

# Sulforaphane Retards the Growth of Human PC-3 Xenografts and Inhibits HDAC Activity in Human Subjects

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Sulforaphane (SFN) is an isothiocyanate found in cruciferous vegetables such as broccoli. This anticarcinogen was first identified as a potent inducer of Phase 2 enzymes, but evidence is mounting that SFN acts through other cancer chemopreventive mechanisms. We recently reported on a novel mechanism of chemoprotection by SFN in human colon cancer cells and prostate epithelial cells, namely the inhibition of histone deacetylase (HDAC). In the present investigation, we sought to test whether SFN also might inhibit HDAC activity *in vivo*. When consumed in the diet at an average daily dose of 7.5  $\mu$ mol per animal for 21 days, SFN suppressed the growth of human PC-3 prostate cancer cells by 40% in male nude mice. There was a significant decrease in HDAC activity in the xenografts, as well as in the prostates and mononuclear blood cells (MBC), of mice treated with SFN, compared to controls. There also was a trend towards increased global histone acetylation in the xenografts, prostates, and MBC. In human subjects, a single dose of 68 g BroccoSprouts inhibited HDAC activity significantly in peripheral blood mononuclear cells (PBMC) 3 and 6 hrs following consumption. These findings provide evidence that one mechanism through which SFN acts as a cancer chemopreventive agent *in vivo* is through the inhibition of HDAC activity. Moreover, the data suggest that HDAC activity in PBMC may be used as a biomarker for assessing exposure to novel dietary HDAC inhibitors in human subjects. *Exp Biol Med* 232:227–234, 2007

**Key words:** prostate cancer; histone deacetylase inhibitor; sulforaphane mechanisms of prevention; diet and cancer

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## Introduction

Targeting the epigenome, including the use of histone deacetylase (HDAC) inhibitors, is a novel strategy for cancer chemoprevention. Recently, drugs classified as HDAC inhibitors have shown promise as potential chemoprevention agents (1–5). HDAC inhibitors are interesting candidates for prevention because they enable re-expression of epigenetically-silenced genes involved in differentiation, cell cycle regulation, apoptosis, angiogenesis, invasion, and metastasis.

HDAC changes have been linked to the progression of several cancers, including prostate cancer (1, 3, 4, 6). For example, *HDAC1* mRNA levels are higher in several prostate cancer cell lines compared with benign prostate hyperplasia, with a concomitant increase in HDAC activity (7). Moreover, HDAC1 expression was increased in primary human prostate cancer tissue compared with prostate hyperplasia (7). Overexpression of HDAC1 in PC-3 cells resulted in an increase in cell proliferation and an overall decrease in cell differentiation (8). HDAC1 expression was highest in hormone refractory prostate cancer (8), suggesting that alterations in HDAC may be of particular importance in the progression to androgen independence. Importantly, inhibitors of HDAC, including suberoylanilide hydroxamic acid (SAHA), valproic acid, depsipeptide, and sodium butyrate have been demonstrated to be effective against prostate cancer in experimental models (9–11).

Epidemiologic evidence suggests that consumption of cruciferous vegetables decreases overall risk for prostate cancer, particularly during the early stages (12–15). Sulforaphane (SFN) is a constituent of cruciferous vegetables, found at high levels in broccoli and broccoli sprouts (16, 17). SFN has been identified as an effective cancer chemoprotective agent in animal models (17–19), as well as in xenograft models of prostate cancer (20). Although the majority of early studies focused on SFN as a potent Phase 2 enzyme inducer, recent work has implicated other mechanisms of SFN action (reviewed in Ref. 2). For example,

studies using various cancer cell lines have shown either cell cycle arrest or apoptosis upon treatment with SFN (20–29).

We recently identified SFN as a novel HDAC inhibitor in colon and prostate cancer cells (30, 31). HDAC inhibition was associated with global increases in histone acetylation, enhanced interactions of acetylated histones with the promoter regions of the *P21* and *BAX* genes, and elevated expression of p21<sup>Cip1/Waf1</sup> and BAX proteins. In addition to a G2/M cell cycle arrest, there was evidence for loss of BCL-2 expression and increased multicaspase activity, resulting in apoptosis.

