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Mechanisms for reducing/ eliminating chronic neuropathic pain with a focus on platelet-rich plasma

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Abstract

Peripheral nerve trauma commonly results in chronic neuropathic pain by up-regulating the synthesis and release of pro-inflammatory mediators from local and invading cells and inducing hyperexcitability of nociceptive neurons and spontaneous electrical activity. The pain decreases when these cells down-regulate genes supporting the pro-inflammatory state, up-regulate genes for expressing anti-inflammatory factors, and modulate genes that reduce nociceptive neuron spontaneous electrical activity. Pharmacological agents, the primary technique for reducing pain, do not eliminate pain, and <50% of patients achieve benefits because they do not address the underlying causes of pain. Alternative techniques providing longer lasting, but not complete or long-term pain relief include surgical interventions, electrical stimulation, and antibody treatment. Anti-inflammatory mediators can reduce pain, but the effect is not complete or long-lasting. Platelet-rich plasma (PRP) contains a readably available evolutionarily developed cocktail of factors that induce longer-lasting and more significant, but not complete, pain relief than other techniques. However, a novel study shows that unique formulations of PRP can induce long-term pain elimination. This review examines (1) the efficacy of drugs, regenerative peripheral nerve interface (RPNI), targeted muscle reinnervation (TMR), and PRP in reducing chronic neuropathic pain, (2) recent clinical data showing that a novel PRP application technique induces long-term chronic neuropathic pain reduction/elimination, and (3) discusses why the novel PRP may be more effective in reducing/eliminating chronic neuropathic pain.

KEYWORDS

axon regeneration, chronic neuropathic pain, nerve repair, pain elimination, plateletrich plasma, platelets

Impact statement

Peripheral nerve trauma and surgical interventions result in 60% of individuals suffering chronic neuropathic pain. The standard technique for reducing pain is pharmacological agents, although they may not be effective, may reduce but not eliminate pain, are not long-lasting, are strongly addictive, and their side effects may preclude their use. Physiologically, chronic pain is reduced/eliminated when injured axons reinnervate their targets. However, because, following nerve repairs, <50% of patients recover function, most patients suffer chronic pain. Novel techniques are required that induce meaningful recovery or directly reduce/eliminate chronic neuropathic pain. This review examines the efficacy of pharmacological agents and other techniques for their analgesic efficacies. It then discusses a novel technique involving platelet-rich plasma (PRP), which reliably and rapidly induces long-term chronic neuropathic pain reduction/elimination. Finally, it briefly discusses various platelet-released factors that may be responsible for this influence and their mechanisms of action.

Introduction

Up to 16% of the US population suffers chronic neuropathic pain due to trauma, amputation, and surgery [1, 2], with peripheral nerve trauma and surgical interventions leading to pain in 60% of patients [3–5]. For those who undergo peripheral nerve surgical procedures, one study found >50% have significant pain reduction [6], while another 73% continued to have or developed pain [7]. The pain was chronic and intense for about 30% [3, 8], debilitating for many [9], and challenging to treat [10, 11]. Of patients presenting to pain clinics reporting chronic neuropathic pain, 78% suffered pain after 6 months, decreasing to 56% after 12 months [12].

Surgical interventions [13–15], electrical stimulation [16–18] antibodies against pro-inflammatory mediators and their receptors [19, 20], and drugs that block nociceptive neurons' hyperexcitability and spontaneous ectopic electrical activity [21–23] provide long-term chronic neuropathic pain relief, but not elimination, to <50% of patients [24, 25].

Extensive evidence shows that injury-induced inflammation underlies neuropathic pain [3]. This suggests that administering anti-inflammatory agents should reduce chronic pain. However, clinically, administering anti-inflammatory drugs prolongs rather than shortens the time to pain elimination, while administering pro-inflammatory mediators reduces pain more rapidly [26]. While counterintuitive, this is because inflammation induces neutrophil invasion and up-regulates the synthesis and release of pro-inflammatory factors, which trigger an anti-inflammatory response [26]. Therefore, reducing/eliminating chronic neuropathic pain requires understanding which cells

are recruited by injuries, the sequences of their recruitment, and what leads to the up- or down-regulation of specifically released factors.

This paper examines the efficacy of drugs and PRP in reducing pain and the results of two novel clinical techniques involving PRP, which induce long-term chronic neuropathic pain reduction/elimination. Finally, the paper discusses the pro-algesic and analgesic roles played by some platelet-released factors that induce pain reduction/elimination.

Pharmaceutical agents

Clinically, pharmacological agents are best for reducing pain and providing adequate pain control to 30%–40% of patients [27]. Among the most effective opioid receptor agonists are strong [1, 28–34] followed by weak [35, 36] opioids [28], anticonvulsive drugs [37], such as gabapentin [38–42], tricyclic antidepressants [43], and the selective norepinephrine and anti-epileptic drug pregabalin [44]. While opioids are the most effective [45], their efficacies are increased by combining them with other drugs [46]. The clinical efficacies of other techniques, such as the local application of capsaicin [47] and lidocaine [48], are less well-established and are still being tested [49]. Recently, suzetrigine was FDA approved (first in class JAN 2025) as a non-opioid analgesic of comparable efficacy to higher-potency opioids. [50]. Its efficacy compared to PRP is not known. However, it has been shown to induce mild to moderate severe adverse events [50], while PRP induces no known adverse events.

The anesthetic ketamine is effective against chronic neuropathic pain [51]. It is considered to act by inhibiting the N-methyl-D-aspartate (NMDA) receptor and possibly other mechanisms, such as enhancing descending inhibition and central site anti-inflammatory actions [51]. However, short-term NMDA infusions induced potent analgesia only during its administration, while prolonged infusion (4–14 days) induces analgesia for up to 3 months following infusion [51]. Unfortunately, ketamine's clinical side effects include nausea/vomiting, psychedelic symptoms (hallucinations, memory defects, panic attacks), cardiovascular stimulation, and somnolence, with a minority of patients suffering hepatotoxicity [51].

No pharmacological agent provides long-term analgesia [52], and for patients with chronic pain, 54% use opioid medications daily, despite up to 97% reporting inadequate pain relief [6, 53]. However, their use is limited because of adverse effects [54, 55] and problems with abuse, dependence, overdose, and death [54, 55]. Therefore, it is essential to balance opioid pain control and the development of opioid dependence [56]. These difficulties can be reduced by multimodal analgesic plans, non-opioid medications, and regional application techniques [31, 56]. Nevertheless, novel pain relief techniques are required [57],

including developing alternative forms of nerve surgery [6], and pharmacological agents.

Targeted muscle reinnervation (TMR) and regenerative peripheral nerve interface (RPNI)

Removing painful neuromas reduces but does not eliminate pain [13], and there is a high rate of neuroma and pain redevelopment [58]. However, following neuroma removal, the pain that normally develops is reduced by securing the nerve stump to an autograft or allograft [59, 60]. For lower extremity amputations, pain is reduced by nerve capping or implanting nerve stumps in bone [24, 61]. However, there is still no long-term chronic neuropathic pain reduction [62].

The most effective techniques for preventing or reducing chronic neuropathic pain or post-amputation neuroma pain are regenerative peripheral nerve interface (RPNI) and targeted muscle reinnervation (TMR) [63–67]. RPNI involves coapting a nerve stump into a small denervated muscle grafts, while TMR involves coapting the proximal nerve stump to the proximal motor nerve innervating a small muscle graft. Thus, following neuroma excision, both RPNI and TMR reduce pain development [68] and clinically reduce post-amputation neuroma pain in 75–100% of patients [64–67, 69–71] and phantom limb pain in 45–80%. However, TMR has the significant limitation of being only effective if applied <3 months post-trauma [72], requires sacrificing a motor nerve and cannot be used if a goal is to both reduce pain and restore function.

Target reinnervation and cessation of axon regeneration

Abnormal spontaneous electrical activity of regenerating dorsal root axons is closely associated with chronic neuropathic pain [73–75]. Clinically, chronic pain reduction/elimination occurs only slightly before or in association with initial signs of functional recovery [76]. These findings led to the hypothesis that pain remains chronic when axons are regenerating [73] and only decreases or is eliminated when axons reinnervate targets, stop regenerating [63, 73, 77, 78], take up a target-derived factor/s [76, 79–81], which silence hyperexcitable nociceptive axons [63].

Supporting this hypothesis is that the extent of pain reduction decreases proportionately with the increasing extent of functional recovery [82, 83]. In rats, pain behavior is reduced or eliminated when axon regeneration is stopped/inhibited [73, 84], such as by applying semaphorin 3A [85] and injecting small-interfering RNA (siRNA) into axotomized sensory ganglia to

block growth-associated protein-43 (GAP-43) expression [73, 86]. This hypothesis is consistent with studies showing that TMR reduces/eliminates chronic neuropathic pain [69, 77, 87], including complex regional pain syndrome (CRPST) type II [88]. This idea is also consistent with rat chronic pain behavior being blocked by tetrodotoxin (TTX), GAP-43 knockdown, and semaphorin 3A, which stop axon regeneration and the electrical activity of nociceptive neurons [73]. Target reinnervation and the cessation of axon regeneration are consistent with TMR and RPNI reducing/eliminating chronic neuropathic pain, which occurs in 71%–100% of the subjects [66, 69].

