

Antithromboplastic Activity of Monosodium Alpha Tocopherol Phosphate and Inositol Phosphatide.* (17714)

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Thomas(1) showed that intravenous injections of thromboplastin, a sedimentable tissue component, into mice caused a characteristic response consisting of ataxia, convulsions and coma. The easy reproduction of these symptoms with small, relatively constant quantities of thromboplastin provided a more or less quantitative method for titrating the thromboplastic activity of these suspensions. Thomas further showed that this response could be prevented by both heparin and congo red. This paper deals with the inhibition of thromboplastin by two other substances, monosodium alpha tocopherol phosphate[†] and inositol phosphatide[‡]. Overman(2) reported the antithromboplastic action of inositol phosphatide on the basis of *in vitro* work. No similar studies seem to have been carried out with the alpha tocopherol preparation.

Method. These experiments were carried out on mice, each weighing from 12 to 14 g. Mixed brain and lung thromboplastin[§] was standardized by the method of Thomas(1) as follows: It was diluted with distilled water to make up concentrations varying from 1:10 to 1:50. Each solution was incubated for 60 minutes at 37.5°F and was injected into the tail vein of a mouse, which was observed for the characteristic response of ataxia, convulsions and coma. The minimum effective dose which produced this response was 0.2 ml of thromboplastin in 1:30 dilution. Alpha

tocopherol was then added to a 1:20 thromboplastin solution in amounts varying from 7.5 to 100 γ . Immediate injection of the solution into the tail veins of mice in 0.2 ml aliquots was ineffective in inhibiting the characteristic response to thromboplastin. When, however, the solution was incubated for 60 minutes at 37.5°C and was similarly injected, it was found that 15 γ of alpha tocopherol phosphate was effective for this purpose, while 7.5 γ per 0.2 ml aliquot lacked sufficient antithromboplastic activity to prevent the reaction.

Results. Similar experiments with inositol phosphatide showed (1) that without incubation the test solution had no inhibitive effect and (2) that 36 γ of the substance per 2 ml aliquot was capable of inhibiting the response of a mouse to a 1:20 dilution of thromboplastin when the same aliquot was used. At 18 γ no antithromboplastic activity was observed. Since it had been shown that both monosodium alpha tocopherol phosphate and inositol phosphatide were antithromboplastic in the concentrations used in this experiment after the solutions had been incubated, another experiment was done to determine whether the antithromboplastic action would occur if the test solutions were injected into the mouse prior to the injection of a 0.2 ml aliquot of the minimum effective dose of thromboplastin. After 15 to 100 γ of monosodium alpha tocopherol phosphate per 0.2 ml aliquot had been injected intravenously into the mouse, either 5 or 30 minutes were permitted to elapse before 0.2 ml of the 1:20 thromboplastin dilution was injected into the same mouse. All of the mice showed the characteristic response to thromboplastin, no antithromboplastic activity being demonstrated in any of them. At 1,000 γ inhibition of thromboplastin occurred after a 5 and a 30 minute interval.

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1. Thomas, L., *Bull. Johns Hopkins Hosp.*, 1947, v81, 1.

† Supplied by Mr. Rudolph Frundt of the Warren-Teed Products Co., Inc.

‡ Supplied by Dr. George Mast of C.S.C. Pharmaceuticals.

2. Overman, A. S., and Wright, I. S., *J. Biol. Chem.*, 1948, v174.

§ Sol U Plastin supplied by Mr. Blanchard of Schafflin and Co.

of inositol phosphatide, compared with a value of 72.8 dynes cm² for distilled water.

These observations suggest the possibility (1) that thromboplastin presents a surface which can adsorb monosodium alpha tocopherol phosphate and inositol phosphatide, (2) that the utilization of this surface by these surface-acting agents explains why their injection into mice in a solution of thromboplastin prevents the typical reaction which normally occurs in these animals following the injection of thromboplastin alone, and (3) that the addition of fibrinogen to the solution interferes with this protective action because

of the surface presented by fibrinogen molecules.

Summary. Experimental studies on mice showed that the typical reaction of these animals to the injection of thromboplastin can be prevented if monosodium alpha tocopherol phosphate or inositol phosphatide is incubated with the thromboplastin before it is injected. When fibrinogen is added to the test solution, the inhibitory effect of each of these substances is lost. It is possible that alterations in surface tension may explain these phenomena.

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Use of Maxted's Method for Group Classification of Hemolytic Streptococci.*† (17715)

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Maxted(1) has recently described a method for grouping beta-hemolytic streptococci in which the streptococci are lysed by a substance, presumably an enzyme, produced by a strain of *Streptomyces albus*. Such lysates contain the streptococcal C substance and can be used as antigens in the precipitin reaction for group classification. The simplicity, reliability and usefulness of Maxted's technique have been confirmed in a study of 1,010 consecutive strains of streptococci.

Methods. The strain of *Streptomyces albus*, obtained from Maxted, was grown on

the following medium which is a modification of that described previously(1).

Bacto-peptone (Difco)	0.5 g
Yeast extract, desiccated	0.6 "
Dibasic sodium phosphate, hydrous	0.2 "
Glucose	0.2 "
Fildes' extract(2)	2.5 ml
Agar	1.25 g
Distilled water	100.00 ml

These constituents, with the exception of the Fildes' extract, were mixed and dispensed into a 1000 ml Roux bottle. After sterilization at 120°C for 20 minutes, the bottle was cooled to about 40°C, the Fildes' extract added, and the agar medium allowed to gel with the bottle in the horizontal position. The surface was inoculated evenly with 1 ml of a heavy suspension of *Streptomyces albus* spores in 0.85% sodium chloride solution. After incubation at 35°C for 5 days, the culture was frozen at -10°C for 12 hours and then allowed to thaw at room temperature. This procedure separated the agar gel of the

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1. Maxted, W. R., *Lancet*, 1948, v253, 255.

2. Fildes, P., *Brit. J. Exp. Path.*, 1920, v1, 129.