Based on these findings (30, 31), we sought to determine whether SFN might act as an HDAC inhibitor *in vivo*, using a prostate xenograft model, as well as in peripheral blood mononuclear cells (PBMC) from human volunteers consuming broccoli sprouts. Here, we report that dietary administration of SFN, in both mice and humans, inhibited HDAC activity significantly and suppressed the growth of human PC-3 xenografts in nude mice. These findings provide evidence that SFN may exert its chemopreventive effects, at least in part, through inhibition of HDAC activity *in vivo*.

## Materials and Methods

**Xenografts.** Male athymic nude BALB/c (nu/nu) mice (5 weeks old) were purchased from Charles River (Wilmington, MA) and maintained in accordance with Institutional Animal Care and Use Committee guidelines. PC-3 cells were mixed in a 1:1 ratio of complete media (Roswell Park Memorial Institute-1640 + 10% FBS) and High Concentration Growth Factors Matrigel Matrix (Becton Dickinson, Bedford, MA). A suspension of 10<sup>6</sup> cells (50  $\mu$ l) was injected subcutaneously into the right flank of each mouse. Tumor volume was calculated using the following formula for the volume of an ellipsoid: length  $\times$  width<sup>2</sup>  $\times$  0.5236 ( $\pi/6$ ).

**Treatments.** Mice were randomized into groups of 10 animals. In the test group, SFN (LKT, St. Paul, MN) was mixed into pelleted AIN93G diet without *t*-butylhydroquinone at a concentration of 443 mg/kg (2.5  $\mu$ mol/g) diet (Research Diets, New Brunswick, NJ). AIN93G diet does not contain any isothiocyanates. Control mice received the same diet without added SFN. Diets were  $\gamma$ -irradiated at 2.5 Mrads and fed to mice *ad libitum*. Food intake and body weights were monitored throughout the study, and the animals were killed by CO<sub>2</sub> inhalation 21 days after xenograft implantation.

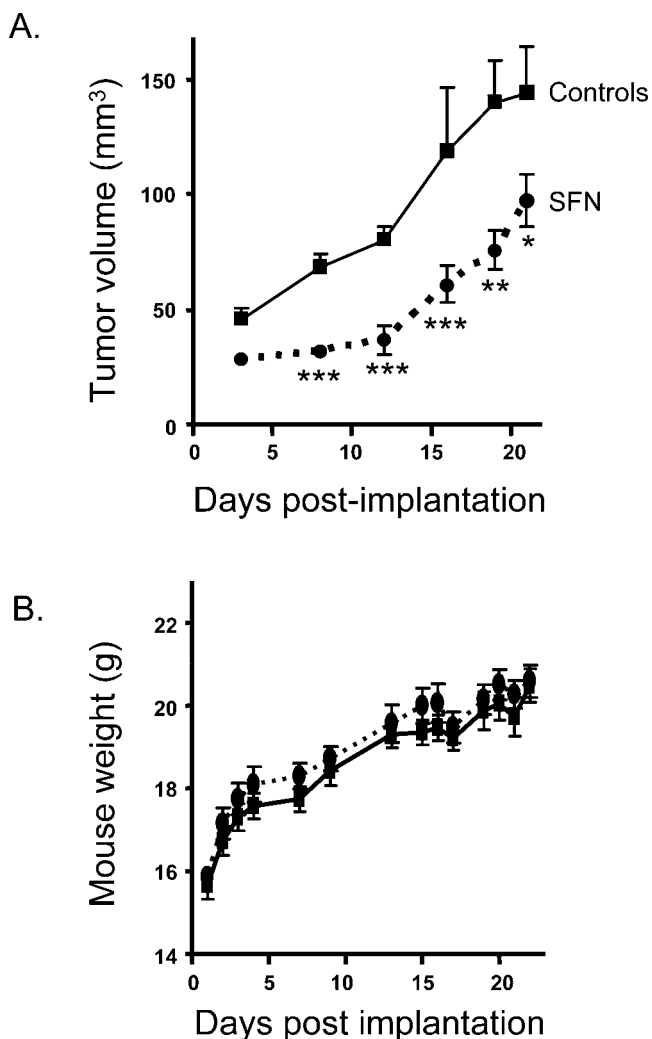
**Mononuclear Blood Cell (MBC) Isolation.** Spleens were homogenized with a Dounce homogenizer, and a single cell suspension was made by passing the homogenate through a 21-gauge needle followed by a 23-gauge needle. The cell suspension was placed on top of Fico/Lite LE (Atlanta Biologicals, Lawrenceville, GA) and centrifuged for 25 mins at 2000 *g*. The MBC layer was collected and washed in phosphate-buffered saline (PBS) by

spinning for 10 mins at 900 *g*. The MBC pellet was placed in 90% FBS/10% dimethylsulfoxide and slow-frozen in isopropyl alcohol at  $-80^{\circ}\text{C}$  overnight.

