval of 3 hr and using smaller doses (6).

Our data confirm the earlier report of Larsen (6) and are consistent with the TRH studies in the euthyroid rat presented above. Either 2.5 µg of T₄ (Fig. 6) or 0.625 µg of T₃ (Fig. 7) caused a significantly greater depression of basal plasma TSH in hypothyroid rats 2 hr later than did administration of saline.

The effect of T₄, but not that of T₃, was blocked by iopanoic acid. Propylthiouracil partially blocked the effect of T₄ (Fig. 6), but to a much lesser extent than did iopanoic acid. Propylthiouracil was not tested with T₃.

Discussion. The primary object of these studies was to determine whether the early effect of T₄ on inhibiting TRH-induced TSH secretion was more likely due to a direct effect of T₄ on the thyrotroph or to the rapid conversion of T₄ to T₃ within the pituitary cell. The report of Boado *et al.* (3) indicated that there was no inhibition of the early T₄ suppression of TRH-induced TSH secretion in rats treated with iopanoic acid, a substance known to markedly depress intrapituitary conversion of T₄ to T₃, thus suggesting that T₄ itself might have some intrinsic action. Other evidence had suggested that the action of T₄ at the pituitary might be entirely dependent on its conversion to T₃ (6–9, 20, 25). Two potential problems in interpreting the data of Boado *et al.* are that quite large supraphysiologic doses of T₄ were given (10

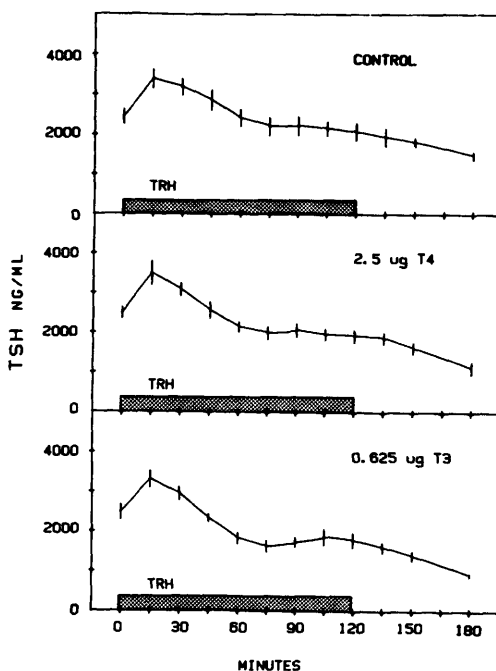


FIG. 5. Effect of 2.5 µg T₄ or 0.625 µg T₃ at 0 min on TRH-induced TSH secretion in hypothyroid rats. There were four rats in each group. The data are plotted as in Fig. 1.

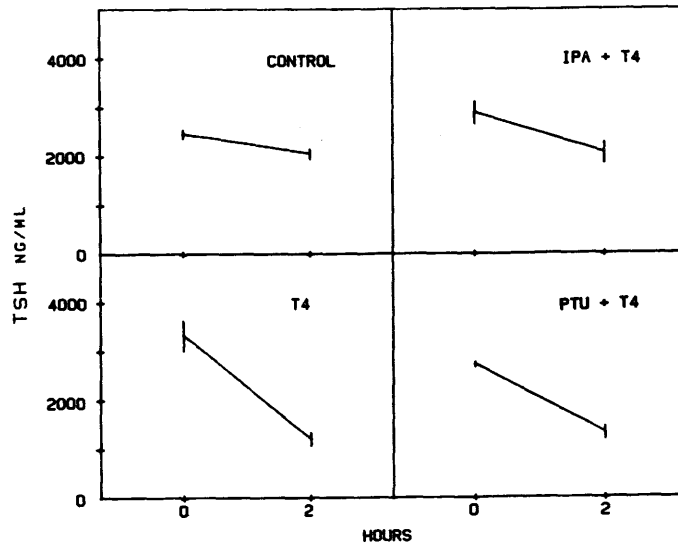


FIG. 6. Effect of iopanoic acid or PTU on 2.5 µg T₄ inhibition of basal TSH secretion in hypothyroid rats. There were four to five rats in each group. T₄ was given in a bolus intrajugular injection at 0 h. Jugular blood was sampled for TSH concentration just before and 2 hr after T₄ injection. There was a slight drop in plasma TSH in the vehicle-injected control group. However, there was a significantly greater drop in plasma TSH in the group given T₄ alone ($P < 0.01$). The group given IPA plus T₄ was not significantly different from the control ($P > 0.05$). However, that given PTU plus T₄ was statistically different from both the control ($P < 0.05$) and the T₄ only ($P < 0.05$) groups.

