## Changes in Renal Phosphate Reabsorption in the Aged Rat (44268)

SUSAN E. MULRONEY, 1 CRAIG WODA, AND AVIAD HARAMATI

Department of Physiology and Biophysics, Georgetown University School of Medicine, Washington, DC 20007

Abstract. Depletion of inorganic phosphate (P<sub>i</sub>) reserves occurs frequently in aged animals and can result in diminished bone mineralization and osteoporosis. This altered P, balance results from a reduction in intestinal P, absorption and an elevation in renal P<sub>i</sub> excretion. Since the kidney plays a central role in maintaining P<sub>i</sub> homeostasis, we tested whether the increased phosphaturia seen with aging is a consequence of changes in the intrinsic tubular capacity to reabsorb P, (TmP<sub>i</sub>). Male Wistar rats (12-, 18-, and 24-months-old) were acutely thyroparathyroidectomized (TPTX) and prepared for renal clearance studies in the presence and absence of fixed levels of parathyroid hormone (synthetic PTH-(1-34), 1U/kg/min). The maximum capacity for Pi transport (TmP<sub>i</sub>) was assessed by infusion of P<sub>i</sub> at progressively higher rates (0-6 µmol/min) to increase the filtered load of Pi and facilitate the determination of the TmP<sub>i</sub>. TmP<sub>i</sub> declined significantly with age  $(3.51 \pm 0.12 \text{ vs } 3.04 \pm 0.19 \text{ vs } 2.30 \pm 0.18$  $\mu$ mol/ml, for 12-, 18-, and 24-month-old rats, respectively, P < 0.05) in TPTX rats. Administration of PTH markedly reduced the TmP, in all age groups. Although the TmP, attained was similar among the age groups (1.15  $\pm$  0.13 vs 1.15  $\pm$  0.06 vs 1.03  $\pm$ 0.09 µmol/ml, for 12-, 18-, and 24-month-old rats, respectively), the magnitude of the reduction in the presence of PTH declined from 67% in 12-month-old rats to 62% and 55% in 18- and 24-month-old rats, respectively. These results demonstrate that aging is associated with a PTH-independent decrease in the intrinsic capacity of the kidney to reabsorb phosphate. Further, the kidney of the aged rat can respond to a pharmacological dose of PTH with appropriate reductions in the TmP, although the magnitude of the response declines with age. [P.S.E.B.M. 1998, Vol 218]

ging is associated with alterations in phosphate  $(P_i)$  homeostasis. Serum  $P_i$  concentrations are decreased and urinary excretion rates of  $P_i$  are elevated in the aged rat (25-month-old) (1, 2), compared with the young (3, 4). The elevated urinary  $P_i$  excretion together with reduced intestinal  $P_i$  absorption (5) results in a negative  $P_i$  balance in the older animal. Since inorganic  $P_i$  is a main constituent of the bone matrix and is required for proper bone mineralization, depletion of  $P_i$  in the aging animal can disrupt bone homeostasis and may promote and/or accelerate the onset of osteoporosis. Circulating hormones known to influence the

extracellular levels of  $P_i$  have been shown to change with age (1, 6-8). For example, immunoreactive parathyroid hormone (iPTH) is elevated in the senescent animal (1, 8), whereas circulating levels of 1,25-dihydroxyvitamin  $D_3$  are significantly lower (1, 6) compared to the immature, developing animal. Advancing age has also been shown to cause a decrease in both the pituitary secretion of growth hormone (GH) and circulating serum levels of insulin-like growth factor-I (IGF-I) (9), which are known to enhance the tubular reabsorption of  $P_i$ .

Aging also affects various morphological, hemodynamic and functional aspects of the kidney. By age 70 in humans, approximately 20% of total kidney size, primarily the cortical region, is lost (10), and by age 80, renal blood flow decreases by approximately 40% (11). Similarly, a progressive decline in proximal tubular length and volume has been noted with aging, as well as a thickening of the Bowman's capsule basement membrane (12) and a significant decline (30%–50%) in glomerular number (13). Anderson *et al.* have shown that glomerular damage initiates atrophy of both afferent and efferent arterioles leading to glomerular sclerosis and decreased glomerular filtration by

Received June 25, 1997. [P.S.E.B.M. 1998, Vol 218] Accepted December 3, 1997.