**Western Blotting.** Frozen portions of tumor tissue were thawed in lysis buffer and ground using a Potter-Elvehjem homogenizer (Teflon drill). Whole prostates were homogenized and centrifuged at 15,000 rpm for 5 mins, and the supernatant was collected. Protein concentrations were determined by the Lowry assay. Proteins (10–20  $\mu$ g) were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis on a 4%–12% bis-Tris gel (Novex, San Diego, CA) and were transferred to nitrocellulose membrane (Invitrogen, Carlsbad, CA). Equal protein loading was confirmed with Amido black staining and  $\beta$ -actin levels. The membrane was blocked for 1 hr with 2% bovine serum albumin, followed by either overnight incubation with primary antibody at 4 $^{\circ}\text{C}$  or 1 hr incubation with primary antibody at room temperature, and was finally incubated for 1 hr with secondary antibody conjugated with horseradish peroxidase (Bio-Rad, Hercules, CA). Antibody dilutions were as follows: acetylated histone H3, 1:100 (Upstate, Charlottesville, VA); acetylated histone H4, 1:100 (Upstate); Bax, 1:100 (Santa Cruz Biotechnologies, Santa Cruz, CA); and  $\beta$ -actin, 1:5000 (Sigma, St. Louis, MO). Antibodies were selected on the basis of their ability to recognize both human and mouse proteins. Detection was by Western Lightning Chemiluminescence Reagent Plus (PE Life Sciences, Boston, MA) with image analysis on an AlphaInnotech photodocumentation system (Hayward, CA). Image quantification was determined by National Institutes of Health ImageJ software (Bethesda, MD). Expression levels were normalized to  $\beta$ -actin.

**HDAC Activity Assay.** HDAC activity was determined using the Fluor-de-Lys HDAC activity assay kit (Biomol, Plymouth Meeting, PA), as reported previously (31). Homogenates (25  $\mu$ g of total protein) from tumor tissue, prostates, or MBC were incubated with Fluor-de-Lys substrate in triplicate for 12 min at 37 $^{\circ}\text{C}$  to initiate the HDAC reaction. Fluor-de-Lys Developer was then added, and the mixture was incubated for another 10 min at room temperature. Fluorescence was measured using a Spectra Max Gemini XS fluorescent plate reader (Molecular Devices, Sunnyvale, CA), with excitation at 360 nm and emission at 460 nm.

**Human Study.** The Institutional Review Board at Oregon State University approved the protocol for this investigation, and all participants provided written consent before enrollment. In brief, healthy volunteers ( $n = 3$ ) were selected for this study on the basis of age (18–55 yrs), nonnutritional supplement use ( $>6$  months), and exercise status ( $<5$  hrs/week of aerobic activity). Volunteers were asked to refrain from cruciferous vegetable intake for 48 hrs. Each subject consumed 68 g BroccoSprouts broccoli sprouts (approximately 105 mg SFN; equivalent to approximately 570 g of mature broccoli) with a bagel and cream cheese. Blood was collected at 0, 3, 6, 24, and 48 hrs after



**Figure 1.** SFN retards the growth of PC-3 xenografts. (A) PC-3 cells were implanted into male nude mice (10 mice per group). SFN was administered in the diet (443 mg/kg; 2.5  $\mu$ mol/g) beginning on the day of implantation. Tumor volume was determined as described in Materials and Methods. (B) No adverse effect of SFN on mouse body weight. Data are expressed as mean  $\pm$  SE. \* $P$  < 0.05, \*\* $P$  < 0.01, \*\*\* $P$  < 0.001.

consumption of broccoli sprouts. Approximately 8 ml of blood was drawn into EDTA vacutainer blood collection tubes. The blood was layered on top of Histopaque (Sigma) and centrifuged at 400  $g$  for 30 min at room temperature. The PBMC layer was removed, placed in a clean 15-ml tube, washed with PBS three times, and then frozen at  $-80^{\circ}\text{C}$  until HDAC activity analysis.

**Statistics.** One-way analysis of variance or Student's  $t$  test was used to assess the differences between groups. Differences among treatments were tested by Dunnett's test.