PRP and the reduction/elimination of chronic neuropathic pain

One of the challenges in using PRP is consistency in the findings between different studies. Thus, some clinical studies concluded that PRP provided little or no pain relief for tendinosis or rotator cuff tears [89–91]. Meta-analyses of multiple studies support this conclusion [89, 92, 93]. However, other clinical studies found that PRP reduced pain associated with tendinosis [94, 95], tendon injury [96–98], rotator cuff tears [99, 100] osteoarthritis [101, 102], plantar fasciitis [103], and muscle injuries [104]. These findings were supported by meta-analysis [105]. Animal model studies show PRP reduces pain caused by many types of injuries [106, 107], such as skin burn-induced neuropathic pain [108], painful lesions caused by *mycobacterium leprae* (leprosy bacteria) [109], and rat spinal cord injury sites [110]. Clinically, PRP also reduces peripheral nerve pain when applied to digital [111] and sciatic [112] nerve crush sites, pudendal nerve neurolysis surgery sites [113], the median nerve at the carpal tunnel's proximal edge [114], and when injected into the perineurium of patients suffering from diabetic neuropathic pain [115]. These techniques result in >80% of patients achieving ca. three months of pain relief [116].

Clinically, a single epidural PRP injection provides lower back pain relief for up to 6 months [117, 118] while reducing complex chronic degenerative spinal pain [119]. Multiple PRP epidural injections provide temporary lumbar radicular pain relief [120], which is significantly increased by adding corticosteroids, although this is effective for only about 50% of patients [121, 122]. In comparative studies, the analgesic efficacy of PRP is similar to [120, 123] or better than [94, 124] that is provided by injecting a corticosteroid. An effective alternative technique for rats involves wrapping nerves in a PRP-coated NeuraWrap Nerve Protector [125]. Nevertheless, two meta-analyses found that in animal models and clinically, although PRP induces pain relief and nerve regeneration, the effect is not long-lasting [114, 126].

Variability in PRP efficacy in reducing neuropathic pain

The efficacy of PRP in reducing pain and the duration of the suppression varies significantly between individuals [127] and studies [128]. This variability is best explained by significant differences in the composition of the PRP due to how it is prepared [129]. These techniques include single vs. double-spin PRP separation and >30 different types of commercially available PRP separation systems [130–132]. These different techniques yield PRP with platelet concentrations ranging from 0.52- to 9.3-fold relative to whole blood [133]. PRP differences are also caused by how and when the platelets are activated before or when the PRP is applied, and the centrifugation parameters [132, 134, 135], which affect the ratios of unactivated vs. activated platelets, numbers of other different cell types, levels of bioactive factors, and leukocyte concentration [128, 130, 135–138], when platelets release their factors, (3) the ratios of the factors released, and (4) the level of factor bioactivity [139].

When comparing the analgesic efficacy of PRP, parameters that are never considered are how it is prepared and applied and the uncontrolled differences in the physiology, health, and products consumed by patients [128, 140, 141]. Thus, platelet aggregation, and therefore its efficacy, is increased by smoking [142], while platelet activation and aggregation are decreased by alcohol consumption [143], which also reduces the response of platelets to thrombin [144] and collagen. Platelet aggregation is also reduced by diets containing isoflavones [145], caffeine [146], quercetin (a flavonoid) [147], and anthocyanins [148]. However, platelet aggregation is increased by diets high in saturated fats [149], sugar [150], and simple carbohydrates [149]. Finally, platelets of patients with high blood pressure have a decreased whole blood platelet count [151], and their platelets have lower bioactive factor concentrations than those with normal blood pressure [152]. Therefore, without standardizing how PRP is prepared and applied, it is not possible to eliminate the variability in the efficacy of PRP and to ensure that PRP exerts its maximum potential effects [138].

Novel clinical PRP application techniques induce long-term chronic neuropathic pain reduction/elimination

Recent case studies show that PRP induces long-lasting and complete pain elimination. These applications involved bridging nerve gaps with an autograft within a PRP-filled collagen tube [153–158] or only a PRP-filled collagen tube [159, 160]. The first technique reduced the pain in 8% and eliminated it in 92% of subjects, while the second eliminated pain in all the subjects. The pain reduction/elimination lasted throughout the 1.1–15 years follow-up. Thus, platelet-released

factors have the capacity to induce long-term pain elimination.

Novel PRP techniques are superior to TMR and RPNI

The novel PRP techniques are superior to TMR and RPNI. (1) They reduce/eliminate pain while promoting meaningful recovery. (2) While TMR is effective when applied up to 3 months post-trauma, its efficacy decreases with longer delays [72]. (3) RPNI requires survival or a small muscle graft, which PRP does not. (4) RPNI requires sacrificing a motor nerve, while using PRP does not.

What underlies the high level of efficacy of the novel PRP application techniques?

While PRP provides short-term pain reduction/elimination [161, 162], the pain returns to 86% of patients [163]. This raises the question of why the novel PRP application techniques induce long-term pain reduction/elimination in 92%–100% of patients. The best explanation is in how the PRP was prepared and applied. First, applying PRP in a liquid form (without fibrin polymerization) causes the platelets to release all their factors within a few hours, while applying PRP in a polymerized fibrin form allows the platelets to release their factors over days [164], thus allowing the factors significantly more time to act. The novel PRP technique involved applying PRP in a polymerized fibrin form.

Second, although the optimal platelet concentration to provide maximal pain relief has not been determined, the degree of pain relief provided by PRP is reported to be linearly related to the number of platelets, the number and concentration of the growth and inflammatory factors they contain, and the number of neutrophils and monocytes [165]. Double centrifugation is the most commonly used PRP separation technique, yielding a ca. 4-fold increase in platelet concentration. This concentration is consistent with the finding that the degree of pain reduction associated with tennis elbow increases as the PRP platelet concentration is increased to 4–6-fold [166], while for knee osteoarthritis [167] and tendinopathies [168], a 3–4 fold concentration increase is recommended. However, it has also been reported that while a 2-fold increased platelet concentration yields good results for tissue healing, a 5-fold increase reduces healing [169, 170], and *in vitro* kills human tenocytes [171]. However, PRP used in the novel techniques prepared using GPS III concentrator tubes (Zimmer Biomet, Warsaw, IN) had a 9.3-fold increased platelet concentration.

Third, PRP from the GPS III concentrator tubes increased leukocytes by four-fold. Fourth, while most studies involve

applying a small amount of PRP (≤ 1 mL), long-term pain relief is associated with the application of a significantly larger volume (4–6 mL). Fifth, long-term pain relief was associated with applying PRP to long (4–16 cm) vs. short (<0.5 cm) lengths of nerve [172]. Sixth, most studies showing a temporary analgesic influence of PRP involve applying it to the surface of nerves. However, PRP provides longer-lasting analgesia when the nerve and applied PRP are surrounded by a collagen tube. The tube reduces the diffusion of platelet-released factors away from the site, resulting in a higher effective concentration of the factors, which allows them to act on the axons for a longer time. This hypothesis is supported by the finding that the efficacy of PRP applied to a rat nerve crush site is increased by surrounding the application site with a collagen tube [112].

Platelet-released factors

Platelets contain and release >300 identified factors [173]. While some are pre-packaged, with different types of platelet granules containing different factors, others are synthesized by platelets. The platelet's environment determines the factor synthesis and release pattern [174–176]. Thus, physiologically, platelets release their factors in a specific order, with some released fast and others more slowly [173, 177, 178]. For example, nerve growth factor (NGF) is released within minutes, while brain-derived neurotrophic factor (BDNF) is released more slowly [176]. This sequence is critical to allow the factors to perform specifically timed functions, such as releasing pro-inflammatory factors first, followed by releasing anti-inflammatory factors, which suppress pain [179].

Platelet-released factors reduce/eliminate pain by additional mechanisms, but journal length limitations do not allow a complete discussion of the platelet-released factors that may be involved in reducing/eliminating pain. However, nerve injury induces voltage gated sodium channel (Na(v)) 1.3 channel expression in nociceptive and higher-order spinal sensory neurons, leading to their hyperexcitability, spontaneous ectopic electrical activity, and the development of neuropathic pain [180–182]. Therefore, one mechanism for reducing/eliminating chronic neuropathic pain is to down-regulate the expression of these channels, thus eliminating the spontaneous electrical activity that underlies pain.

Drugs, such as local anesthetics and other Na(v) channel-blocking techniques, reduce neuropathic pain by inhibiting nociceptive axon spontaneous ectopic nerve activity and hyperactivity [183–185]. The pharmacological blockade of sodium channel activity reduces ectopic electrical activity [185, 186] and reverses nerve injury-induced hyperalgesia [187]. This has been attributed to the blocking of Na(v) 1.8 and Na(v) 1.7 channels, leading to reduced or eliminated nociceptive neuron hyperexcitability. However, the role of Na(v)

1.7 in neuropathic pain must be further investigated, and new analysis of a mouse Na(v)1.8 knockout suggests it is not involved in changing the neurons' post-injury pain threshold following peripheral nerve injury [188]. However, it has also been shown that administering antisense oligonucleotides against Na(v) 1.8 administered intrathecally completely reverses neuropathic pain behavior [189].