0037-9727/98/2181-0062\$10.50/0 Copyright © 1998 by the Society for Experimental Biology and Medicine

<sup>&</sup>lt;sup>1</sup> To whom requests for reprints should be addressed at Department of Physiology and Biophysics, Georgetown University School of Medicine, Rm. 253 Basic Science Building, 3900 Reservoir Rd, NW, Washington, DC 20007. This work was supported by grant DK36111 from the NIH.

the kidney (14). In adult humans, there is an accelerated rate of nephron loss between the ages of 40 and 80 with a 50% nephron loss by 75 years of age (15). These age-related renal changes have been shown to result in alterations in the tubular solute transport of the remaining nephrons. However, to what extent aging alters the intrinsic tubular capacity to reabsorb  $P_i$  (TmP<sub>i</sub>) is not well known and is the focus of the present study.

## Materials and Methods

Renal clearance studies were performed on male Wistar rats (provided by the Gerontology Research Center, NIH, Baltimore, MD) at 12-, 18-, and 24-months-of-age. The rats were maintained on a normal rat chow diet (Ralston Purina, St. Louis, MO) containing 0.86% phosphate and 1.2% calcium, and food and water were given *ad libitum*. Room temperature was held between 22°–24°C, and rats were provided with a 12-hr-on/12-hr-off light cycle.

On the day of the experiment, the rats were anesthetized by an intraperitoneal injection of Inactin (80-100 mg/kg) (Promonta, Hamburg, FRG) and prepared for acute renal clearance studies. Body temperature was maintained at  $37^{\circ} \pm 0.5^{\circ}$ C via a servo-controlled heat lamp (Yellow Springs Instruments, Yellow Springs, OH) and a thermoregulated table and monitored by a rectal probe. A tracheostomy was performed so that the animals could breathe spontaneously. Catheters were inserted into the jugular vein for infusing solutions, the carotid artery for blood pressure measurement and blood collections, and the bladder for urine collections.

To remove endogenous levels of circulating parathyroid hormone, all rats underwent a thyroparathyroidectomy (TPTX) on the day of the experiment. An FEP<sub>i</sub> of  $<1-2\pm1\%$  during the control collection period was considered a successful TPTX in the aged animals. Following TPTX, a 2% solution of inulin was infused at a rate of 2% body wt/hr for 90 min to allow the animal to recover and attain a steady state.

**Experiments in the Absence of Parathyroid Hormone.** Following the 90-min recovery period, a 30-min control urine clearance was collected. Increasing concentrations of  $P_i$  (1, 2, 3, and 6  $\mu$ mol/min) were added to the inulin infusion to increase the filtered load of  $P_i$  and facilitate the determination of the Tm $P_i$ .  $P_i$  infusions proceeded for 20 min before sequential 30-min clearance periods began. Blood samples were obtained at the midpoint of each clearance period.

**Experiments in the Presence of Parathyroid Hormone.** In this series, the rats were prepared as previously stated, with the exception that exogenous PTH was administered throughout the experiment. PTH (1-34, Beckman Instruments, Palo Alto, CA) was given in a bolus injection (33 U/kg) iv, and followed with a sustained infusion of 1 U/kg/min for the duration of the experiment.

**Analysis.** Inulin concentrations in plasma and urine were determined by the anthrone method (16), and GFR was

equated with the clearance of inulin.  $P_i$  concentrations were measured by the phosphomolybdate method (17). Reabsorbed  $P_i$  ( $R_{Pi}$ ) was calculated as the difference between the amount of  $P_i$  filtered ( $FL_{Pi} = P_{Pi} \times GFR$ ), and the amount of  $P_i$  excreted ( $U_{Pi}V$ ). The maximum capacity of  $P_i$  reabsorption normalized by the GFR ( $TmP_i$ ) was calculated as the mean of the highest values of  $R_{Pi}/GFR$  for each animal within a group. All values are expressed as means  $\pm$  SE. A one-way analysis of variance followed by Scheffe tests were used for comparisons between groups. Paired Student t tests were used to make statistical comparisons within groups.

## Results

**Experiments in the Absence of Parathyroid Hormone.** Mean arterial blood pressure (MAP) was not significantly different between groups  $(136 \pm 4 \text{ vs } 130 \pm 6 \text{ vs } 127 \pm 4 \text{ mmHg}$  for 12-, 18-, and 24-month-old rats, respectively) and was maintained throughout the experiment. Table I provides indices of renal function in TPTX animals in the absence of parathyroid hormone. From 12- to 18-months-of-age, there were significant (P < 0.05) decreases in the basal plasma  $P_i$  concentration  $(2.22 \pm 0.24 \text{ vs } 1.26 \pm 0.13 \text{ mM}$  for 12- and 18-month-old rats, respectively) and GFR  $(3.33 \pm 0.37 \text{ vs } 1.48 \pm 0.21 \text{ ml/min}$  for 12- and 18-month-old rats, respectively). Moreover, while the plasma  $P_i$  concentration was stable between 18 and 24 months, aging was associated with a progressive decline in GFR.