## Results

We recently identified SFN as an HDAC inhibitor in human prostate epithelial cells, with HDAC inhibition resulting in cell cycle arrest and apoptosis (30). In the present study, we sought to verify that SFN was an HDAC

**Table 1.** No Effect of SFN on Food Intake and Body Weight Gain<sup>a</sup>

	Average daily food intake (g)	Average body weight at end of study (g)
Control	3.11 $\pm$ 0.99	20.48 $\pm$ 1.22
SFN	3.26 $\pm$ 0.98	20.58 $\pm$ 1.23

<sup>a</sup> Nude mice (10 animals per group) were given  $\gamma$ -irradiated AIN93G diet (controls), or the same diet containing 443 mg SFN per kg, from the day that PC-3 cells were implanted as xenografts until the study was terminated at 3 weeks. Food and water were provided *ad libitum*. For body weights at earlier time points, see Figure 1B.

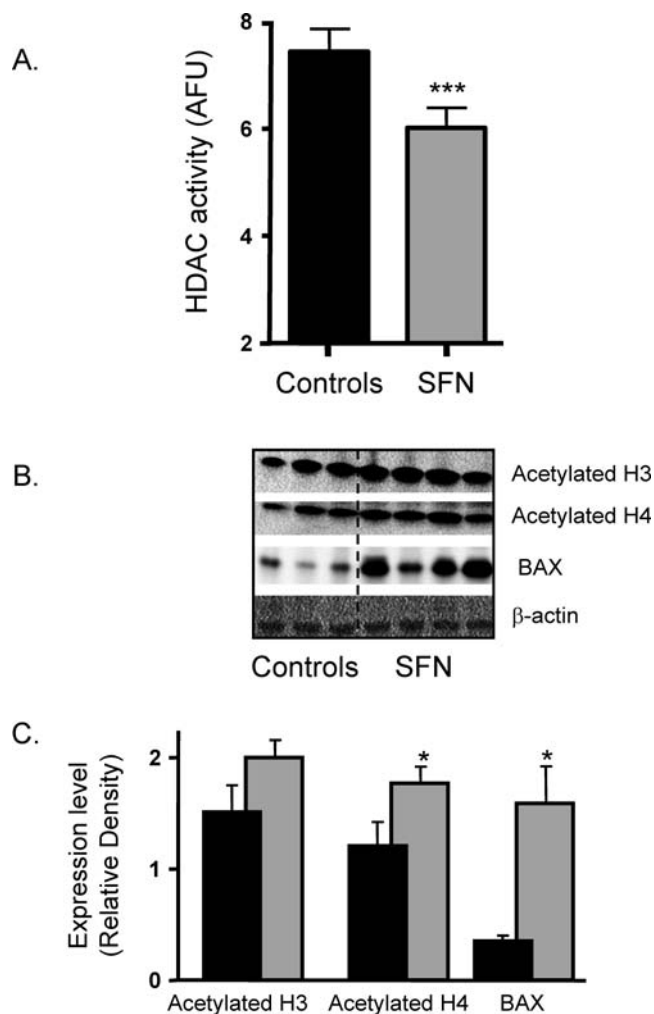
inhibitor *in vivo*, leading to retardation of PC-3 xenograft growth, and to determine whether human consumption of SFN-rich broccoli sprouts would result in inhibition of HDAC activity in PBMC. SFN administered to mice in the diet inhibited xenograft growth significantly compared with controls as early as 3 days after implantation (Fig. 1A). Although the protective effects diminished somewhat at later times, SFN retarded xenograft growth significantly throughout the 21-day duration of this study.

There were no adverse effects of SFN treatment on animal health, food intake, or body weight. The average daily food intake did not differ between controls and treatment groups (Table 1). Moreover, body weight was not significantly different throughout the duration of the study, suggesting that SFN was essentially nontoxic at the dietary concentration used in the present investigation (Fig. 1B). Mice consumed approximately 7.5  $\mu$ mol SFN (1.3 mg) per day.