This is in contrast to the finding of Lai et al who reported that antisense oligonucleotides directed against Na(v) 1.8 administered intrathecally completely reverse neuropathic pain behavior [18]. It is possible that this discrepancy could be due to the up-regulation of the Na(v)1.7 channel seen in the Na(v)1.8 knockout mouse [12], which might mask an otherwise important role for Na(v)1.8 in neuropathic pain.

However, the pain suppression is not long-lasting. However, in rats, nerve injury-induced chronic pain is reduced by Na(v) 1.3 knockdown [190]. Further, intrathecal IL-10 infusion reduces neuropathic pain [191, 192] in part by down-regulating sodium channel expression in dorsal root ganglion (DRG) neurons [179], resulting in blocking nociceptive neurons' hyperexcitability and spontaneous ectopic electrical activity [21–23, 185, 189, 193]. Although platelets do not contain IL-10, they release large amounts of prostaglandin E2 (PGE2), which induces interleukin-10 (IL-10) release [194, 195], which reduces pain [196–198]. Thus, multiple platelet-released factors can induce long-term chronic neuropathic pain reduction/elimination.

Conclusion

Tissue injury-induced inflammation is the primary trigger of neuropathic pain, with chronic inflammation resulting in chronic pain. Injury induces the release of pro-inflammatory factors from local cells and other cells recruited to the injury site. While inflammation and pain are adverse events, they are required to trigger the normal physiological responses that induce the transition of a pro-inflammatory environment into an anti-inflammatory one, which is necessary for healing and pain elimination. Although some factors initially play pro-inflammatory roles, over time, they begin to play anti-inflammatory roles. Their roles depend on when they act after injury, what other factors are present, the cells on which they act, and the receptors on those cells. Thus, controlling pain requires controlling which factors are released and when. Platelets are an evolutionarily developed toolbox containing a physiological cocktail of factors for controlling cellular environments to promote healing and pain relief. While most studies find PRP only induces short-lived pain relief, two novel clinical techniques show that PRP can induce long-term chronic neuropathic pain elimination in all subjects. Further studies must determine which platelet-released factors, ratios, and concentrations induce these effects.

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All authors contributed to the article and approved the submitted version.

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References

- DiBonaventura MD, Sadosky A, Concialdi K, Hopps M, Kudel I, Parsons B, et al. The prevalence of probable neuropathic pain in the US: results from a multimodal general-population health survey. *J Pain Res* (2017) **10**:2525–38. doi:10.2147/jpr.s127014
- Toth C, Lander J, Wiebe S. The prevalence and impact of chronic pain with neuropathic pain symptoms in the general population. *Pain Med* (2009) **10**(5): 918–29. doi:10.1111/j.1526-4637.2009.00655.x
- Teixeira MJ, da Paz MG, Bina MT, Santos SN, Raicher I, Galhardoni R, et al. Neuropathic pain after brachial plexus avulsion—central and peripheral mechanisms. *BMC Neurol* (2015) **15**:73. doi:10.1186/s12883-015-0329-x
- Campbell J, Neuroma Pain In, Gebhart GF, Schmidt RF, editors. *Encyclopedia of pain 2*. Berlin: Springer-Verlag (2013). p. 2056–8.
- Poppler LH, Mackinnon SE. The role of the peripheral nerve surgeon in the treatment of pain. *Neurotherapeutics* (2019) **16**(1):9–25. doi:10.1007/s13311-018-00695-z
- Felder JM, Ducic I. Impact of nerve surgery on opioid and medication use in patients with chronic nerve injuries. *Plast Reconstr Surg Glob open* (2021) **9**(9): e3789. doi:10.1097/gox.0000000000003789
- Miclescu A, Straatmann A, Gkatziani P, Butler S, Karlsten R, Gordh T. Chronic neuropathic pain after traumatic peripheral nerve injuries in the upper extremity: prevalence, demographic and surgical determinants, impact on health and on pain medication. *Scand J Pain* (2019) **20**(1):95–108. doi:10.1515/sjpain-2019-0111
- Samii M, Bear-Henney S, Ludemann W, Tatagiba M, Blömer U. Treatment of refractory pain after brachial plexus avulsion with dorsal root entry zone lesions. *Neurosurgery* (2001) **48**(6):1269–77. doi:10.1227/00006123-200106000-00016
- Colloca L, Ludman T, Bouhassira D, Baron R, Dickenson AH, Yarnitsky D, et al. Neuropathic pain. *Nat Rev Dis Primers* (2017) **3**:17002. doi:10.1038/nrdp.2017.2
- Attal N, Cruccu G, Haanpää M, Hansson P, Jensen TS, Nurmikko T, et al. EFNS guidelines on pharmacological treatment of neuropathic pain. *Eur J Neurol* (2006) **13**(11):1153–69. doi:10.1111/j.1468-1331.2006.01511.x
- Finnerup NB, Otto M, Jensen TS, Sindrup SH. An evidence-based algorithm for the treatment of neuropathic pain. *MedGenMed* (2007) **9**(2):36.
- Hayes C, Browne S, Lantry G, Burstall R. Neuropathic pain in the acute pain service: a prospective survey. *Acute Pain* (2002) **4**(2):45–8. doi:10.1016/s1366-0071(02)00026-8
- Nikolajsen L, Black JA, Kroner K, Jensen TS, Waxman SG. Neuroma removal for neuropathic pain: efficacy and predictive value of lidocaine infusion. *The Clin J Pain* (2010) **26**(9):788–93. doi:10.1097/ajp.0b013e3181ed0823
- Souza JM, Cheesborough JE, Ko JH, Cho MS, Kuiken TA, Dumanian GA. Targeted muscle reinnervation: a novel approach to postamputation neuroma pain. *Clin Orthopaedics and Relat Res* (2014) **472**(10):2984–90. doi:10.1007/s11999-014-3528-7
- Calcagni M, Zimmermann S, Scaglioni MF, Giesen T, Giovanoli P, Fakin RM. The novel treatment of SVF-enriched fat grafting for painful end-neuromas of superficial radial nerve. *Microsurgery* (2018) **38**(3):264–9. doi:10.1002/micr.30122
- Lopez-Alvarez VM, Cobiañchi S, Navarro X. Chronic electrical stimulation reduces hyperalgesia and associated spinal changes induced by peripheral nerve injury. *Neuromodulation: Technology Neural Interf* (2019) **22**(5):509–18. doi:10.1111/ner.12927

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- Billet B, Hanssens K, De Coster O, Santos A, Rotte A, Minne V. High-frequency (10 kHz) spinal cord stimulation for the treatment of focal, chronic postsurgical neuropathic pain: results from a prospective study in Belgium. *Pain Manag* (2021) **12**:75–85. doi:10.2217/pmt-2021-0045
- Lee KY, Bae C, Lee D, Kagan Z, Bradley K, Chung JM, et al. Low-intensity, kilohertz frequency spinal cord stimulation differently affects excitatory and inhibitory neurons in the rodent superficial dorsal horn. *Neuroscience* (2020) **428**:132–9. doi:10.1016/j.neuroscience.2019.12.031
- Kato K, Kikuchi S, Shubayev VI, Myers R. Distribution and tumor necrosis factor- α isoform binding specificity of locally administered etanercept into injured and uninjured rat sciatic nerve. *Neuroscience* (2009) **160**(2):492–500. doi:10.1016/j.neuroscience.2009.02.038
- Kremer M, Yalcin I, Goumon Y, Wurtz X, Nexon L, Daniel D, et al. A dual noradrenergic mechanism for the relief of neuropathic allodynia by the antidepressant drugs duloxetine and amitriptyline. *J Neurosci* (2018) **38**(46): 9934–54. doi:10.1523/jneurosci.1004-18.2018
- Suter MR, Papaloizos M, Berde CB, Woolf C, Gilliard N, Spahn D, et al. Development of neuropathic pain in the rat spared nerve injury model is not prevented by a peripheral nerve block. *Anesthesiology* (2003) **99**(6):1402–8. doi:10.1097/0000542-200312000-00025
- Capano A, Weaver R, Burkman E. Evaluation of the effects of CBD hemp extract on opioid use and quality of life indicators in chronic pain patients: a prospective cohort study. *Postgrad Med* (2020) **132**(1):56–61. doi:10.1080/00325481.2019.1685298
- Chaplan SR, Guo HQ, Lee DH, Luo L, Liu C, Kuei C, et al. Neuronal hyperpolarization-activated pacemaker channels drive neuropathic pain. *J Neurosci* (2003) **23**(4):1169–78. doi:10.1523/jneurosci.23-04-01169.2003
- Wu J, Chiu DT. Painful neuromas: a review of treatment modalities. *Ann Plast Surg* (1999) **43**(6):661–7. doi:10.1097/0000637-199912000-00016
- Allen CB, Williamson TK, Norwood SM, Gupta A. Do electrical stimulation devices reduce pain and improve function? A comparative review. *Pain Ther* (2023) **12**(6):1339–54. doi:10.1007/s40122-023-00554-6
- Parisien M, Lima LV, Dagostino C, El-Hachem N, Drury GL, Grant AV, et al. Acute inflammatory response via neutrophil activation protects against the development of chronic pain. *Sci Transl Med* (2022) **14**(644):eabj9954. doi:10.1126/scitranslmed.abj9954
- Decrouy-Duruz V, Christen T, Raffoul W. Evaluation of surgical treatment for neuropathic pain from neuroma in patients with injured peripheral nerves. *J Neurosurg* (2018) **128**(4):1235–40. doi:10.3171/2017.1.jns.161778
- Freeman R. New and developing drugs for the treatment of neuropathic pain in diabetes. *Curr Diab Rep* (2013) **13**(4):500–8. doi:10.1007/s11892-013-0396-6
- Finnerup NB, Sindrup SH, Jensen TS. The evidence for pharmacological treatment of neuropathic pain. *Pain* (2010) **150**(3):573–81. doi:10.1016/j.pain.2010.06.019
- Caruso R, Ostuzzi G, Turrini G, Ballette F, Recla E, Dall'Olio R, et al. Beyond pain: can antidepressants improve depressive symptoms and quality of life in patients with neuropathic pain? A systematic review and meta-analysis. *Pain* (2019) **160**:2186–98. doi:10.1097/j.pain.0000000000001622
- Gisev N, Nielsen S, Campbell G, Santo T, Jr, Mant A, Bruno R, et al. Antidepressant use among people prescribed opioids for chronic noncancer pain. *Pain Med* (2019) **20**:2450–8. doi:10.1093/pm/pnz009