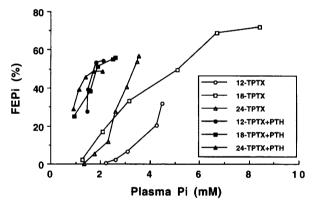
Infusions of P<sub>i</sub> caused a progressive elevation of the plasma P<sub>i</sub> levels that increased the filtered load of P<sub>i</sub>, although the GFR remained relatively constant within each age group. The increase in filtered load of Pi caused an elevation in the reabsorption of P<sub>i</sub>, but also in the fractional excretion of P<sub>i</sub> (FE<sub>Pi</sub>). Figure 1 depicts this relationship, showing the progressive rise in the fractional excretion of P<sub>i</sub> when the plasma P<sub>i</sub> concentrations increased. The curve shifted further to the left as age increased indicating an age-related increase in sensitivity to the phosphaturic effect of P<sub>i</sub> infusion. This change is likely explained by an agerelated reduction in tubular P<sub>i</sub> reabsorption. As shown in Figure 2, the TmP<sub>i</sub> was significantly (P < 0.05) lower in the older animals  $(3.04 \pm 0.19 \text{ and } 2.30 \pm 0.18 \mu \text{mol/ml})$  for 18and 24-month-old rats, respectively) compared to the 12month-old adult rats (3.51  $\pm$  0.12  $\mu$ mol/ml). The further decrease in TmP<sub>i</sub> between 18- and 24-month-old rats was not due to a decrease in food intake  $(12.0 \pm 1.3 \text{ vs } 11.5 \pm 1.4 \text{ m})$ g/day, respectively, n.s.). Thus, their daily intake of P<sub>i</sub> was comparable (~2.5 mmol/day), and in general, should have provided an adequate supply of P<sub>i</sub> for the animals. Thus, the intrinsic ability of the kidney to reabsorb P<sub>i</sub> (in the absence of PTH) declines precipitously with age.

Experiments in the Presence of Parathyroid Hormone. MAP was no different between age groups  $(146 \pm 5 \text{ vs } 133 \pm 4 \text{ vs } 152 \pm 10 \text{ mmHg for } 12\text{-}, 18\text{-}, \text{ and } 24\text{-month-old rats, respectively})$  and maintained throughout the experiment. The effects of a pharmacologic dose of PTH