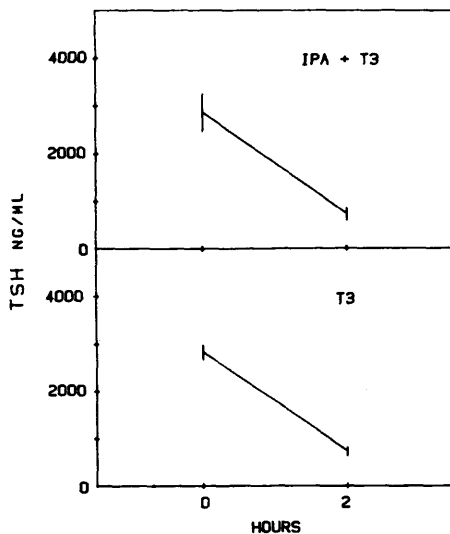


FIG. 7. Effect of iopanoic acid on 0.625 µg T₃ inhibition of basal TSH secretion in hypothyroid rats. The experiment was performed as in Fig. 6. There were five animals in the IPA plus T₃ group and four in the T₃ only group. There was no significant difference between the two groups in the depression of plasma TSH induced by T₃ ($P > 0.1$).

µg/100 g body wt) and the study was made within 30 min after administration of T₄ to minimize any possible conversion of T₄ to T₃.

Our initial experiments were designed to determine the earliest time at which a clear inhibition by either T₄ or T₃ of the TRH-induced TSH stimulation could be detected and to ascertain the smallest dose of each hormone which could unequivocally produce this inhibition. We felt that such knowledge would permit us to evaluate more conclusively whether there might be a direct effect of T₄ on this phenomenon. T₃ was approximately four times more potent than T₄ in inhibiting TRH-induced TSH secretion under the conditions of our experiments, as would be expected from a vast number of earlier comparisons using a variety of parameters. Clear inhibition of TRH-induced TSH secretion could be seen only after 45 min from the time of administration of either thyroid hormone.

Our data indicate that the failure of Boado *et al.* to detect an inhibitory effect of iopan-

oic acid on the ability of T₄ to depress TRH-induced TSH secretion may be because they made their study less than 30 min after administering T₄, probably earlier than a significant effect of T₄ or T₃ could be detected. Also, Boado *et al.* used the rather high dose of 10 µg T₄/100 g body wt. Under these conditions there may have been some low-affinity binding of T₄ to nuclear or other receptors to induce an inhibition of TSH secretion which would not be seen at T₄ concentrations closer to normal.

In our experiments, pretreatment with iopanoic acid blocked the ability of T₄ but not of T₃ to suppress TRH-induced TSH secretion 2 hr after administration of the respective thyroid hormone. Pretreatment with iopanoic acid also blocked the ability of T₄, but not of T₃, to depress the elevated basal plasma TSH concentration of hypothyroid rats within 2 hr.

In conclusion, our data are consistent with the theory proposed by Silva and Larsen (6, 14) that T₄ inhibits TSH secretion only after conversion to T₃, whether this conversion is in peripheral tissues or within the pituitary itself. It thus seems likely that T₄ conversion to T₃ must occur very rapidly and the quantity of T₃ thus generated is sufficient to inhibit TSH secretion.