32. Lovaglio AC, Socolovsky M, Di Masi G, Bonilla G. Treatment of neuropathic pain after peripheral nerve and brachial plexus traumatic injury. *Neurol India* (2019) **67**(Suppl. ment):S32–S37. doi:10.4103/0028-3886.250699
33. Watson PCN, Moulin D, Watt-Watson J, Gordon A, Eisenhoffer J. Controlled-release oxycodone relieves neuropathic pain: a randomized controlled trial in painful diabetic neuropathy. *Pain* (2003) **105**(1–2):71–8. doi:10.1016/s0304-3959(03)00160-x
34. Sindrup SH, Andersen G, Madsen C, Smith T, Brøsen K, Jensen TS. Tramadol relieves pain and allodynia in polyneuropathy: a randomised, double-blind, controlled trial. *Pain* (1999) **83**(1):85–90. doi:10.1016/s0304-3959(99)00079-2
35. Harati Y, Gooch C, Swenson M, Edelman S, Greene D, Raskin P, et al. Double-blind randomized trial of tramadol for the treatment of the pain of diabetic neuropathy. *Neurology* (1998) **50**(6):1842–6. doi:10.1212/wnl.50.6.1842
36. Sato R, Sekiguchi M, Konno SI. Acetaminophen combined with tramadol is more effective than acetaminophen or tramadol to reduce neuropathic root pain: an experimental study with application of nucleus pulposus in a rat model. *Eur Spine J* (2020) **29**(1):169–78. doi:10.1007/s00586-019-06190-z
37. Vinik A. Clinical review: use of antiepileptic drugs in the treatment of chronic painful diabetic neuropathy. *The J Clin Endocrinol and Metab* (2005) **90**(8):4936–45. doi:10.1210/jc.2004-2376
38. Backonja MM. Gabapentin monotherapy for the symptomatic treatment of painful neuropathy: a multicenter, double-blind, placebo-controlled trial in patients with diabetes mellitus. *Epilepsia* (1999) **40**:S57–S74. doi:10.1111/j.1528-1157.1999.tb00934.x
39. Vannier JL, Belkheyar Z, Oberlin C, Montravers P. Management of neuropathic pain after brachial plexus injury in adult patients: a report of 60 cases. *Ann Françaises d'Anesthésie de Réanimation* (2008) **27**(11):890–5. doi:10.1016/j.annfar.2008.08.013
40. Sindou MP, Blondet E, Emery E, Mertens P. Microsurgical lesioning in the dorsal root entry zone for pain due to brachial plexus avulsion: a prospective series of 55 patients. *J Neurosurg* (2005) **102**(6):1018–28. doi:10.3171/jns.2005.102.6.1018
41. van Dongen R, Cohen SP, van Kleef M, Mekhail N, Huygen F. 22. Traumatic plexus lesion. *Pain Pract* (2011) **11**(4):414–20. doi:10.1111/j.1533-2500.2011.00451.x
42. Allegri N, Mennuni S, Rulli E, Vanacore N, Corli O, Floriani I, et al. Systematic review and meta-analysis on neuropsychological effects of long-term use of opioids in patients with chronic noncancer pain. *Pain Pract* (2019) **19**(3):328–43. doi:10.1111/papr.12741
43. Guan J, Tanaka S, Kawakami K. Anticonvulsants or antidepressants in combination pharmacotherapy for treatment of neuropathic pain in cancer patients: a systematic review and meta-analysis. *The Clin J Pain* (2016) **32**(8):719–25. doi:10.1097/ajp.0000000000000310
44. Rosenstock J, Tuchman M, LaMoreaux L, Sharma U. Pregabalin for the treatment of painful diabetic peripheral neuropathy: a double-blind, placebo-controlled trial. *Pain* (2004) **110**(3):628–38. doi:10.1016/j.pain.2004.05.001
45. Torrance N, Smith BH, Watson MC, Bennett MI. Medication and treatment use in primary care patients with chronic pain of predominantly neuropathic origin. *Fam Pract* (2007) **24**(5):481–5. doi:10.1093/fampra/cmm042
46. Stein C. New concepts in opioid analgesia. *Expert Opin Investig Drugs* (2018) **27**(10):765–75. doi:10.1080/13543784.2018.1516204
47. Papagianni A, Siedler G, Sommer C, Üçeyler N. Capsaicin 8% patch reversibly reduces A-delta fiber evoked potential amplitudes. *PAIN Rep* (2018) **3**(2):e644. doi:10.1097/pr9.0000000000000644
48. Wolff RF, Bala MM, Westwood M, Kessels A, Kleijnen J. 5% lidocaine medicated plaster in painful diabetic peripheral neuropathy (DPN): a systematic review. *Swiss Med Wkly* (2010) **140**(21–22):297–306. doi:10.4414/smw.2010.12995
49. Schreiber AK, Nones CF, Reis RC, Chichorro JG, Cunha JM. Diabetic neuropathic pain: physiopathology and treatment. *World J Diabetes* (2015) **6**(3):432–44. doi:10.4239/wjd.v6.i3.432
50. Bertoch T, D'Aunno D, McCoun J. Suzetrigine, a non-opioid NaV1.8 inhibitor for treatment of moderate-to-severe acute pain: two phase 3 randomized clinical trials. *Anesthesiology* (2025). doi:10.1097/ALN.0000000000005460
51. Niesters M, Martini C, Dahan A. Ketamine for chronic pain: risks and benefits. *Br J Clin Pharmacol* (2014) **77**(2):357–67. doi:10.1111/bcp.12094
52. Piotrowska A, Starnowska-Sokół J, Makuch W, Mika J, Witkowska E, Tymecka D, et al. Novel bifunctional hybrid compounds designed to enhance the effects of opioids and antagonize the pronociceptive effects of non-opioid peptides as potent analgesics in a rat model of neuropathic pain. *Pain* (2020) **162**:432–45. doi:10.1097/j.pain.0000000000002045
53. American Pain Foundation. Overview of American pain surveys: 2005–2006. *J Pain Palliat Care Pharmacother* (2008) **22**(1):33–8. doi:10.1080/15360280801989344
54. Benyamin R, Trescot AM, Datta S, Buenaventura R, Adlaka R, Sehgal N, et al. Opioid complications and side effects. *Pain Physician* (2008) **11**(2 Suppl. 1):S105–20.
55. Katz WA, Barkin RL. Dilemmas in chronic/persistent pain management. *Disease-a-Month* (2010) **56**(4):233–50. doi:10.1016/j.disamonth.2009.12.006
56. Gross JL, Perate AR, Elkassabany NM. Pain management in trauma in the age of the opioid crisis. *Anesthesiology Clin* (2019) **37**(1):79–91. doi:10.1016/j.andclin.2018.09.010
57. Lin CS, Lin YC, Lao HC, Chen CC. Interventional treatments for postherpetic neuralgia: a systematic review. *Pain Physician* (2019) **22**(3):209–28.
58. Pet MA, Ko JH, Friedly JL, Smith DG. Traction neurectomy for treatment of painful residual limb neuroma in lower extremity amputees. *J Orthopaedic Trauma* (2015) **29**(9):e321–5. doi:10.1097/bot.0000000000000337
59. Zuniga JR, Yates DM. Factors determining outcome after trigeminal nerve surgery for neuropathic pain. *J Oral Maxillofacial Surg* (2016) **74**(7):1323–9. doi:10.1016/j.joms.2016.02.005
60. Ducic I, Yoon J, Eberlin KR. Treatment of neuroma-induced chronic pain and management of nerve defects with processed nerve allografts. *Plast Reconstr Surg - Glob Open* (2019) **7**(12):e2467. doi:10.1097/gox.0000000000002467
61. Mass DP, Ciano MC, Tortosa R, Newmeyer WL, Kilgore ES. Treatment of painful hand neuromas by their transfer into bone. *Plast Reconstr Surg* (1984) **74**(2):182–5. doi:10.1097/00006534-198408000-00002
62. Ives GC, Kung TA, Nghiem BT, Ursu DC, Brown DL, Cederna PS, et al. Current state of the surgical treatment of terminal neuromas. *Neurosurgery* (2018) **83**(3):354–64. doi:10.1093/neuros/nyx500
63. Chappell AG, Jordan SW, Dumanian GA. Targeted muscle reinnervation for treatment of neuropathic pain. *Clin Plast Surg* (2020) **47**(2):285–93. doi:10.1016/j.cps.2020.01.002
64. Dumanian GA, Potter BK, Mioton LM, Ko JH, Cheesborough JE, Souza JM, et al. Targeted muscle reinnervation treats neuroma and phantom pain in major limb amputees: a randomized clinical trial. *Ann Surg* (2019) **270**(2):238–46. doi:10.1097/sla.0000000000003088
65. Janes LE, Fracol ME, Dumanian GA, Ko JH. Targeted muscle reinnervation for the treatment of neuroma. *Hand Clin* (2021) **37**(3):345–59. doi:10.1016/j.hcl.2021.05.002
66. Mauch JT, Kao DS, Friedly JL, Liu Y. Targeted muscle reinnervation and regenerative peripheral nerve interfaces for pain prophylaxis and treatment: a systematic review. *PM&R* (2023) **15**:1457–65. doi:10.1002/pmrj.12972
67. Al-Ajam Y, Woollard A, Kang N. Advances in upper limb loss rehabilitation: the role of targeted muscle reinnervation and regenerative peripheral nerve interfaces. *Plast Aesthet Res* (2022) **9**(63):63. doi:10.20517/2347-9264.2022.24
68. Senger JLB, Hardy P, Thorkelsson A, Duia S, Hsiao R, Kemp SWP, et al. A direct comparison of targeted muscle reinnervation and regenerative peripheral nerve interfaces to prevent neuroma pain. *Neurosurgery* (2023) **93**(5):1180–91. doi:10.1227/neu.0000000000002541
69. Shamoun F, Shamoun V, Akhavan A, Tuffaha SH. Target receptors of regenerating nerves: neuroma formation and current treatment options. *Front Mol Neurosci* (2022) **15**:859221. doi:10.3389/fnmol.2022.859221
70. Valerio IL, Dumanian GA, Jordan SW, Mioton LM, Bowen BJ, West JM, et al. Preemptive treatment of phantom and residual limb pain with targeted muscle reinnervation at the time of major limb amputation. *J Am Coll Surgeons* (2019) **228**(3):217–26. doi:10.1016/j.jamcollsurg.2018.12.015
71. Dellon AL, Mackinnon SE. Treatment of the painful neuroma by neuroma resection and muscle implantation. *Plast Reconstr Surg* (1986) **77**(3):427–36. doi:10.1097/00006534-198603000-00016
72. Roth E, Linehan A, Weihrauch D, Stucky C, Hogan Q, Hoben G. Targeted muscle reinnervation prevents and reverses rat pain behaviors after nerve transection. *Pain* (2023) **164**(2):316–24. doi:10.1097/j.pain.0000000000002702
73. Xie W, Strong JA, Zhang JM. Active nerve regeneration with failed target reinnervation drives persistent neuropathic pain. *eNeuro* (2017) **4**(1):ENEURO.0008–17.2017. doi:10.1523/eneuro.0008-17.2017
74. Serra J, Bostock H, Sola R, Aleu J, García E, Cokic B, et al. Microneurographic identification of spontaneous activity in C-nociceptors in neuropathic pain states in humans and rats. *Pain* (2012) **153**(1):42–55. doi:10.1016/j.pain.2011.08.015
75. Xie W, Strong JA, Meij JTA, Zhang JM, Yu L. Neuropathic pain: early spontaneous afferent activity is the trigger. *Pain* (2005) **116**(3):243–56. doi:10.1016/j.pain.2005.04.017
76. Woolf CJ, Salter MW. Neuronal plasticity: increasing the gain in pain. *Science* (2000) **288**(5472):1765–8. doi:10.1126/science.288.5472.1765