Table I. Effect of Phosphate Infusion in Acutely TPTX Aging Rats

Age	n	P <sub>i</sub> Infused µmol/min	P <sub>Pi</sub> m <i>M</i>	GFR ml/min	FL <sub>Pi</sub> µmol/min	FE <sub>Pi</sub> %	R <sub>Pi</sub> µmol/ml	R <sub>ei</sub> /GFR ml/min
12 month	(5)	0	2.22 ± 0.24	$3.33 \pm 0.37$	7.08 ± 0.91	$0.4 \pm 0.3$	7.04 ± 0.90	2.21 ± 0.24
		1	$2.21 \pm 0.13$	$3.56 \pm 0.19$	$7.84 \pm 0.67$	$0.5 \pm 0.4$	$7.81 \pm 0.66$	$2.20 \pm 0.13$
		2	$2.62 \pm 0.23$	$3.05 \pm 0.29$	8.01 ± 1.22	$2.3 \pm 1.7$	$7.83 \pm 1.33$	$2.55 \pm 0.23$
		3	$3.09 \pm 0.28$	$3.56 \pm 0.16$	11.12 ± 0.97	$6.5 \pm 1.6$	$10.60 \pm 1.07$	$2.92 \pm 0.31$
		6	$4.26 \pm 0.23$	$3.83 \pm 0.34$	16.18 ± 0.91	$20.4 \pm 1.7$	$13.51 \pm 0.89$	$3.52 \pm 0.14$
		6	$4.49 \pm 0.27$	$3.10 \pm 0.36$	$14.24 \pm 0.87$	$31.8 \pm 2.1$	$10.50 \pm 1.33$	$3.22 \pm 0.31$
18 month	(6)	0	$1.26 \pm 0.13$	$1.48 \pm 0.21$	$1.82 \pm 0.25$	$2.2 \pm 1.0$	$1.78 \pm 0.25$	$1.24 \pm 0.14$
	` ,	1	$2.08 \pm 0.11$	$2.17 \pm 0.52$	$4.50 \pm 1.19$	$16.7 \pm 4.7$	$3.81 \pm 1.04$	$1.75 \pm 0.15$
		2	$3.17 \pm 0.28$	$1.97 \pm 0.43$	$6.10 \pm 1.37$	$33.1 \pm 4.7$	$4.04 \pm 0.92$	$2.24 \pm 0.28$
		3	$5.12 \pm 0.19$	$1.89 \pm 0.29$	$9.42 \pm 1.23$	$49.3 \pm 3.5$	$4.89 \pm 0.81$	$2.58 \pm 0.16$
		6	$6.72 \pm 0.42$	$1.80 \pm 0.38$	$12.02 \pm 2.73$	$68.9 \pm 7.5$	4.42 ± 1.76	$2.52 \pm 0.41$
		6	$8.44 \pm 0.84$	1.71 ± 0.31	$13.40 \pm 1.79$	$72.0 \pm 6.8$	$3.84 \pm 1.32$	$2.39 \pm 0.42$
24 month	(6)	0	$1.34 \pm 0.23$	$1.08 \pm 0.19$	$1.35 \pm 0.20$	$0.2 \pm 0.0$	$1.35 \pm 0.20$	$1.33 \pm 0.23$
	` '	3	1.77 ± 0.14	$1.48 \pm 0.19$	$2.59 \pm 0.31$	$5.0 \pm 2.2$	$2.44 \pm 0.26$	$1.68 \pm 0.13$
		3	$2.31 \pm 0.08$	$1.23 \pm 0.27$	$2.78 \pm 0.56$	$11.5 \pm 2.5$	$2.44 \pm 0.49$	$2.05 \pm 0.08$
		3	$2.62 \pm 0.12$	$1.62 \pm 0.21$	$4.16 \pm 0.42$	$27.6 \pm 5.3$	$3.03 \pm 0.38$	$1.88 \pm 0.11$
		6	$3.05 \pm 0.18$	1.51 ± 0.15	$4.56 \pm 0.36$	$40.5 \pm 3.7$	$2.68 \pm 0.20$	$1.80 \pm 0.07$
		6	$3.48 \pm 0.15$	$1.63 \pm 0.21$	$5.67 \pm 0.75$	$53.6 \pm 5.7$	$2.50 \pm 0.30$	$1.59 \pm 0.17$
		6	$3.55 \pm 0.19$	$1.94 \pm 0.16$	$6.85 \pm 0.62$	$56.9 \pm 3.4$	$2.94 \pm 0.35$	$1.53 \pm 0.15$

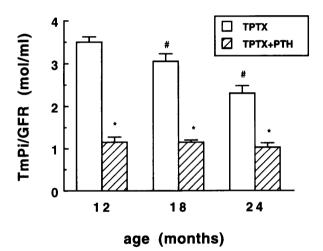
*Note.* Values are means  $\pm$  SE; n, no. of animals in each group.  $P_i$ , phosphate;  $P_{P_i}$ , plasma phosphate concentration;  $FE_{P_i}$ , fractional excretion of phosphate;  $FL_{P_i}$ , filtered load of phosphate;  $R_{P_i}$ , absolute phosphate reabsorption; GFR, glomerular filtration rate; TPTX, thyroparathyroid-ectomized. Statistical comparisons are discussed in text.



**Figure 1.** Relationship between fractional excretion of phosphate (FEP<sub>i</sub>) and plasma phosphate concentration (Plasma P<sub>i</sub>) as a function of age. Data were obtained in thyroparathyroidectomized (TPTX) rats with and without infusion of parathyroid hormone (PTH).

on basal renal function in different aged animals is shown in Table II. In the presence of PTH, basal plasma  $P_i$  levels were significantly lower in all age groups when compared to TPTX-only rats  $(1.50 \pm 0.13$  and  $0.96 \pm 0.08$  and  $0.92 \pm 0.06$  mM vs  $2.22 \pm 0.24$  and  $1.26 \pm 0.13$  and  $1.34 \pm 0.23$  mM in 12-, 18-, and 24-month-old rats, respectively, P < 0.05). Further, a significant (P < 0.05) age-related decline was evident in both basal plasma  $P_i$  concentration and GFR from 12- to 18-months-of-age although there were no differences between the senescent groups (18- vs 24-months-of-age).

Infusion of  $P_i$ , in the presence of PTH, resulted in smaller increases in plasma  $P_i$  concentrations and filtered load of  $P_i$  (Table II), compared to  $P_i$  infusions in the absence of PTH (Table I). This was due to the much higher basal rates of urinary  $P_i$  excretion seen in the presence of PTH.



