

for 24 hours. (5) One 15 mg troche decreases bacterial counts significantly for at least one hour. (6) A second troche administered one hour after the first adds considerably to the antibacterial effect.

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Liver Storage of Vitamin B₁₂ by the Rat.* (19139)

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Assays for vit. B₁₂ by the use of rat and chick growth, or microbiological methods, are well known. Although available information (1,2) indicates that the amount of vit. B₁₂ in the liver or other organs was lower for animals fed a vit. B₁₂ deficient diet as compared to a supplemented diet, it was of interest to extend these studies and to determine the vit. B₁₂ potency of the livers of rats fed graded amounts of vit. B₁₂. Further, the liver storage of the vitamin, when fed as the crystalline vitamin or as a food, could be evaluated as a method for determining the vit. B₁₂ potency of foods and could be compared with data obtained by the microbiological and animal growth methods. These studies have been conducted, and data have also been obtained on the effect of the level of vit. B₁₂ fed on the liver weight to body weight ratio.

Experimental and results. Weanling male rats of the Holtzman strain were fed a basal ration (corn-soybean oil meal-iodinated casein) without added vit. B₁₂ as a depletion ration for a period of 2 weeks. The rats were then randomized into groups and fed either 0, 3, 6, 9, 15 or 24 μ g of crystalline vit. B₁₂ per kilo of ration or crude supplements of unknown vit. B₁₂ potency for a period of 4 weeks. The ration and procedure used have

been described previously(3). Seven rats were used in each group. After completion of the rat growth assay, all animals from selected groups were retained for studies on the storage of vit. B₁₂ in the liver. The animals were sacrificed by decapitation, the livers removed and weighed. Two gram samples were homogenized in a Waring blender with 50 ml of 0.1 M phosphate buffer, pH 6.8, for one minute, then autoclaved for 5 minutes at 15 lb pressure. After autoclaving, the samples were cooled, made to 100 ml volume, filtered and stored in the refrigerator. Aliquots of these samples were taken for microbiological assay according to the method previously described(4,5). The enzymatic digestion step was eliminated since investigations with short-term autoclaving of the samples as compared to subsequent enzyme

TABLE I. Vitamin B₁₂ Content of Rat Livers As Assayed Using *Lactobacillus leichmannii*.

μ g B ₁₂ added/kg ration	μ g B ₁₂ / g liver*	μ g B ₁₂ / liver	Liver wt (% of body wt)
0	.026 $\pm$.005	.200	6.20
3	.030 $\pm$.003	.306	6.49
6	.037 $\pm$.011	.361	6.09
9	.047 $\pm$.009	.474	5.77
15	.048 $\pm$.006	.442	5.20
24	.066 $\pm$.017	.621	5.00

* Mean and stand. dev.

* We are indebted to Merck and Co. for supplying the vit. B₁₂ used in these studies. Journal Paper No. 45, American Meat Institute Foundation.

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TABLE II. Vitamin B₁₂ Activity of Liver With and Without Alkali Treatment.

Material	Test organism	Sample treatment	
		Autoclaved pH 7, µg/g	Autoclaved Alkali, µg/g
Beef liver	<i>L. leichmannii</i>	.77	.146
	<i>Euglena gracilis</i>	.74	0
Rat liver (B ₁₂ deficient diet)	<i>L. leichmannii</i>	.04	.031
	<i>Euglena gracilis</i>	.02	0
Rat liver (B ₁₂ supplemented diet)*	<i>L. leichmannii</i>	.068	.057
	<i>Euglena gracilis</i>	.054	0

* 24 µg of vit. B₁₂/kg of diet.

treatment gave identical results on three assays. Duplicate analyses were made for each liver and *Lactobacillus leichmannii* 327 (ATCC 7831) was used as the test organism.

The vit. B₁₂ potency of the liver (0.066 ± .017 µg/g) for the group fed an optimum level of vit. B₁₂ was considerably higher than that for the basal group (0.026 ± .005 µg/g) as shown in Table I. In a subsequent experiment, livers from rats fed the basal diet and basal diet plus 24 µg B₁₂ per kg diet were analyzed for vit. B₁₂ in a similar manner. In this experiment, the basal group averaged 0.046 µg/g of vit. B₁₂ per g of liver, and the supplemented group averaged 0.108 µg. These values for the B₁₂ potency of rat livers are approximately 1/10 to 1/20 those observed for beef liver(5).