77. Lanier ST, Jordan SW, Ko JH, Dumanian GA. Targeted muscle reinnervation as a solution for nerve pain. *Plast and Reconstr Surg* (2020) **146**(5):651e–63e. doi:10.1097/prs.00000000000007235
78. Peters BR, Russo SA, West JM, Moore AM, Schulz SA. Targeted muscle reinnervation for the management of pain in the setting of major limb amputation. *SAGE Open Med* (2020) **8**:2050312120959180. doi:10.1177/2050312120959180
79. Jankowski MP, Lawson JJ, McIlwraith SL, Rau KK, Anderson CE, Albers KM, et al. Sensitization of cutaneous nociceptors after nerve transection and regeneration: possible role of target-derived neurotrophic factor signaling. *J Neurosci* (2009) **29**(6):1636–47. doi:10.1523/jneurosci.3474-08.2009
80. Pet MA, Ko JH, Friedly JL, Mourad PD, Smith DG. Does targeted nerve implantation reduce neuroma pain in amputees? *Clin Orthopaedics and Relat Res* (2014) **472**(10):2991–3001. doi:10.1007/s11999-014-3602-1
81. Valerio I, Schulz SA, West J, Westenberg RF, Eberlin KR. Targeted muscle reinnervation combined with a vascularized pedicled regenerative peripheral nerve interface. *Plast Reconstr Surg - Glob Open* (2020) **8**(3):e2689. doi:10.1097/gox.0000000000002689
82. Kato N, Htut M, Taggart M, Carlstedt T, Birch R. The effects of operative delay on the relief of neuropathic pain after injury to the brachial plexus: a review of 148 cases. *The J Bone Joint Surg Br volume* (2006) **88-B**(6):756–9. doi:10.1302/0301-620x.88b6.16995
83. Berman JS, Birch R, Anand P. Pain following human brachial plexus injury with spinal cord root avulsion and the effect of surgery. *Pain* (1998) **75**(2-3):199–207. doi:10.1016/s0304-3959(97)00220-0
84. Decosterd I, Woolf CJ. Spared nerve injury: an animal model of persistent peripheral neuropathic pain. *Pain* (2000) **87**(2):149–58. doi:10.1016/s0304-3959(00)00276-1
85. Zhang J, Liu W, Zhang X, Lin S, Yan J, Ye J. Sema3A inhibits axonal regeneration of retinal ganglion cells via ROCK2. *Brain Res* (2020) **1727**:146555. doi:10.1016/j.brainres.2019.146555
86. Wu F, Miao X, Chen J, Sun Y, Liu Z, Tao Y, et al. Down-regulation of GAP-43 by inhibition of caspases-3 in a rat model of neuropathic pain. *Int J Clin Exp Pathol* (2012) **5**(9):948–55.
87. Lister RC, Tsui JM, Naram A. A technical guide for sciatic nerve targeted muscle reinnervation in a transfemoral amputee. *Plast Reconstr Surg - Glob Open* (2022) **10**(9):e4525. doi:10.1097/gox.00000000000004525
88. Shin SE, Haffner ZK, Chang BL, Kleiber GM. A pilot investigation into targeted muscle reinnervation for complex regional pain syndrome, type II. *Plast Reconstr Surg - Glob Open* (2022) **10**(12):e4718. doi:10.1097/gox.00000000000004718
89. Fu CJ, Sun JB, Bi ZG, Wang XM, Yang CL. Evaluation of platelet-rich plasma and fibrin matrix to assist in healing and repair of rotator cuff injuries: a systematic review and meta-analysis. *Clin Rehabil* (2017) **31**(2):158–72. doi:10.1177/0269215516634815
90. Ribeiro Ad G, Riccoli Junior W, Silva AR, Polesello GC, Guimarães RP. Prp in the treatment of trochanteric syndrome: a pilot study. *Acta Ortop Bras* (2016) **24**(4):208–12. doi:10.1590/1413-785220162404159837
91. Verhaegen F, Brys P, Debeer P. Rotator cuff healing after needling of a calcific deposit using platelet-rich plasma augmentation: a randomized, prospective clinical trial. *J Shoulder Elbow Surg* (2016) **25**(2):169–73. doi:10.1016/j.jse.2015.10.009
92. Grassi A, Napoli F, Romandini I, Samuelsson K, Zaffagnini S, Candrian C, et al. Is platelet-rich plasma (PRP) effective in the treatment of acute muscle injuries? A systematic review and meta-analysis. *Sports Med* (2018) **48**(4):971–89. doi:10.1007/s40279-018-0860-1
93. Khan M, Bedi A. Cochrane in CORR ®: platelet-rich therapies for musculoskeletal soft tissue injuries (review). *Clin Orthopaedics and Relat Res* (2015) **473**(7):2207–13. (Review). doi:10.1007/s11999-015-4207-z
94. Fitzpatrick J, Bulsara MK, O'Donnell J, McCrory PR, Zheng MH. The effectiveness of platelet-rich plasma injections in gluteal tendinopathy: a randomized, double-blind controlled trial comparing a single platelet-rich plasma injection with a single corticosteroid injection. *Am J Sports Med* (2018) **46**:933–9. doi:10.1177/0363546517745525
95. Yoshida M, Funasaki H, Marumo K. Efficacy of autologous leukocyte-reduced platelet-rich plasma therapy for patellar tendinopathy in a rat treadmill model. *Muscle Ligaments Tendons J* (2019) **06**(2):205–15. doi:10.32098/mltj.02.2016.07
96. Cook JL, Smith PA, Bozynski CC, Kuroki K, Cook CR, Stoker AM, et al. Multiple injections of leukoreduced platelet rich plasma reduce pain and functional impairment in a canine model of ACL and meniscal deficiency. *J Orthopaedic Res* (2016) **34**(4):607–15. doi:10.1002/jor.23054
97. Dyson-Hudson TA, Hogaboom NS, Nakamura R, Terry A, Malanga GA. Ultrasound-guided platelet-rich plasma injection for the treatment of recalcitrant rotator cuff disease in wheelchair users with spinal cord injury: a pilot study. *The J Spinal Cord Med* (2020) **45**:42–8. doi:10.1080/10790268.2020.1754676