**Figure 2.** Age-related changes in the maximum capacity of phosphate reabsorption (TmP<sub>i</sub>/GFR) in thyroparathyroidectomized (TPTX) rats and in TPTX rats infused with parathyroid hormone TPTX + PTH). Values were determined by averaging highest  $R_{\rm Pl}/GFR$  in each animal within an age group. Statistical comparisons are as follows: TPTX 12-month > 18-month > 24-month, P < 0.05. No significant difference (P > 0.05) was seen between any age group in the presence of PTH. \* = means significantly different from TPTX-only aged matched males, # = means significantly different from the 12-month-old age group.

However, the phosphaturic response to  $P_i$  infusion in the presence of PTH was enhanced in all groups (Figure 1). Although  $FE_{P_i}$ , in the presence of PTH, was significantly (P < 0.05) higher in all age groups compared to TPTX-only group, there was no age-related differences. The increased  $P_i$  excretion was associated with a marked decrease in  $TmP_i$  (Fig. 2). In fact, PTH reduced the  $TmP_i$  to similar levels between the different age groups  $(1.15 \pm 0.13 \text{ vs } 1.15 \pm 0.06 \text{ vs } 1.03 \pm 0.09 \text{ } \mu \text{mol/ml}$ , for 12-, 18- and 24-month-old rats,

Table II. Effect of Phosphate Infusions in Acutely TPTX Aging Rats in the Presence of PTH

Age	n	P <sub>i</sub> Infused µmol/min	P <sub>Pi</sub> m <i>M</i>	GFR ml/min	FL <sub>Pi</sub> µmol/min	FE <sub>Pi</sub> %	R <sub>Pi</sub> µmol/ml	R <sub>Pi</sub> /GFR µmol/min
12 month	(5)	0	1.50 ± 0.13	3.90 ± 0.31	5.82 ± 0.75	27.6 ± 3.6	4.25 ± 0.04	$1.90 \pm 0.13$
	, ,	1	1.48 ± 0.11	$3.83 \pm 0.51$	$5.61 \pm 0.82$	$39.0 \pm 4.8$	$3.56 \pm 0.06$	$0.93 \pm 0.12$
		2	$1.83 \pm 0.11$	$2.91 \pm 0.43$	$5.03 \pm 0.61$	$53.4 \pm 3.5$	$2.53 \pm 0.04$	$0.87 \pm 0.09$
		3	$2.13 \pm 0.16$	$3.07 \pm 0.79$	6.11 ± 1.51	$54.2 \pm 3.5$	$2.98 \pm 0.06$	$0.97 \pm 0.08$
18 month	(6)	0	$0.96 \pm 0.08$	1.56 ± 0.24	$1.49 \pm 0.23$	$24.8 \pm 5.3$	$1.17 \pm 0.23$	$0.74 \pm 0.08$
		1	$1.58 \pm 0.15$	$1.83 \pm 0.16$	$2.87 \pm 0.40$	$38.3 \pm 6.2$	$1.74 \pm 0.29$	$0.94 \pm 0.11$
		2	$1.90 \pm 0.24$	$2.44 \pm 0.29$	$4.71 \pm 0.87$	$51.2 \pm 7.2$	$2.22 \pm 0.50$	$0.88 \pm 0.13$
		3	$2.52 \pm 0.22$	$1.70 \pm 0.26$	$4.18 \pm 0.53$	$54.9 \pm 7.2$	$1.41 \pm 0.05$	$0.93 \pm 0.15$
		3	$2.61 \pm 0.25$	$2.04 \pm 0.12$	$5.24 \pm 0.38$	$55.8 \pm 9.1$	$1.59 \pm 0.16$	$0.88 \pm 0.12$
24 month	96)	0	$0.92 \pm 0.06$	$1.74 \pm 0.18$	$1.57 \pm 0.13$	$28.9 \pm 8.1$	$1.12 \pm 0.17$	$0.66 \pm 0.09$
		1	$1.14 \pm 0.08$	$2.27 \pm 0.26$	$2.59 \pm 0.38$	$39.1 \pm 3.3$	$1.58 \pm 0.25$	$0.69 \pm 0.06$
		2	$1.40 \pm 0.09$	$2.66 \pm 0.18$	$3.71 \pm 0.29$	$45.8 \pm 4.3$	$2.02 \pm 0.25$	$0.77 \pm 0.09$
		3	$1.74 \pm 0.13$	$2.10 \pm 0.27$	$3.66 \pm 0.48$	$48.5 \pm 4.7$	$1.88 \pm 0.34$	$0.92 \pm 0.14$
		3	$2.08 \pm 0.10$	$1.76 \pm 0.32$	$3.71 \pm 0.78$	$48.6 \pm 5.8$	$1.94 \pm 0.56$	$0.94 \pm 0.10$

Note. Values are means ± SE; n, no. of animals in each group. Abbreviations are as described in Table I.

respectively) although the magnitude of the reduction, in the presence of PTH, declined from 67% in 12-month-old rats to 62% and 55% in 18- and 24-month-old rats, respectively.