Since values for the vit. B₁₂ potency of rat liver appeared low compared to that of beef liver, it was of interest to determine how much of the B₁₂ potency in these samples may be attributed to desoxyribosides. Hoffman, *et al.*(6) showed that vit. B₁₂ was destroyed by heating with 0.2 N NaOH at 100°C for 30 minutes and that under these conditions the vit. B₁₂ activity of the desoxyribosides of thymine, guanine, and hypoxanthine was not affected. They also found that about 97% of the total microbiological activity of liver extract disappeared after alkali treatment. Rat liver was subjected to a similar treatment: Two g samples were homogenized in 40 ml 0.1 N NaOH and autoclaved at 15 lb pressure for 30 minutes. The samples were neutralized with 2 N HCl, made to 100 ml volume and filtered. Aliquots were

then taken for microbiological analyses. Rat liver homogenates treated with alkali were found to retain an appreciable amount of vit. B₁₂ activity for *L. leichmannii* (Table II). When rat liver plus crystalline vit. B₁₂ was so treated, the added B₁₂ was destroyed, about 80% of the vit. B₁₂ potency of beef liver was destroyed, but the vit. B₁₂ activity of rat liver was retained. It is known that *L. leichmannii* responds to thymidine and desoxyribosides (7-9), and since rat liver contains alkali stable substances with vit. B₁₂ activity, it is likely that the organism measured desoxyriboside activity after alkali treatment. Vit. B₁₂ complexes that may be present in liver that are stable to alkali could also explain these findings.

This problem was further investigated by the use of the algal flagellate *Euglena gracilis*. Hutner, *et al.*(10) have shown that thymidine does not replace the vit. B₁₂ requirement of this organism. The medium used in these studies was that of Robbins, *et al.*(11) with the exception of NH₄VO₃ which was not included. The method of assay was essentially that of Hutner,[†] *et al.*(10). The organism was cultured on an agar slant containing

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0.5% peptone, 0.1% liver extract, 0.2% sodium acetate and 1.5% agar. The inoculum was prepared in the same medium (pH 7) without the agar. An assay period of 6 days was used, and the amount of growth determined by measuring the turbidity of the cultures with the use of a Coleman Junior Spectrophotometer.

Fresh rat and beef liver homogenates with and without alkali treatment were assayed with *Euglena*. The results indicated that the values for livers from vit. B₁₂-deficient rats were consistently lower in vit. B₁₂ than those for livers from rats fed an optimum level of vit. B₁₂. The values obtained by the vit. B₁₂ *Euglena* method of assay for rat or beef liver treated with alkali were negligible. The results for fresh beef and rat liver with this method of assay compared very well with the values found by the *L. leichmannii* assay (Table II).

These studies indicate that application of the procedure of alkali treatment to low potency samples, that retain a large portion of the vit. B₁₂ activity after alkali treatment for *L. leichmannii*, may not provide information that can be reliably interpreted. Other studies(12) have shown that the desoxyribonucleic acid level of livers from vit. B₁₂-supplemented rats is 10-20% higher than for deficient rats. Thus, the increases observed in the vit. B₁₂ activity of rat liver when the amount fed was increased can be only partly explained by increased desoxyriboside activity. Although the data obtained after alkali treatment are not definitive, the results for *L. leichmannii* and *Euglena*, as well as the rat assay(1), show that the vit. B₁₂ stored in the liver is related to the vit. B₁₂ intake.

It is of interest that the liver weight was greater per unit of body weight for animals fed the low levels of crystalline vit. B₁₂. Among the deficient animals, the liver weight

was found to be 6.2% of the body weight, while the percentage for the B₁₂-supplemented group was 5.0 (Table I). These data are in agreement with those observed in another experiment in which the percentage of body weight that was liver weight was 6.6 for animals fed the basal diet and 5.2 for animals fed the basal diet plus 24 µg vit. B₁₂ per kg.† The percentage for animals fed the supplemented diet with the food intake restricted to that of the basal group was 5.0.

By the use of the liver storage method of assay, values for fresh beef round, pork ham, lamb leg, kidney, liver, and injectable liver extract have been determined. In the cases of pork and lamb, the data for vit. B₁₂ are in close agreement with those for similar samples studied earlier using the microbiological technic. The values for the remainder of the crude materials tend to be somewhat higher. Details of these studies will be reported at a future date.

Summary. The storage of vit. B₁₂ in rat liver was investigated after feeding rats graded levels of crystalline vit. B₁₂ or of crude supplements. The vit. B₁₂ potency was determined with *L. leichmannii* after short-term autoclaving of the liver homogenates at pH 7.0. The vit. B₁₂ potency was shown to increase as the level fed was increased. The values were only 1/10 to 1/20 those for beef liver, however, and the vit. B₁₂ potency of rat liver was not reduced appreciably by alkali treatment. Studies on the desoxyribonucleic acid levels in rat livers and studies with *Euglena gracilis* assays for vit. B₁₂ indicated that the vit. B₁₂ activity remaining, as determined with *L. leichmannii* after alkali treatment, may be due to the presence of alkali stable forms of vit. B₁₂ as well as to desoxyribosides. The liver weight was shown to be greater per unit of body weight for the rats fed the vit. B₁₂-deficient diet as compared to the optimum diet.

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