1. Maruta S, Greer MA. Evidence that thyroxine inhibits TRH-induced TSH secretion only after conversion to 3,5,3'-triiodothyronine. Abstr 60th Ann Mtg Am Thyroid Assoc. pT-42, 1984.
2. Fukuda H, Yasuda N, Greer MA. Acute effects of thyroxine, triiodothyronine, and iodide on thyrotropin secretion. *Endocrinology* 97:924-931, 1975.
3. Boado R, Ulloa E, Zaninovich A. The effects of thyroxine and triiodothyronine on the thyrotropin response to thyrotropin-releasing hormone in normal and iopanoic acid-treated euthyroid rats. *Acta Endocrinol* 101:205-209, 1982.
4. Chopra IJ, Solomon DH, Chua Teco GN. Thyroxine: Just a hormone or a prohormone too. *J Clin Endocrinol Metab* 36:1050-1057, 1973.
5. Frumess RD, Larsen PR. Correlation of serum triiodothyronine (T₃) and thyroxine (T₄) with biologic effects of thyroid hormone replacement in propylthiouracil-treated rats. *Metabolism* 24:547-554, 1975.
6. Larsen PR, Dick TE, Markovitz BP, Kaplan MM, Gard TG. Inhibition of intrapituitary thyroxine to 3,5,3'-triiodothyronine conversion prevents the autocrine suppression of thyrotropin release by thyroxine in hypothyroid rats. *J Clin Invest* 64:117-128, 1979.
7. Kleinmann RE, Vagenakis AG, Braverman LE. The effect of iopanoic acid on the regulation of thyrotropin secretion in euthyroid subjects. *J Clin Endocrinol Metab* 51:399-403, 1980.
8. Burman KD, Smallridge RC, Burge JR, Carlson D, Wartofsky L. Iodate restores the fasting-induced decrement in thyrotropin secretion. *J Clin Endocrinol Metab* 57:597-602, 1983.
9. Kaplan MM. The role of thyroid hormone deiodination in the regulation of hypothalamo-pituitary function. *Neuroendocrinology* 38:254-260, 1984.
10. Schaison G, Thomopoulos P, Leguillouzie D, Thomas G, Moatti M. Effects of sodium ipodate and propylthiouracil in athyreotic human subjects, role of triiodothyronine and pituitary thyroxine monodeiodination in thyrotrophin regulation. *Acta Endocrinol (Copenhagen)* 106:338-345, 1984.
11. Ikeda H, Uchimura H, Greer MA. Comparison of the ability of thyroxine and triiodothyronine to suppress TRH-induced TSH secretion by perfused rat anterior pituitary fragments. *Neuroendocrinology* 41:79-82, 1985.
12. Surks MI, Oppenheimer JH. Concentration of L-thyroxine and L-triiodothyronine specifically bound to nuclear receptors in rat liver and kidney. Quantitative evidence favoring a major role of T₃ in thyroid hormone action. *J Clin Invest* 60:555-562, 1977.
13. Oppenheimer JH, Schwartz HL, Surks MI. Tissue differences in the concentration of triiodothyronine nuclear binding sites in the rat: Liver, kidney, pituitary, heart, brain, spleen, and testis. *Endocrinology* 95:897-903, 1974.
14. Silva JE, Larsen PR. Pituitary nuclear 3,5,3'-triiodothyronine and thyrotropin secretion: An explanation for the effect of thyroxine. *Science* 198:617-620, 1977.
15. Schrey MP, Larsen PR. Evidence for a possible role of Ca⁺⁺ in the 3,5,3'-triiodothyronine inhibition of thyrotropin-releasing hormone-induced secretion of thyrotropin by rat anterior pituitary in vitro. *Endocrinology* 108:1690-1696, 1981.
16. Belchetz PE, Gredley G, Bird D, Himsworth RL. Regulation of thyrotropin secretion by negative feedback of tri-iodothyronine. *J Endocrinol* 76:439-448, 1978.
17. Kaplan MM, Utiger RD. Iodothyronine metabolism in rat liver homogenates. *J Clin Invest* 61:459-471, 1978.
18. Cheron RG, Kaplan MM, Larsen PR. Physiological and pharmacological influences on thyroxine to 3,5,3'-triiodothyronine conversion and nuclear 3,5,3'-triiodothyronine binding in rat anterior pituitary. *J Clin Invest* 64:1402-1414, 1979.
19. Crantz FR, Larsen PR. Rapid thyroxine to 3,5,3'-triiodothyronine conversion and nuclear 3,5,3'-tri-

- iodothyronine binding in rat cerebral cortex and cerebellum. *J Clin Invest* **65**:935-938, 1980.
20. Obregon MJ, Pascual A, Mallol J, Morreale de Escobar G, Escobar del Rey F. Evidence against a major role of L-thyroxine at the pituitary level: Studies in rats treated with iopanoic acid (Telepaque). *Endocrinology* **106**:1827-1836, 1980.
 21. Maruta S, Greer MA. Repeated TRH injections induce a biphasic rise in plasma TSH due to a reversible partial refractoriness of the euthyroid thyrotroph. Abstr 59th Ann Mtg Am Thyroid Assoc. pT-38, 1983.
 22. Maruta S, Greer MA. Repetitive or continuous thyrotropin-releasing hormone administration induces a biphasic rise in plasma thyrotropin in euthyroid but not in hyothyroid rats. *Endocrinology*, **121**:1946-1952, 1987.
 23. Oppenheimer JH, Schwartz HL, Surks MI. Propylthiouracil inhibits the conversion of L-thyroxine to L-triiodothyronine. An explanation of the anti-thyroxine effect of propylthiouracil and evidence supporting the concept that triiodothyronine is the active thyroid hormone. *J Clin Invest* **51**:2493-2497, 1972.
 24. Larsen PR, Frumess RD. Comparison of the biological effects of thyroxine and triiodothyronine in the rat. *Endocrinology* **100**:980-988, 1977.
 25. Silva JE, Larsen PR. Contributions of plasma triiodothyronine and local thyroxine monodeiodination to triiodothyronine to nuclear triiodothyronine receptor saturation in pituitary, liver, and kidney of hypothyroid rats. Further evidence relating saturation of pituitary nuclear triiodothyronine receptors and the acute inhibition of thyroid-stimulating hormone release. *J Clin Invest* **61**:1247-1259, 1978.
 26. Maeda M, Ingbar SH. Effect of alterations in thyroid status on the metabolism of thyroxine and triiodothyronine by rat pituitary gland in vitro. *J Clin Invest* **69**:799-808, 1982.
 27. Visser TJ, Kaplan MM, Leonard JL, Larsen PR. Evidence for two pathways of iodothyronine 5'-deiodination in rat pituitary that differ in kinetics, propylthiouracil sensitivity, and response to hypothyroidism. *J Clin Invest* **71**:992-1002, 1983.
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