## Discussion

The present study demonstrates a progressive PTH-independent, age-related decline in the intrinsic capacity of the renal tubule to reabsorb P<sub>i</sub>. Further, despite the lower TmP<sub>i</sub>, the aged kidney still responds to PTH with a marked reduction in TmP<sub>i</sub>, although the magnitude of the response declines with age. These findings, together with the known elevations in circulating PTH levels in the aged animal, may explain the hypophosphatemia observed in senescence.

There are several potential factors that may contribute to the changes observed in whole kidney P<sub>i</sub> reabsorption in aging animals. Aging is associated not only with nephron loss primarily from the juxtamedullary region (12), but also with changes in proximal tubular cells, such as decreased mitochondria number (18) and reduced basolateral Na<sup>+</sup>-K<sup>+</sup> activated ATPase activity (19). Since P<sub>i</sub> uptake utilizes the Na<sup>+</sup> gradient generated by the basolateral Na<sup>+</sup>/K<sup>+</sup> ATPase pump, a reduction in the activity of this pump would limit the ability of the proximal tubule to reabsorb Pi. Previous studies have also demonstrated that the maximum capacity for P<sub>i</sub> reabsorption is greater in proximal tubules from deep as compared to superficial nephrons (20). Therefore, the combination of a selective juxtamedullary nephron loss, along with a reduction in the ability of the remaining proximal tubular cells to generate a sodium gradient, may contribute, in part, to the diminished ability of the aged kidney to reclaim the filtered load of P<sub>i</sub>.

Sodium-dependent phosphate (Na- $P_i$ ) transport on the luminal surface of the proximal tubule is the rate limiting step by which  $P_i$  is reabsorbed. Using renal brush-border membrane vesicle (BBMV) preparations, Kiebzak and Sacktor reported an age-dependent decline in the  $V_{max}$  of  $P_i$  transporters without a decrease in the affinity ( $K_m$ ) for  $P_i$ . This suggested that aging is associated with either a de-

crease in the number of P<sub>i</sub> carriers or a decrease in the transporter turnover rate in the proximal tubule (21). Recently, Sorribas *et al.* have determined that both NaP<sub>i</sub>-2 mRNA level and protein abundance is decreased in the BBM of proximal tubules from aged (12- to 16-month-old) rats compared to young adult (3- to 4-month-old) rats (22). Whether changes are present in other nephron segments is still unknown.

Alterations in the chemical composition of the lipid bilayer have also been shown to influence the function of both passive and carrier-mediated transport systems located on the cell membrane (23-25). Several laboratories have demonstrated that there is an age-related increase in the cholesterol, sphingomyelin, and phosphatidylinositol content of brush border membranes causing a decrease in membrane fluidity (26-28), which may affect Na-P<sub>i</sub> co-transport from the proximal tubule (29-32). Molitoris et al. report that the significant increase in the V<sub>max</sub> of the Na-P<sub>i</sub> cotransport, during dietary P<sub>i</sub> restriction, is due to a reduction in BBM cholesterol content and increased fluidity (32). More recently, using 24-month-old Fischer 344 rats, the elevated phosphaturia was associated with increases in cortical BBM cholesterol and sphingomyelin content and decreased BBM fluidity (26). Hence, a decline in the number of BBM Na-P; transporters, in addition to an age-related increase in membrane stiffness, may be an important factor contributing to the lower renal reabsorption of P<sub>i</sub> in aging.

In this study, the age-related decline in  $TmP_i$  was found to be independent of PTH. Several humoral factors have been shown to affect the intrinsic renal  $P_i$  reabsorption and may contribute to the age-related reduction in  $TmP_i$ . The GH/IGF-I axis is known to affect the reabsorption of  $P_i$  in a positive fashion, since chronic treatment with GH enhances tubular reabsorption of  $P_i$  (33, 34), and  $TmP_i$  is significantly reduced in hypophysectomized adult rats (35). The primary effects of growth hormone on renal  $P_i$  retention are thought to be mediated through hepatic IGF-I (36). Since circulating levels of both GH and IGF-I decline with age (9, 37), this

could potentially contribute to the increased phosphaturia seen with aging.