At the end of the study, xenografts were examined for possible systemic effects of SFN treatment on HDAC activity. Compared with xenografts recovered from animals on the control diet, HDAC activity was inhibited significantly in xenografts of mice fed SFN ( $P$  < 0.001; Fig. 2A). Interestingly, immunoblotting of xenografts showed a trend towards increased acetylation of histones H3 and H4 among the treatment groups, compared with controls (Fig. 2B and C). These changes, however, were not statistically significant. In the PC-3 xenografts, induction of BAX expression was observed, and this was significant in animals fed SFN (Fig. 2B and C). Because PC-3 cells lack P53, the increase in BAX may be attributed to increased histone acetylation at the BAX promoter, as observed recently in BPH-1 cells treated with SFN *in vitro* (30).

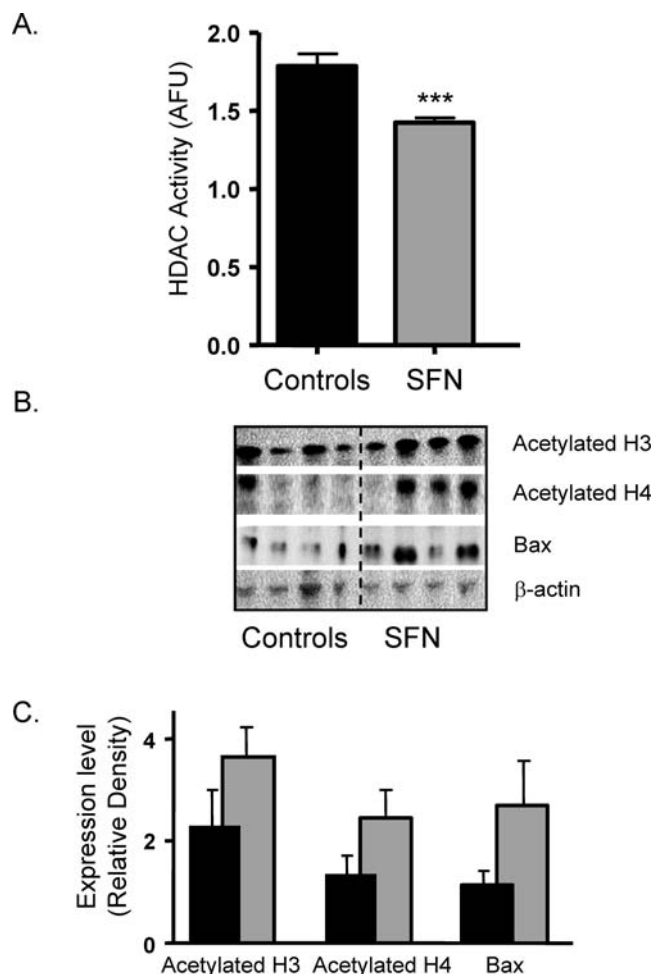
Because HDAC inhibition was detected in the xenografts, we next examined systemic effects of SFN in mouse prostate tissues. HDAC activity was significantly inhibited in the prostates from mice treated with SFN, compared with animals fed the control diet (Fig. 3A). Paralleling the inhibition of HDAC activity, prostates from the SFN-treated mice showed increased acetylated histones H3 and H4, and a concomitant increase in BAX expression (Fig. 3B and C).

Several HDAC inhibitors are currently undergoing clinical trials; an important biomarker of HDAC inhibition used in these trials is acetylated histone status from PBMC



**Figure 2.** SFN inhibits HDAC activity in PC-3 xenografts. (A) HDAC activity (arbitrary fluorescent units [AFU]) was determined in xenografts as described in Materials and Methods. Data are expressed as mean  $\pm$  SE.  $n = 10$ . \*\*\* $P < 0.001$ . (B) Xenografts were immunoblotted for acetylated H3, acetylated H4, and BAX;  $\beta$ -actin was used as the loading control. Each lane represents a xenograft from an individual animal, and the blot is a representative sample from each group of 10 animals. (C) Quantification of immunoblots using National Institutes of Health Image J, normalized for  $\beta$ -actin. Data are expressed as mean  $\pm$  SE. \* $P < 0.05$ .

(32, 33). In the present investigation, HDAC activity was inhibited significantly in MBC obtained from SFN-fed mice, compared with controls (\*\* $P < 0.01$ , Fig. 4A). Moreover, in human subjects, HDAC activity was significantly inhibited in the PBMC of all three subjects as early as 3 hrs after consumption of 68 g of broccoli sprouts (Fig. 4B). In two of the subjects, HDAC inhibition also was seen at the 6-hr time point, and by 24–48 hrs HDAC activity had returned to the level detected at 0 hrs (Fig. 4C). In a third subject, the starting HDAC activity at 0 hrs was lower than expected, and for this reason the highest HDAC activity was seen at 48 hrs. (Fig. 4C). A longer period of restriction from cruciferous vegetable intake might be required in some subjects to attain peak HDAC activity levels in BMC.