98. Moezi M, Tahririan M, Motifard M, Nemati M, Nemati A. Ultrasound guided platelet-rich plasma injection for the treatment of rotator cuff tendinopathy. *Adv Biomed Res* (2016) **5**:200. doi:10.4103/2277-9175.190939
99. Kim SJ, Kim EK, Kim SJ, Song DH. Effects of bone marrow aspirate concentrate and platelet-rich plasma on patients with partial tear of the rotator cuff tendon. *J Orthop Surg Res* (2018) **13**(1):1. doi:10.1186/s13018-017-0693-x
100. Sengodan VC, Kurian S, Ramasamy R. Treatment of partial rotator cuff tear with ultrasound-guided platelet-rich plasma. *J Clin Imaging Sci* (2017) **7**(32):32. doi:10.4103/jcis.jcis_26_17
101. Sanchez M, Guadilla J, Fiz N, Andia I. Ultrasound-guided platelet-rich plasma injections for the treatment of osteoarthritis of the hip. *Rheumatology* (2012) **51**(1):144–50. doi:10.1093/rheumatology/ker303
102. Patel S, Dhillon MS, Aggarwal S, Marwaha N, Jain A. Treatment with platelet-rich plasma is more effective than placebo for knee osteoarthritis: a prospective, double-blind, randomized trial. *Am J Sports Med* (2013) **41**(2):356–64. doi:10.1177/0363546512471299
103. Wilson JJ, Lee KS, Miller AT, Wang S. Platelet-rich plasma for the treatment of chronic plantar fasciopathy in adults: a case series. *Foot and Ankle Specialist* (2014) **7**(1):61–7. doi:10.1177/1938640013509671
104. Rossi LA, Molina Romoli AR, Bertona Altieri BA, Burgos Flor JA, Scordo WE, Elizondo CM. Does platelet-rich plasma decrease time to return to sports in acute muscle tear? A randomized controlled trial. *Knee Surg Sports Traumatol Arthrosc* (2017) **25**(10):3319–25. doi:10.1007/s00167-016-4129-7
105. Hurley ET, Lim Fat D, Moran CJ, Mullett H. The efficacy of platelet-rich plasma and platelet-rich fibrin in arthroscopic rotator cuff repair: a meta-analysis of randomized controlled trials. *Am J Sports Med* (2018) **47**:753–61. doi:10.1177/0363546517751397
106. Fitzpatrick J, Bulsara M, Zheng MH. The effectiveness of platelet-rich plasma in the treatment of tendinopathy: a meta-analysis of randomized controlled clinical trials. *Am J Sports Med* (2017) **45**(1):226–33. doi:10.1177/0363546516643716
107. Kuffler DP. Platelet-rich plasma and the elimination of neuropathic pain. *Mol Neurobiol* (2013) **48**(2):315–32. doi:10.1007/s12035-013-8494-7
108. Huang SH, Wu SH, Lee SS, Lin YN, Chai CY, Lai CS, et al. Platelet-rich plasma injection in burn scar areas alleviates neuropathic scar pain. *Int J Med Sci* (2018) **15**(3):238–47. doi:10.7150/ijms.22563
109. Anjayani S, Wirohadidjojo YW, Adam AM, Suwandi D, Seweng A, Amiruddin MD. Sensory improvement of leprosy peripheral neuropathy in patients treated with perineural injection of platelet-rich plasma. *Int J Dermatol* (2014) **53**(1):109–13. doi:10.1111/ijd.12162
110. Behrooz Z, Ramezani F, Janzadeh A, Rahimi B, Nasirinezhad F. Platelet-rich plasma in umbilical cord blood reduces neuropathic pain in spinal cord injury by altering the expression of ATP receptors. *Physiol and Behav* (2021) **228**:113186. doi:10.1016/j.physbeh.2020.113186
111. Ikumi A, Hara Y, Okano E, Kohyama S, Arai N, Taniguchi Y, et al. Intraoperative local administration of platelet-rich plasma (PRP) during neurolysis surgery for the treatment of digital nerve crush injury. *Case Rep Orthopedics* (2018) **2018**:1–6. doi:10.1155/2018/1275713
112. Vares P, Dehghan MM, Bastami F, Biazar E, Shamloo N, Heidari Keshel S, et al. Effects of platelet-rich fibrin/collagen membrane on sciatic nerve regeneration. *J Craniofac Surg* (2021) **32**(2):794–8. doi:10.1097/scs.00000000000007003
113. Hibner M, Castellanos ME, Drachman D, Balducci J. Repeat operation for treatment of persistent pudendal nerve entrapment after pudendal neurolysis. *J Minimally Invasive Gynecol* (2012) **19**(3):325–30. doi:10.1016/j.jmig.2011.12.022
114. Dong C, Sun Y, Qi Y, Zhu Y, Wei H, Wu D, et al. Effect of platelet-rich plasma injection on mild or moderate carpal tunnel syndrome: an updated systematic review and meta-analysis of randomized controlled trials. *Biomed Res Int* (2020) **2020**:5089378. doi:10.1155/2020/5089378
115. Hassanien M, Elawamy A, Kamel EZ, Khalifa WA, Abolfadl GM, Roushdy ASI, et al. Perineural platelet-rich plasma for diabetic neuropathic pain, could it make a difference? *Pain Med* (2020) **21**(4):757–65. doi:10.1093/pm/pnz140
116. Hernández G, Ospina-Tascón GA, Damiani LP, Estenssoro E, Dubin A, Hurtado J, et al. Effect of a resuscitation strategy targeting peripheral perfusion status vs serum lactate levels on 28-day mortality among patients with septic shock: the ANDROMEDA-SHOCK randomized clinical trial. *JAMA* (2019) **321**(7):654–64. doi:10.1001/jama.2019.0071
117. Bhatia R, Chopra G. Efficacy of platelet rich plasma via lumbar epidural route in chronic prolapsed intervertebral disc patients-A pilot study. *J Clin Diagn Res : JCDR* (2016) **10**(9):UC05–UC07. doi:10.7860/JCDR/2016/21863.8482