In the present study, although the aged rat responded to PTH with an elevation in P<sub>i</sub> excretion and an appropriate reduction in TmP<sub>i</sub>, the magnitude of the response was less compared to younger adult rats. The decline in the PTH response with age may be due to the increased circulating levels of PTH, perhaps as a consequence of reduced renal clearance of PTH (38, 39), which may lead to downregulation of renal PTH receptors. Marcus and Gonzales reported that PTH-stimulated cAMP production is attenuated in cortical renal slices from 12-month-old rats compared to slices from 2-month-old rats (40), whereas Armbrecht et al. demonstrated a decline in the PTH-induced 1,25-(OH)<sub>2</sub>D<sub>3</sub> synthesis in 13-month-old rats compared to 2-month-old rats (41). These findings suggest that the aged kidney may possess a degree of end-organ insensitivity toward the phosphaturic and vitamin D<sub>3</sub>-stimulating effects of PTH. On the other hand, the lower magnitude of response may reflect the reduced GFR and plasma P<sub>i</sub> concentration, and the already higher rate of urinary P<sub>i</sub> excretion. In the absence of a dose-response study, it is not possible to determine changes in the sensitivity of the renal tubule to PTH.

In conclusion, the present study demonstrates a significant age-related decrease in the tubular capacity to reclaim the filtered load of  $P_i$  that is independent of the effects of PTH. In addition, it appears that the aging kidney can still respond to the phosphaturic effect of PTH, although there may be some degree of insensitivity with aging. Interestingly, the decline in  $TmP_i$  seen throughout senescence is consistent with that seen previously, as the highest capacity to reabsorb phosphate occurred in the immature (3- to 4-week-old) rat and progressively declined toward adulthood (52-week-old rat) (4).

The authors gratefully acknowledge the guidance and participation of Dr. Bertram Sacktor who passed away before the study was completed.

- Armbrecht HJ, Forte LR, Halloran BP. Effect of age and dietary calcium on renal 25(OH)D metabolism, serum 1,25(OH)<sub>2</sub>D, and PTH. Am J Physiol 246:F266-F270, 1984.
- Lee DBN, Yaragawa N, Jo O, Yu BP, Beck N. Phosphaturia of aging: Studies on mechanisms. Adv Exp Med Biol 178:103–108, 1984.
- Caverzasio J, Bonjour JP, Fleisch H. Tubular handling of P<sub>i</sub> of young growing and adult rats. Am J Physiol 242(6):F705-F710, 1982.
- Haramati A, Mulroney SE, Webster SK. Developmental changes in the tubular capacity for phosphate reabsorption in the rat. Am J Physiol 255:F287-F291, 1988.
- Armbrecht HJ, Gross CJ, Zenser TV. Effect of dietary calcium and phosphorus restriction on calcium and phosphorus balance in young and old rats. Arch Biochem Biophys 210:179–185, 1981.
- Horst RL, DeLuca HF, Jorgensen NA. The effect of age on calcium absorption and accumulation of 1,25-dihydroxyvitamin D3 in intestinal mucosa of rats. Metab Bone Dis & Relat Res 1:29–32, 1978.
- Kalu DN, Cockerham R, Yu BP, Roos BA. Lifelong dietary modulation of calcitonin levels in rats. Endocrinology 113:2010–2016, 1983.
- 8. Kalu DN, Hardin RH, Cockerham R, Yu BP. Aging and dietary modu-

