

the spatial distribution of the radicals formed in water and their reactions.

1. Patt, H. M., *Physiol. Rev.*, 1953, v33, 35.
2. Thoday, J. M., and Read, J., *Nature*, London, 1947, v160, 608.
3. ———, *Nature*, London, 1949, v163, 133.
4. Giles, N. H., Jr., Beatty, A. V., and Riley, H. P., *Genetics*, 1952, v37, 641.
5. Clark, J. W., Vogel, H. H., Jr., and Jordan, D. L., *Argonne Nat. Lab. Rep.*, 1953, No. 4948, 32.
6. Vogel, H. H., Jr., Blomgren, R. D., and Bohlin, N. J. G., *Nucleonics*, 1953, v11, 28.
7. Smith, D. E., Patt, H. M., Tyree, E. B., and Straube, R. L., *PROC. SOC. EXP. BIOL. AND MED.*, 1950, v73, 198.
8. Patt, H. M., Mayer, S. H., Straube, R. L., and Jackson, E. M., *J. Cell. Comp. Physiol.*, 1953, v42, in press.
9. Patt, H. M., Smith, D. E., Tyree, E. B., and Straube, R. L., *PROC. SOC. EXP. BIOL. AND MED.*, 1950, v73, 18.
10. Gray, L. H., *J. Cell. Comp. Physiol.*, 1952, v39, suppl. 1, 57.
11. Bonet-Maury, P., and Lefort, M., *Nature*, London, 1950, v166, 981.
12. Mayer, S. H., and Patt, H. M., *Fed. Proc.*, 1953, v12, 94.
13. Salerno, P. R., and Friedell, H. L., *Fed. Proc.*, 1953, v12, 364.
14. Patt, H. M., Blackford, M. E., and Straube, R. L., *PROC. SOC. EXP. BIOL. AND MED.*, 1952, v80, 92.
15. Read, J., *Brit. J. Radiol.*, 1952, v25, 336.
16. Patt, H. M., and Blackford, M. E., *Argonne Nat. Lab. Rep.*, 1951, No. 4713, 48.
17. Bellock, S., and Krebs, A. T., *Army Med. Research Lab. Rept.* 1951, No. 6-64-12-06-(43).

Received June 22, 1953. P.S.E.B.M., 1953, v84.

Effect of Carbonic Anhydrase Inhibitor (6063) on Arterial-Alveolar CO₂ Gradient in Man.* (20587)

E. L. BECKER, J. E. HODLER,[†] AND A. P. FISHMAN. (Introduced by Homer W. Smith.)

From the Department of Physiology, New York University College of Medicine, New York City.

Inhibitors of carbonic anhydrase activity have been employed in the effort to block the *in vivo* reaction, $\text{H}_2\text{O} + \text{CO}_2 \rightleftharpoons \text{H}_2\text{CO}_3$. Various physiological mechanisms are dependent upon this reaction. Höber(1) first demonstrated that sulfonilamide, a carbonic anhydrase inhibitor, impairs the acidification mechanisms of the frog kidney. The use in the dog of stronger sulfonamide inhibitors, especially Diamox (6063 or 2-acetylaminio-1, 3,4-thiadiazole-5-sulfonamide), has implicated carbonic anhydrase in the secretion of hydrochloric acid by the stomach(2) and of bicarbonate by the pancreas(3). It has been re-

cently shown(4) that Diamox increases plasma bicarbonate concentration and pCO₂ in marine fishes, an effect possibly attributable to inhibition of erythrocyte carbonic anhydrase.

Roughton(5) showed that carbon dioxide exchange in unanesthetized humans was impeded when sulfonilamide was combined with exhaustive exercise. In anesthetized dogs(6), Diamox is reported to interfere with CO₂ transport, possibly by inhibition of the erythrocyte enzyme system. It was the purpose of this study to determine whether the carbonic anhydrase inhibitor, Diamox, in concentrations adequate to inhibit renal acid excretion, would interfere with the transport of CO₂ from pulmonary capillary blood into the alveoli in normal, non-anesthetized human subjects.

The subjects were 3 convalescent patients without cardiac, renal, or pulmonary diseases from the First (Columbia University) Medical Division of Bellevue Hospital. These pa-

* This work was supported in part by research grants from the Atomic Energy Commission and the National Heart Institute of the National Institutes of Health, Public Health Service. These studies were conducted in the Cardio-Pulmonary Laboratory, Chest Service, Bellevue Hospital, with the technical assistance of Mrs. H. Bodeen.

[†] Fellow of the Genossenschaft für Biologisch-medizinische Stipendien, Basel, Switzerland.

TABLE I. Effects of Diamox (6063) on Resting Man.

	Time, min.	V, ml/min.	Urine Bicar- bonate, mM/L	U _{Na} V, μEq/min.	U _K V, μEq/min.	C _{Cr}	Respiratory gases				Arterial blood		
							VE, M ² /BSA	P ^A CO ₂ , mm Hg	RQ	pH	P _a (CO ₂ , mm Hg	Total CO ₂ , mM/L	
Subject A: Control	0	.9		229	89	128		4.12	43	.89	7.39	46	24.2
	10	.7	39.1			7.35*		3.98	41	.87	7.39	45	24.2
Subject A: After Diamox	30												
	72	5.3		638	244	137							
	83	13.7		700	288								
	86	15.3	33	596	231	130							
	100	14.0		567	213								
	117	8.3		590	222	130							
	125	5.6	57	586	180		4.02	40	.81	7.37	42	21.5	
Subject B: Control	0	2.9				134	4.01	38	.76	7.35	40	20.5	
	12						3.67	39	.83	7.40	40	23.8	
Subject B: After Diamox	120						3.77	40	.81	7.41	41	23.2	
	150						3.55	40	.81	7.38	37	22.2	
							3.62	38	.80	7.37	39	22.0	

* Collected under oil.