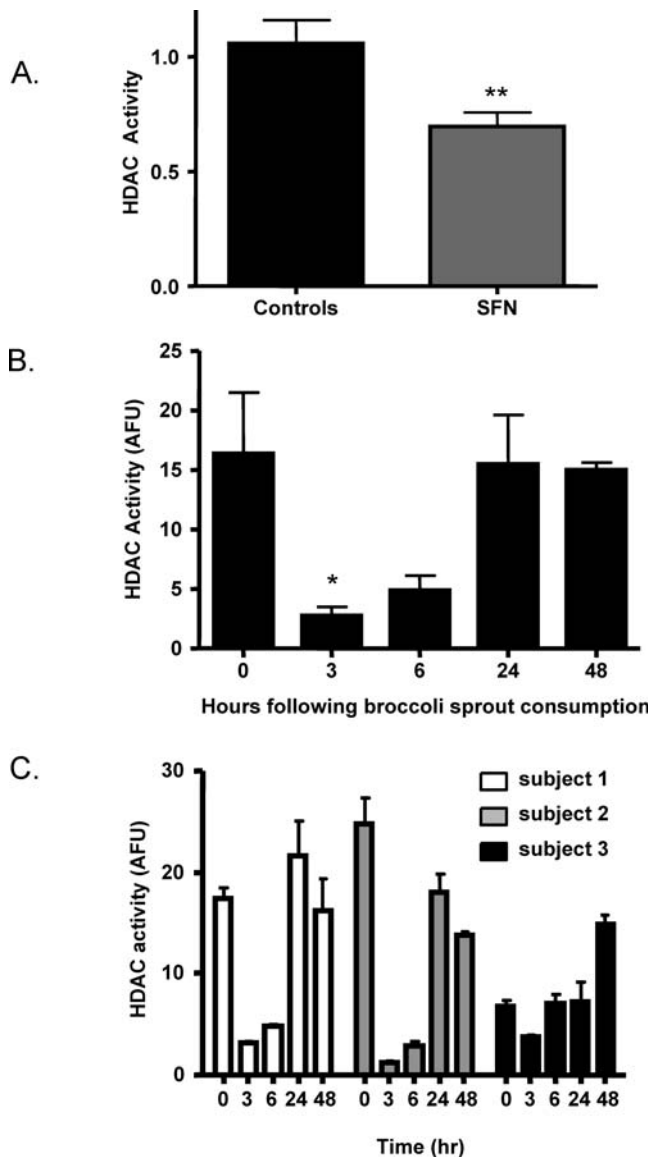


**Figure 3.** SFN increases acetylated histones and induces BAX expression in mouse prostates. (A) HDAC activity was determined in mouse prostate tissues as described in Materials and Methods. Data are expressed as mean  $\pm$  SE.  $n = 10$ . \*\*\* $P < 0.001$ . (B) Prostates were immunoblotted for acetylated H3, acetylated H4, and BAX;  $\beta$ -actin was included as a loading control. Each lane represents a prostate from an individual animal, and the blot is a representative sample from each group of 10 animals. (C) Quantification of immunoblots using National Institutes of Health Image J, normalized for  $\beta$ -actin. Data are expressed as mean  $\pm$  SE.

## Discussion

We recently reported that SFN inhibited HDAC activity in BPH-1, LnCaP, and PC-3 prostate epithelial cells, with accumulation of acetylated histones associated with the BAX promoter, increased levels of BAX mRNA and protein, loss of procaspase-3, and induction of multicaspase activity leading to apoptosis (30). Other groups have reported that SFN given to mice at a dose of 5.6  $\mu$ mol three times per week by gavage slowed the growth of PC-3 xenografts and caused induction of apoptosis (20). In the present study, we confirmed that dietary SFN also retarded the growth of PC-3 xenografts *in vivo*, with evidence for HDAC inhibition and induction of BAX in xenografts and prostate tissues of mice. Importantly, HDAC activity also was inhibited in MBC from mice given SFN, and in PBMC from human subjects





**Figure 4.** SFN inhibits HDAC activity in mouse MBC and human PBMC. (A) HDAC activity in mouse MBC was determined as described in Materials and Methods; mean  $\pm$  SE.  $**P < 0.01$  for SFN *versus* controls. (B) Human subjects consumed 68 g of BroccoSprouts and blood was obtained at the times indicated in the figure. PBMC were isolated and HDAC activity assays were performed as described in Materials and Methods. Data are expressed as mean  $\pm$  SE.  $n = 3$ .  $*P < 0.05$ . (C) Data for HDAC activity in PBMC from each subject.

consuming broccoli sprouts, which are a rich source of SFN (17–20).