118. Anitua E, Kirchner F. Intradiscal and intra-articular facet infiltrations with plasma rich in growth factors reduce pain in patients with chronic low back pain. *J Craniovertebral Junction Spine* (2016) 7(4):250–6. doi:10.4103/0974-8237.193260
119. Ruiz-Lopez R, Tsai YC. A randomized double-blind controlled pilot study comparing leucocyte-rich platelet-rich plasma and corticosteroid in caudal epidural injection for complex chronic degenerative spinal pain. *Pain Pract* (2020) 20(6):639–46. doi:10.1111/papr.12893
120. Bise S, Dallaudiere B, Pesquer L, Pedram M, Meyer P, Antoun MB, et al. Comparison of interlaminar CT-guided epidural platelet-rich plasma versus steroid injection in patients with lumbar radicular pain. *Eur Radiol* (2020) 30(6):3152–60. doi:10.1007/s00330-020-06733-9
121. Centeno C, Markle J, Dodson E, Stemper I, Hyzy M, Williams C, et al. The use of lumbar epidural injection of platelet lysate for treatment of radicular pain. *J Exp Orthopaedics* (2017) 4(1):38. doi:10.1186/s40634-017-0113-5
122. Auffero D, Vincent H, Sampson S. Regenerative injection treatment in the spine: review and case series with platelet rich plasma. *J Stem Cells Res Rev and Rep* (2015) 2:1019.
123. Becker C, Heidersdorf S, Drewlo S, de Rodriguez SZ, Krämer J, Willburger RE. Efficacy of epidural perineural injections with autologous conditioned serum for lumbar radicular compression: an investigator-initiated, prospective, double-blind, reference-controlled study. *Spine (Phila Pa 1976)* (2007) 32(17):1803–8. doi:10.1097/brs.0b013e3181076514
124. Uzun H, Bitik O, Uzun O, Ersoy US, Aktaş E. Platelet-rich plasma versus corticosteroid injections for carpal tunnel syndrome. *J Plast Surg Hand Surg* (2017) 51(5):301–5. doi:10.1080/2000656x.2016.1260025
125. Hibner M, Castellanos ME, Drachman D, Balducci J. Repeat operation for treatment of persistent pudendal nerve entrapment after pudendal neurolysis. *J Minimally Invasive Gynecol* (2012) 19:325–30. doi:10.1016/j.jmig.2011.12.022
126. Lin CP, Chang KV, Huang YK, Wu WT, Özçakar L. Regenerative injections including 5% dextrose and platelet-rich plasma for the treatment of carpal tunnel syndrome: a systematic review and network meta-analysis. *Pharmaceuticals (Basel)* (2020) 13(3):49. doi:10.3390/ph13030049
127. Kuffler DP. Differing efficacies of autologous platelet-rich plasma in reducing pain following rotator-cuff injury in a single patient. *J Pain Res* (2018) 11:2239–45. doi:10.2147/jpr.s169647
128. Kuffler DP. Variables affecting the potential efficacy of PRP in providing chronic pain relief. *J Pain Res* (2018) 12:109–16. doi:10.2147/jpr.s190065
129. Shao S, Pan R, Chen Y. Autologous platelet-rich plasma for diabetic foot ulcer. *Trends Endocrinol and Metab* (2020) 31(12):885–90. doi:10.1016/j.tem.2020.10.003
130. Castillo TN, Pouliot MA, Kim HJ, Drago J. Comparison of growth factor and platelet concentration from commercial platelet-rich plasma separation systems. *Am J Sports Med* (2011) 39(2):266–71. doi:10.1177/0363546510387517
131. Hsu WK, Mishra A, Rodeo SR, Fu F, Terry MA, Randelli P, et al. Platelet-rich plasma in orthopaedic applications: evidence-based recommendations for treatment. *J Am Acad Orthopaedic Surgeons* (2013) 21(12):739–48. doi:10.5435/jaas-22-08-469
132. Oudelaar BW, Peerbooms JC, Huis in 't Veld R, Vochoeteloo AJ. Concentrations of blood components in commercial platelet-rich plasma separation systems: a review of the literature. *Am J Sports Med* (2019) 47(2):479–87. doi:10.1177/0363546517746112
133. Marx RE. Platelet-rich plasma (PRP): what is PRP and what is not PRP? *Implant Dentistry* (2001) 10(4):225–8. doi:10.1097/00008505-200110000-00002
134. Arora S, Agnihotri N. Platelet derived biomaterials for therapeutic use: review of technical aspects. *Indian J Hematol Blood Transfus* (2017) 33(2):159–67. doi:10.1007/s12288-016-0669-8
135. Dohan Ehrenfest DM, Pinto NR, Pereda A, Jiménez P, Corso MD, Kang BS, et al. The impact of the centrifuge characteristics and centrifugation protocols on the cells, growth factors, and fibrin architecture of a leukocyte- and platelet-rich fibrin (L-PRF) clot and membrane. *Platelets* (2018) 29(2):171–84. doi:10.1080/09537104.2017.1293812
136. Weibrich G, Kleis WK, Hitzler WE, Hafner G. Comparison of the platelet concentrate collection system with the plasma-rich-in-growth-factors kit to produce platelet-rich plasma: a technical report. *The Int J Oral and Maxill Implants* (2005) 20(1):118–23.
137. Schar MO, Diaz-Romero J, Kohl S, Zumstein MA, Nesic D. Platelet-rich concentrates differentially release growth factors and induce cell migration *in vitro*. *Clin Orthopaedics and Relat Res* (2015) 473(5):1635–43. doi:10.1007/s11999-015-4192-2
138. Mazzocca AD, McCarthy MB, Chowaniec DM, Cote MP, Romeo AA, Bradley JP, et al. Platelet-rich plasma differs according to preparation method and human variability. *J Bone Joint Surg* (2012) 94(4):308–16. doi:10.2106/jbjs.k.00430
139. Cavallo C, Roffi A, Grigolo B, Mariani E, Pratelli L, Merli G, et al. Platelet-rich plasma: the choice of activation method affects the release of bioactive molecules. *Biomed Res Int* (2016) 2016:1–7. doi:10.1155/2016/6591717
140. Eppley BL, Woodell JE, Higgins J. Platelet quantification and growth factor analysis from platelet-rich plasma: implications for wound healing. *Plast Reconstr Surg* (2004) 114(6):1502–8. doi:10.1097/01.prs.0000138251.07040.51
141. Kuffler DP. Platelet-rich plasma promotes axon regeneration, wound healing, and pain reduction: fact or fiction. *Mol Neurobiol* (2015) 52(2):990–1014. doi:10.1007/s12035-015-9251-x
142. Belch JJ, McArdle BM, Burns P, Lowe GDO, Forbes CD. The effects of acute smoking on platelet behaviour, fibrinolysis and haemorheology in habitual smokers. *Thromb Haemost* (1984) 51(1):006–8. doi:10.1055/s-0038-1660996
143. Mukamal KJ, Massaro JM, Ault KA, Mittleman MA, Sutherland PA, Lipinska I, et al. Alcohol consumption and platelet activation and aggregation among women and men: the Framingham Offspring Study. *Alcohol Clin and Exp Res* (2005) 29(10):1906–12. doi:10.1097/01.alc.0000183011.86768.61
144. Olas B, Wachowicz B, Saluk-Juszczak J, Zieliński T. Effect of resveratrol, a natural polyphenolic compound, on platelet activation induced by endotoxin or thrombin. *Thromb Res* (2002) 107(3–4):141–5. doi:10.1016/s0049-3848(02)00273-6
145. Williams JK, Clarkson TB. Dietary soy isoflavones inhibit *in-vivo* constrictor responses of coronary arteries to collagen-induced platelet activation. *Coron Artery Dis* (1998) 9(11):759–64. doi:10.1097/00019501-199809110-00009
146. Frary CD, Johnson RK, Wang MQ. Food sources and intakes of caffeine in the diets of persons in the United States. *J Am Diet Assoc* (2005) 105(1):110–3. doi:10.1016/j.jada.2004.10.027
147. Hubbard GP, Wolfram S, Lovegrove JA, Gibbins J. Ingestion of quercetin inhibits platelet aggregation and essential components of the collagen-stimulated platelet activation pathway in humans. *J Thromb Haemost* (2004) 2(12):2138–45. doi:10.1111/j.1538-7836.2004.01067.x
148. Alvarez-Suarez JM, Giampieri F, Tulipani S, Casoli T, Di Stefano G, González-Paramás AM, et al. One-month strawberry-rich anthocyanin supplementation ameliorates cardiovascular risk, oxidative stress markers and platelet activation in humans. *The J Nutr Biochem* (2014) 25(3):289–94. doi:10.1016/j.jnutbio.2013.11.002
149. de Lorgeril M, Renaud S, Mamelle N, Salen P, Martin JL, Monjaud I, et al. Mediterranean alpha-linolenic acid-rich diet in secondary prevention of coronary heart disease. *Lancet* (1994) 343(8911):1454–9. doi:10.1016/s0140-6736(94)92580-1
150. Sudic D, Razmara M, Forslund M, Ji Q, Hjerdahl P, Li N. High glucose levels enhance platelet activation: involvement of multiple mechanisms. *Br J Haematol* (2006) 133(3):315–22. doi:10.1111/j.1365-2141.2006.06012.x
151. Ahmed Y, van Iddekinge B, Paul C, Sullivan MHF, Elder MG. Retrospective analysis of platelet numbers and volumes in normal pregnancy and in pre-eclampsia. *BJOG: An Int J Obstet and Gynaecol* (1993) 100(3):216–20. doi:10.1111/j.1471-0528.1993.tb15233.x
152. Yokogoshi H, Wurtman RJ. Meal composition and plasma amino acid ratios: effect of various proteins or carbohydrates, and of various protein concentrations. *Metabolism* (1986) 35(9):837–42. doi:10.1016/0026-0495(86)90225-8
153. Micheo WF, Foy CA, Kuffler DP. A novel technique restores function while eliminating intractable neuropathic pain in a 71-year-old diabetic patient under challenging injury conditions. *J Reconstr Microsurg Open* (2023) 08(1):e23–e27. doi:10.1055/s-0042-1757323
154. Foy CA, Micheo WF, Kuffler DP. Inducing ulnar nerve function while eliminating claw hand and reducing chronic neuropathic pain. *Plast Reconstr Surg - Glob Open* (2023) 11(4):e4927. doi:10.1097/gox.0000000000004927
155. Micheo WF, Foy CA, Kuffler DP. Novel technique for restoring function and eliminating chronic neuropathic pain to a 71-year-old diabetic patient with a 12 cm peripheral nerve gap repaired 1.3 years post-trauma. *J Reconstr Microsurg Open* (2022). (in press). doi:10.1055/s-0042-1757323
156. Foy CA, Micheo WF, Kuffler DP. Functional recovery following repair of long nerve gaps in senior patient 2.6 Years posttrauma. *Plast Reconstr Surg - Glob Open* (2021) 9(9):e3831. doi:10.1097/gox.0000000000003831
157. Foy C, Micheo W, Kuffler DP. Sensory and motor recovery following the repair of three long nerve gap in a senior patient 2.6 Years post-trauma. *Plast and Reconstr Surg* (2021). (in press). doi:10.1097/GOX.0000000000003831
158. Santiago Figureoa JP. KD. Novel technique for the long-term elimination of chronic excruciating neuropathic pain. *J Pain Management* (2021). (in press). doi:10.1055/s-0042-1757323
159. Kuffler DP, Reyes O, Sosa JJ, Santiago-Figureoa J. Neurological recovery across a 12-cm-long ulnar nerve gap repaired 3.25 years post trauma: case report. *Neurosurgery* (2011) 69(6):E1321–E1326. doi:10.1227/neu.0b013e31822a9fd2