- lation of rat skeleton and parathyroid hormone. Endocrinology 115:1239-1247, 1984.
- Kelijman M. Age-related alterations of the growth hormone/insulinlike-growth-factor I axis. J Am Geriatr Soc 39:295-307, 1991.
- Meyer BR. Renal function in aging. J Am Geriatr Soc 37:791-800, 1989.
- Meyer BR, Hirsch BE. Renal function and the care of the elderly. Compr Ther 16(9):30-37, 1990.
- Darmady EM, Offer J, Woodhouse MA. The parameters of the aging kidney. J Pathol 109:195-207, 1973.
- Kee CC. Age-related changes in the renal system: Causes, consequences, and nursing implications. Geriatric Nursing 13(2):80–83, 1992.
- Anderson S, Brenner BM. Effects of aging on the renal glomerulus. Am J Med 80:435–442, 1986.
- Huether SE. Structure and function of the renal and urologic systems.
  In: Schrefer S, Ed. Pathophysiology: The Biologic Basis for Disease in Adults and Children. St. Louis: Mosby-Year Books, pp1212–1234, 1994.
- Fuhr J, Kaczmarczyk J, Kruttgen CD. Eine einfache colormetrische Methode zur Inulinbestimmung fur Nierenclear-anceuntersuchungen bei Stoffwechselgesunden und Diabetikern. Klinische Wochenschrift 33:729-730, 1955.
- 17. Chen PS, Toribara TY, Warner H. Microdetermination of phosphorus. Anal Chem 28:1756–1758, 1956.
- Barrows CH Jr., Falzone JA Jr., Shock NW. Age differences in the succinoxidase activity of homogenates and mitochondria from the livers and kidneys of rats. J Gerontol 15:130-133, 1960.
- Beauchene RE, Fanestil DD, Barrows CH. The effect of age on active transport and sodium-potassium-activated ATPase activity in renal tissue of rats. J Gerontol 20:306-310, 1965.
- Haramati A. Tubular capacity of phosphate reabsorption in superficial and deep nephrons. Am J Physiol 248:F729–F733, 1985.
- Kiebzak G, Sacktor B. Effect of age on renal conservation of phosphate in the rat. Am J Physiol 251:F399-F407, 1986.
- Sorribas V, Lotscher M, Loffing J, Biber J, Kaissling B, Murer H, Levi M. Cellular mechanisms of the age-related decrease in renal phosphate reabsorption. Kidney Int 50:855–863, 1996.
- Brenga G, Holmes RP. Interaction between components in biological membranes and their implications for membrane function. Prog Biophys Mol Biol 43:195-257, 1984.
- Specktor AA, Yorek MA. Membrane lipid composition and cellular function. J Lipid Res 26:1015–1035, 1985.
- Stubbs CD, Smith AD. The modification of mammalian membrane polyunsaturated fatty acid composition in relation to membrane fluidity and function. Biochim Biophys Acta 779:89–137, 1984.
- Levi M, Jameson DM, Wieb Van Der Meer B. Role of BBM lipid composition and fluidity in impaired renal P<sub>i</sub> transport in aged rat. Am J Physiol 256:F85–F94, 1989.
- Pratz J, Corman B. Age-related changes in enzyme activities, protein content, and lipid composition of rat kidney brush border membranes. Biochim Biophys Acta 814:265-273, 1985.
- Pratz J, Ripoche P, Corman B. Cholesterol content and water and solute permeabilities of kidney membranes from aging rats. Am J Physiol 253:R8-R14, 1987.
- De Smedt H, Kinne R. Temperature dependence of solute transport and enzyme activities in hog renal brush border membrane vesicles. Biochim Biophys Acta 648:247-253, 1981.
- Friedlander G, Shahedi M, Le Grimellec C, Amiel C. Increase in membrane fluidity and opening of tight junctions have similar effects on sodium-coupled uptakes in renal epithelial cells. J Biol Chem 263:11183-11188, 1988.
- Levi M, Baird BM, Wilson PV. Cholesterol modulates rats renal brush border membrane phosphate transport. J Clin Invest 85(1):231–237, 1990.
- 32. Molitoris BA, Alfrey AC, Harris RA, Simon FR. Renal apical mem-

- brane cholesterol and fluidity in regulation of phosphate transport. Am J Physiol **249:**F12–F19, 1985.
- 33. Corvilain J, Abramow M. Some effects of human growth hormone on renal hemodynamics and on tubular phosphate transport in man. J Clin Invest 41:1230–1235, 1962.
- Corvilain J, Abramow M. Effect of growth hormone on tubular transport of phosphate in normal and parathyroidectomized dogs. J Clin Invest 43:1608–1612, 1964.
- Caverzasio J, Faundez R, Fleisch H, Bonjour JP. Tubular adaptation to P<sub>i</sub> restriction in hypophysectomized rats Pflugers Arch 39(1):17-21, 1981.
- Caverzasio J, Bonjour JP. Insulin-like growth factor I stimulates Nadependent P<sub>i</sub> transport in cultured kidney cells. Am J Physiol 257:F712-F717, 1989.

- Gregerman RI. Mechanisms of age-related alterations of hormone secretion and action: An overview of 30 years of progress. Exp Gerontol 21:345–365. 1986.
- 38. Fujita T, Ohata M, Orimo H, Yoshikawa M. Age and parathyroid hormone inactivation by kidney tissue. J Gerontol 26:20-23, 1971.
- Fujita T, Okano K, Orimo H, Ohata M, Yoshikawa M. Age and fate of parathyroid hormone. J Gerontol 27:25–27, 1972.
- Marcus R, Gonzales D. Age-related change in parathyroid hormonedependent cyclic AMP formation in rat kidney. Mech Ageing Dev Mol 20:353–360, 1982.
- 41. Armbrecht HJ, Wongsurawat N, Zenser TV, Davis BB. Differential effects of parathyroid hormone on the renal 1,25-dihydroxyvitamin  $D_3$  and 24,25-dihydroxyvitamin  $D_3$  production of young and adult rats. Endocrinology **111**:1339–1344, 1982.