SFN is an effective chemopreventive agent and induces Phase 2 enzymes involved in carcinogen detoxification and excretion (17–20). However, SFN also has been shown to suppress colonic aberrant crypts when administered solely after the carcinogen exposure period (18), suggesting a role for mechanisms postinitiation. SFN induced apoptosis in, and slowed the growth of, PC-3 xenografts *in vivo* (20), and cell culture models have demonstrated cell cycle arrest and apoptosis using SFN as a test agent (20–22, 24, 25, 27–30,

36–39). Several mechanisms by which SFN modulates cell cycle arrest and apoptosis in cancer cells have been proposed (reviewed in Refs. 2, 40). Based on the present findings and our previous work in colon and prostate cancer cells (30, 31), we propose that SFN acts as an HDAC inhibitor, augmenting histone acetylation and derepressing genes such as *P21* and *Bax*, leading to cancer prevention via cell cycle arrest and apoptosis.

Interestingly, the HDAC activity levels varied between tissue types, based on the fluorescent values that measure relative HDAC activity. For example, the arbitrary fluorescent units for average HDAC activity in the xenografts (human PC-3 prostate cancer) from control mice were approximately 8-fold higher than the arbitrary fluorescent units for average HDAC activity in the prostates in control mice. PC-3 cells are derived from an androgen-independent, late-stage human prostate carcinoma; the higher HDAC activity levels in these cells compared with normal murine prostate tissue confirms the importance of HDAC activity in the progression to tumorigenesis. Interestingly, there seemed to be a difference in the extent of accumulation of acetylated histones in the prostate compared with the PC-3 xenografts (Figs. 2B and 3B). This may be because of differences in relative HDAC activity levels before the SFN intervention, metabolism of SFN in the two sites, or the accumulation and retention of SFN in xenografts versus murine prostates. It is also possible that cancerous cells have a greater ability to resist the downstream effects of HDAC inhibition, that is, accumulation of acetylated histones, than normal cells. This would suggest that targeting epigenetic alterations early in the carcinogenesis process through prevention may be a better strategy than using HDAC inhibitors as chemotherapeutic agents in a more advanced stage of cancer. Further studies should address the efficacy of prevention versus treatment.

We also tested a prototypical HDAC inhibitor, trichostatin A (TSA), in the mouse xenograft model, starting on the day of implantation and administered by subcutaneous injection (0.5 mg/kg body weight) every day for 21 days; TSA was not as effective as SFN at inhibiting xenograft growth, and there was no evidence for additive or synergistic effects of SFN plus TSA coexposure (data not shown). TSA is a potent HDAC inhibitor *in vitro*, but it can be converted rapidly to inactive metabolites *in vivo* (41). In previous studies, an inhibitor of the mercapturic acid pathway attenuated the HDAC inhibitory effects of SFN *in vitro*, but increased the effectiveness of TSA (31), indicating that TSA parent compound is required for HDAC inhibition. Further work is needed on the possible synergistic effects of TSA plus SFN *in vivo*.

Phase 1 clinical trials have been completed for HDAC inhibitors such as SAHA and depsipeptide in which the test agents were administered by continuous intravenous (iv) infusion for 2 hrs (32, 33), and acetylated histone H3 levels in PBMC were used as a biomarker of effectiveness (32, 33). In the present investigation, SFN was given in the diet

for 3 weeks and HDAC inhibition was observed in MBC and systemic tissues at the end of the study, hinting at a possible sustained effect of SFN compared with pharmacologic HDAC inhibitors given *via* the iv route of administration. However, we noted that the inhibition of xenograft development by SFN was most obvious early after implantation (Fig. 1A). Compensatory HDAC or histone acetyltransferase (HAT) expression might reduce the efficacy of SFN over time, but it will be necessary to confirm this possibility in follow-up studies. Such a finding could have important implications for human clinical trials in individuals who might develop resistance to HDAC inhibitors administered for prolonged periods. Cell culture studies are currently in progress to further understand the kinetics of HDAC inhibition by SFN, in order to determine whether there are changes in HDAC protein levels over time in response to SFN and to characterize potential HDAC subtypes that may be more sensitive to inhibition by SFN.

