

intracutaneous injection is not merely followed by a prompt increase in capillary permeability; but within a surprisingly short interval of 25 to 30 minutes there is considerable evidence of leucocytic migration into the treated areas. The small vessels are packed with polymorphonuclears closely adhering to the endothelial wall in the act of migrating into the extra-capillary spaces which reveal already moderate infiltration of these cells. Furthermore, the substance is positively chemotactic when placed in capillary glass tubes sealed at one end and introduced into an inflamed peritoneal cavity, the tubes soon become filled with leucocytes. The inactive crystalline material obtained from blood serum and injected intracutaneously, distilled water, or saline are all totally devoid of inducing such prompt accumulation and migration of leucocytes through the endothelial wall. Turpentine, which in the skin causes a rapid increase in capillary filtration as indicated by the pronounced seepage of dye in the affected area, is equally incapable of exhibiting as rapid a migration of leucocytes as the active crystalline material. The implications of these observations are obvious. The studies are being continued and will be reported *in extenso* elsewhere.

Conclusions. For the sake of convenience the name *leukotaxine* is tentatively proposed for this active crystalline nitrogenous substance which is evidently released by injured tissue and is readily recovered in inflammatory exudates. "Leukotaxine" *per se* appears to be capable of rapidly initiating the usual sequences of the inflammatory reaction first by inducing a prompt increase in capillary permeability and secondly by causing an extremely rapid aggregation and migration of leucocytes through the endothelial wall.

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Cane Molasses versus Beet Molasses as a Source of Vitamin B₆ and Lactoflavin.

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Regardless of the fact that Nelson, Heller and Fulmer¹ published experimental evidence concerning the value of cane molasses as a source of the whole vitamin B complex, molasses is still generally looked upon as a qualitatively inert food constituent.

In 1935 the experimental data of Nelson, Heller and Fulmer re-

¹ Nelson, V. E., Heller, V. G., and Fulmer, E. I., *Indust. and Engin. Chem.*, 1925, **17**, 199.

ceived substantial confirmation. Birch, György and Harris² showed that cane molasses is very rich in vitamin B₆. As molasses is one of the major components of the diet of pellagrins in the cotton districts of the United States, the high vitamin B₆ potency of molasses indicates that vitamin B₆ has no antipellagra action for human beings.

Furthermore, Jukes and Lepkovsky³ recently have found in cane molasses a fairly good source of the preventive factor for chicken pellagra, the so-called "filtrate" factor; it therefore seems unlikely that lack of the chicken pellagra factor would be responsible for the occurrence of human pellagra.

Nelson, Heller and Fulmer measured the whole vitamin B complex, its composite nature being unknown at that time, and they established the fact that cane molasses is far superior to beet molasses as a source of vitamin B. I have repeated these experiments, using

TABLE I.
Average values for distribution of lactoflavin and vitamin B₆ in molasses.

Molasses Sample Fed to Rats	Amt. Given Daily gm.	Tests for Lacto- flavin (Vitamins B ₁ and B ₆ pro- vided):	Tests for Vitamin B ₆ (Vitamin B ₁ and Lacto- flavin provided):	
		Aver. Weekly In- crease in Wt. of Rats gm.	Aver. Week- ly Increase in Wt. of Rats gm.	Healing of Specific Dermatitis
Cane molasses I, blackstrap (Louisiana)	0.075	—	3	No
	0.125	—	5	Yes
	0.25	—	6.5	"
	0.5	—	—	—
	0.75	2.5	—	—
	1.5	7.5	—	—
Beet molasses (Germany)	0.25	—	0	No
	0.5	—	-1	"
	0.75	0	1	"
	1.0	—	-1	"
	1.5	-4	—	—
Beet molasses (Michigan)	0.75	—	-1	No
	1.5	—	0	"
Syrugold	0.25	—	-2	"
	0.5	—	2	"
	0.75	—	0	"
	1.5	-1	—	—
Karo (blue label)	0.5	—	-2	No
	1.0	—	2	"
	2.0	—	3	Inconstant
	2.5	1.5	2.5	Yes
	5.0	-2	—	—

² Birch, T. W., György, P., and Harris, L. J., *Biochem. J.*, 1935, **29**, 2330.

³ Jukes, T. H., and Lepkovsky, S., *J. Biol. Chem.*, 1936, **114**, 117.

my method for the separate determination of lactoflavin and vitamin B₆.⁴ I have also tested Karo (blue label), which contains a molasses preparation, and Syrugold, a refined cane molasses preparation (National Sugar Refining Co.), for potency of vitamin B₆ and lactoflavin. The results are recorded in Table I.

In addition to the sample of blackstrap molasses given in Table I, I tested 8 other samples of crude cane molasses for their vitamin B₆ content. In 7 samples, from Louisiana, the vitamin B₆ titer varied between 0.1 and 0.3 gm. per rat per day (in two it was 0.1 gm., in three 0.15 gm. and in two 0.3 gm.); in the eighth sample, of unknown provenience, the titer was 0.4 gm. per rat per day. In a specimen of commercial blackstrap treacle put up by Fowler (England), the rat-day dose of vitamin B₆ was found² in 0.25 gm. Beet molasses samples from Germany and from Michigan were both devoid of vitamin B₆ (see Table I).

The Karo sample recorded in Table I showed B₆ activity at the daily dose per rat of 2 to 2.5 gm., whereas in 2 other samples of blue label Karo I failed to detect any curative action for deficiency of vitamin B₆. The implications of this finding in connection with questions of infant feeding will be discussed elsewhere.⁵

Conclusions. Crude cane molasses is a good source of vitamin B₆, the B₆ titer varying from 0.1 to 0.4 gm. per rat per day, with an average of 0.15 gm. Crude cane molasses also contains small amounts of lactoflavin. Beet molasses is practically devoid of both vitamin B₆ and lactoflavin.

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Influence of Carbon Dioxide upon Blood Pressure Reaction to Oxygen Deficiency.

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Gellhorn¹ showed recently that CO₂ either completely offsets or greatly diminishes the effects of oxygen deficiency on various sensory and psychic phenomena. This effect may be thought to be due to the effect of CO₂ on respiration whereby the acapnia associated

⁴ György, P., *Biochem. J.*, 1935, **29**, 741.

⁵ György, P., in preparation.

¹ Gellhorn, E., *Nature*, 1936, **137**, 700; *Am. J. Physiol.*, 1936, **117**, 75; *Proc. Am. Physiol. Soc.*, 1936, p. 59; *Sigma Xi Quarterly*, 1937 (June), in press.