TABLE II. Effects of Diamox (6063) on Man during Mild Exercise.

	Time, min.	Urine			Ccr	Respiratory gases				Arterial blood		
		V, ml/min.	Bicar- bonate, mM/L	Unav, μEq/min.		UkV, μEq/min.	pH	VE, M ² /BSA	VO ₂ , M ² /BSA	RQ	pH	P _a CO ₂ , mm Hg
Rest	0	.7		50		6.81	4.98	141	.89	7.41	42	23.4
Exercise	45	3.3	4.35	51	41.4	6.73	6.9	267	.82	7.41	42	23.0
Diamox	54											
Rest	75	5.77		165	180	7.3	4.33	146	.80	7.39	42	20.3
	110	7.22		446	380	7.7						
	125	4.34	54.8	335	293	7.7						
Exercise	147	3.64	55.2	396	298	7.8	6.68	253	.81	7.36	44	21.8
	212	2.19		314	234	8.0						

tients were trained by repeated cardio-pulmonary studies until they could relax with constant oxygen consumption and RQ despite arterial puncture and gas collection. Two of the subjects were studied at rest, the other was also studied during a steady state of mild, active leg exercise.

Methods. After a period of rest an indwelling urethral catheter and brachial arterial needle were placed in position. Observations were begun ½ hour later. Expiratory gas, arterial blood and urine were collected and analyzed according to methods described elsewhere(7). End expiratory gas was collected by a Rahn(8) sampler and analyzed for CO₂ and O₂ using the micro-Sholander gas apparatus. Blood and urine CO₂-contents were determined by the method of Van Slyke and Neill(9). Arterial blood pCO₂ was calculated from the CO₂ content, pH and per cent oxygen saturation, using the line charts of Van Slyke and Sendroy(10), pH was determined directly by the McInnes-Belchen glass electrode. Potassium and sodium concentrations were determined by an internal standard flame photometer. Glomerular filtration rate was estimated from the endogenous creatinine clearance (CCR) using the method of Bonsnes and Taussky(11). After 2 control periods, 50 mg per kg body weight of Diamox† were administered orally. This dose is approximately 5 times that needed to produce an alkaline urine in man, as reported by others. Observations were then repeated one and 2 hours later.

Results. In the resting subjects, Diamox caused an increase in the excretion of sodium, potassium and bicarbonate, and an increase in urine flow. (Table I). These renal effects were not associated with any change in minute ventilation V_E, oxygen consumption V_{O₂}, respiratory exchange ratio (RQ), arterial or alveolar carbon dioxide tension (PaCO₂ and PA_{CO₂}). Mild exercise, sufficient to double the oxygen consumption, did not reveal any im-

pairment in the elimination of carbon dioxide (Table II).

These observations demonstrate that in the resting subject, Diamox may have a substantial renal effect with no interference with the passage of CO₂ from pulmonary capillary blood into alveoli. The combination of Diamox plus increased CO₂ production induced by mild exercise also failed to impede CO₂ elimination into the alveoli. This failure to influence pulmonary CO₂ excretion may be due to the large concentration of carbonic anhydrase in the human erythrocyte as compared to other tissues.

Summary. Diamox (6063) was administered per os to 3 human subjects without pulmonary disease in single doses of 50 mg per kg body weight. This dose elicited typical renal effects (diuresis and increased excretion of bicarbonate, sodium and potassium) but failed, even when combined with mild exercise, to interfere with CO₂ elimination from pulmonary capillary blood into alveolar air.

1. Höber, R., *Proc. Soc. Exp. Biol. and Med.*, 1941, v49, 87.
2. Janowitz, H. D., Colcher, H., and Hollander, F., *Fed. Proc.*, 1952, v11, 78.
3. Hollander, F., and Birnbaum, D., *Trans. N. Y. Acad. Sci.*, 1952, Ser. II, v15, 54.
4. Heinemann, H. O., and Hodler, J. E., *Bull. Mt. Desert Island Biol. Lab.*, 1953.
5. Roughton, F. J. W., Dill, D. B., Darling, R. C., Graybiel, A., Knehr, C. A., and Talbott, J. H., *Am. J. Physiol.*, 1941, v135, 77.
6. Tomashefski, F., Clark, R. T., and Chinn, H. I., *Fed. Proc.*, 1953, v12, 144.
7. Baldwin, E. deF., Courmand, A., and Richards, D. W., *Medicine*, 1948, v27, 243.
8. Rahn, H., and Otis, A. B., *J. App. Physiol.*, 1949, v1, 717.
9. Van Slyke, D. C., and Neill, J. M., *J. Biol. Chem.*, 1924, v61, 523.
10. Van Slyke, D. D., and Sendroy, J., *J. Biol. Chem.*, 1928, v79, 781.
11. Bonsnes, R. W., and Taussky, H. H., *ibid.*, 1945, v158, 581.

† Diamox was kindly supplied to us by the American Cyanamid Co., Stamford, Conn.