The xenograft model is useful for testing potential chemopreventive and chemotherapeutic agents *in vivo*, but we considered it important to examine biological effects in the intended target tissue. The accumulation of acetylated histones in the prostate of SFN-treated mice and the activation of BAX suggested that SFN indeed entered the prostate in a bioactive form and attained concentrations necessary for HDAC inhibition. HDAC activity assays confirmed the inhibition by SFN in prostate and MBC of mice. We therefore conducted a pilot study with human volunteers to determine whether consumption of broccoli sprouts would inhibit HDAC activity in human PBMC. Based on the approximate SFN content in BroccoSprouts, we calculated that 68 g of broccoli sprouts (approximately 1 cup) would provide a dose of SFN in humans comparable to that which inhibited HDAC activity in colonic mucosa, prostate, and PBMC of mice (42). During the period 3–6 hrs after ingestion, broccoli sprouts strongly inhibited HDAC activity in human PBMC (Fig. 4B and C). To our knowledge, this is the first study to show that a naturally-consumed food product, namely broccoli sprouts, exerted such a marked effect on HDAC activity in humans. A recent study reported the pharmacokinetics of SFN in humans and suggested that after consumption of broccoli containing high levels of SFN, some SFN or its metabolites may be retained in various tissues rather than rapidly excreted (43). Further work will be necessary to characterize the concentration of SFN metabolites in human plasma and urine and the concordance with HDAC activity, acetylated histones, and other acetylated proteins after single or repeated consumption of broccoli sprouts.

Clinical trials with pharmacologic HDAC inhibitors utilize the degree of histone acetylation as a biomarker for inhibition of HDAC activity (32, 33). In this small preliminary pilot study, we chose to use a more direct measure of HDAC activity through the use of an HDAC activity assay. In the results reported here from the xenograft study and in our previous studies (30, 31, 42), we have found

that a decrease in HDAC activity correlates with an increase in accumulation of acetylated histones. Although clinical trials have focused on alterations in acetylated histone status as a biomarker for HDAC inhibition, the use of an HDAC activity assay requires less sample processing and the potential for high throughput analysis. Further studies are underway to validate this assay and confirm similar alterations in acetylated histone status in human volunteers consuming broccoli sprouts. In addition to acetylation of histones, HDACs also target various nonhistone proteins for deacetylation, such as p53,  $\alpha$ -tubulin, and estrogen receptor  $\alpha$  (44), ultimately affecting the activity of the proteins. Because the alterations in the activities of these proteins play a role in cell cycle progression and apoptosis, it will be important to examine the acetylation status of nonhistone proteins in future studies.

Importantly, this study found that administration of both purified SFN and SFN from broccoli sprouts resulted in HDAC inhibition. Because of the cost of purified SFN, the use of SFN-rich foods, such as broccoli sprouts, will greatly increase the accessibility of this important chemopreventive agent. The purified SFN used in this study was a racemic mixture of two stereoisomers, whereas SFN found in nature is only the L enantiomer. In cell culture, there was no difference in HDAC inhibition by the purified D or L enantiomers or by a 50:50 mixture (data not shown), suggesting that the inhibition of HDAC activity by SFN is not dependent on the specific isomer. Crystallization studies with SFN and human HDAC proteins would confirm this observation.

In summary, the present investigation showed that dietary administration of SFN retarded the growth of human PC-3 prostate cancer cells in nude mice. SFN inhibited HDAC activity in the xenografts, prostates, and MBC, and there was a trend towards increased histone acetylation. Consumption of a SFN-rich food (broccoli sprouts) strongly inhibited HDAC activity in MBC of human subjects. We conclude that further studies are warranted on dietary HDAC inhibitors and prostate cancer prevention, and on the use of HDAC activity as a biomarker in clinical trials. When combined with pharmacologic HDAC inhibitors or other chemopreventive agents, dietary HDAC inhibitors might improve the efficacy of intervention strategies and reduce the effective dose used in more conventional therapeutic treatments, thereby ameliorating untoward side effects.

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