160. Reyes O, Sosa IJ, Santiago J, Kuffler DP. A novel technique leading to complete sensory and motor recovery across a long peripheral nerve gap. *PR Health Sci J* (2007) **26**(3):225–8.
161. Sucuoğlu H, Üstünsoy S. The short-term effect of PRP on chronic pain in knee osteoarthritis. *Agri* (2019) **31**(2):63–9. doi:10.14744/agri.2019.81489
162. Urits I, Viswanath O, Galasso AC, Sottosani ER, Mahan KM, Aiudi CM, et al. Platelet-rich plasma for the treatment of low back pain: a comprehensive review. *Curr Pain Headache Rep* (2019) **23**(7):52. doi:10.1007/s11916-019-0797-6
163. Salomon D, Miloro M, Kolokythas A. Outcomes of immediate allograft reconstruction of long-span defects of the inferior alveolar nerve. *J Oral Maxillofacial Surg* (2016) **74**(12):2507–14. doi:10.1016/j.joms.2016.05.029
164. Barber FA. PRP as an adjunct to rotator cuff tendon repair. *Sports Med Arthrosc Rev* (2018) **26**(2):42–7. doi:10.1097/jsa.0000000000000193
165. Sundman EA, Cole BJ, Fortier LA. Growth factor and catabolic cytokine concentrations are influenced by the cellular composition of platelet-rich plasma. *Am J Sports Med* (2011) **39**(10):2135–40. doi:10.1177/0363546511417792
166. Raeissadat SA, Sedighpour L, Rayegani SM, Bahrami MH, Bayat M, Rahimi R. Effect of platelet-rich plasma (PRP) versus autologous whole blood on pain and function improvement in tennis elbow: a randomized clinical trial. *Pain Res Treat* (2014) **2014**:1–8. doi:10.1155/2014/191525
167. Milants C, Bruyere O, Kaux JF. Response to: comment on responders to platelet-rich plasma in osteoarthritis: a technical analysis. *Biomed Res Int* (2018) **2018**:1–2. doi:10.1155/2018/2718156
168. Kaux JF, Bouvard M, Lecut C, Oury C, Gothot A, Sanchez M, et al. Reflections about the optimisation of the treatment of tendinopathies with PRP. *Muscle Ligaments Tendons J* (2019) **05**(1):1–4. doi:10.32098/mltj.01.2015.01
169. Yamaguchi R, Terashima H, Yoneyama S, Tadano S, Ohkohchi N. Effects of platelet-rich plasma on intestinal anastomotic healing in rats: PRP concentration is a key factor. *J Surg Res* (2012) **173**(2):258–66. doi:10.1016/j.jss.2010.10.001
170. Laver L, Marom N, Dnyanesh L, Mei-Dan O, Espregueira-Mendes J, Gobbi A. PRP for degenerative cartilage disease: a systematic review of clinical studies. *Cartilage* (2017) **8**(4):341–64. doi:10.1177/1947603516670709
171. Giusti I, D'Ascenzo S, Mancò A, Di Stefano G, Di Francesco M, Rugghetti A, et al. Platelet concentration in platelet-rich plasma affects tenocyte behavior *in vitro*. *Biomed Res Int* (2014) **2014**:630870. doi:10.1155/2014/630870
172. Santiago-Figueroa J, Sosa IJ, Reyes O. Reducing and eliminating human neuropathic pain following peripheral nerve trauma. *J of Pain Management. J Pain Management* (2011) **4**(4):387–94.
173. Golebiewska EM, Poole AW. Platelet secretion: from haemostasis to wound healing and beyond. *Blood Rev* (2015) **29**(3):153–62. doi:10.1016/j.blre.2014.10.003
174. Lindemann S, Tolley ND, Dixon DA, McIntyre TM, Prescott SM, Zimmerman GA, et al. Activated platelets mediate inflammatory signaling by regulated interleukin 1 β synthesis. *The J Cell Biol* (2001) **154**(3):485–90. doi:10.1083/jcb.200105058
175. Weyrich AS, Schwartz H, Kraiss LW, Zimmerman G. Protein synthesis by platelets: historical and new perspectives. *J Thromb Haemost* (2009) **7**(2):241–6. doi:10.1111/j.1538-7836.2008.03211.x
176. Kniewallner KM, Grimm N, Humpel C. Platelet-derived nerve growth factor supports the survival of cholinergic neurons in organotypic rat brain slices. *Neurosci Lett* (2014) **574**:64–9. doi:10.1016/j.neulet.2014.05.033
177. Jonnalagadda D, Izu LT, Whiteheart SW. Platelet secretion is kinetically heterogeneous in an agonist-responsive manner. *Blood* (2012) **120**(26):5209–16. doi:10.1182/blood-2012-07-445080
178. Mariani E, Roffi A, Cattini L, Pulsatelli L, Assirelli E, Krishnakumar GS, et al. Release kinetic of pro- and anti-inflammatory biomolecules from platelet-rich plasma and functional study on osteoarthritis synovial fibroblasts. *Cytotherapy* (2020) **22**(7):344–53. doi:10.1016/j.jcyt.2020.02.006
179. Shen KF, Zhu HQ, Wei XH, Wang J, Li YY, Pang RP, et al. Interleukin-10 down-regulates voltage gated sodium channels in rat dorsal root ganglion neurons. *Exp Neurol* (2013) **247**:466–75. doi:10.1016/j.expneurol.2013.01.018
180. Estacion M, Waxman SG. The response of Na(V)1.3 sodium channels to ramp stimuli: multiple components and mechanisms. *J Neurophysiol* (2013) **109**(2):306–14. doi:10.1152/jn.00438.2012
181. Hains BC, Klein JP, Saab CY, Craner MJ, Black JA, Waxman SG. Upregulation of sodium channel Nav1.3 and functional involvement in neuronal hyperexcitability associated with central neuropathic pain after spinal cord injury. *J Neurosci* (2003) **23**(26):8881–92. doi:10.1523/jneurosci.23-26-08881.2003
182. Cummins TR, Sheets PL, Waxman SG. The roles of sodium channels in nociception: implications for mechanisms of pain. *Pain* (2007) **131**(3):243–57. doi:10.1016/j.pain.2007.07.026
183. Burchiel KJ. Carbamazepine inhibits spontaneous activity in experimental neuromas. *Exp Neurol* (1988) **102**(2):249–53. doi:10.1016/0014-4886(88)90101-x
184. Matzner O, Devor M. Hyperexcitability at sites of nerve injury depends on voltage-sensitive Na⁺ channels. *J Neurophysiol* (1994) **72**(1):349–59. doi:10.1152/jn.1994.72.1.349
185. Devor M, Wall PD, Catalan N. Systemic lidocaine silences ectopic neuroma and DRG discharge without blocking nerve conduction. *Pain* (1992) **48**(2):261–8. doi:10.1016/0304-3959(92)90067-1
186. Omana-Zapata I, Khabbaz MA, Hunter JC, Clarke DE, Bley KR. Tetrodotoxin inhibits neuropathic ectopic activity in neuromas, dorsal root ganglia and dorsal horn neurons. *Pain* (1997) **72**(1-2):41–9. doi:10.1016/s0304-3959(97)00012-2
187. Chabal C, Jacobson L, Little J. Intrathecal fentanyl depresses nociceptive flexion reflexes in patients with chronic pain. *Anesthesiology* (1989) **70**(2):226–9. doi:10.1097/0000542-198902000-00008
188. Kerr BJ, Souslova V, McMahon SB, Wood JN. A role for the TTX-resistant sodium channel Nav 1.8 in NGF-induced hyperalgesia, but not neuropathic pain. *Neuroreport* (2001) **12**(14):3077–80. doi:10.1097/00001756-2001110080-00019
189. Lai J, Gold MS, Kim CS, Bian D, Ossipov MH, Hunter JC, et al. Inhibition of neuropathic pain by decreased expression of the tetrodotoxin-resistant sodium channel, Nav1.8. *Pain* (2002) **95**(1-2):143–52. doi:10.1016/s0304-3959(01)00391-8
190. Samad OA, Tan AM, Cheng X, Foster E, Dib-Hajj SD, Waxman SG. Virus-mediated shRNA knockdown of Na(v)1.3 in rat dorsal root ganglion attenuates nerve injury-induced neuropathic pain. *Mol Ther* (2013) **21**(1):49–56. doi:10.1038/mt.2012.169
191. Ledeboer A, Jekich BM, Sloane EM, Mahoney JH, Langer SJ, Milligan ED, et al. Intrathecal interleukin-10 gene therapy attenuates paclitaxel-induced mechanical allodynia and proinflammatory cytokine expression in dorsal root ganglia in rats. *Brain Behav Immun* (2007) **21**(5):686–98. doi:10.1016/j.bbi.2006.10.012
192. Yao MZ, Gu JF, Wang JH, Sun LY, Lang MF, Liu J, et al. Interleukin-2 gene therapy of chronic neuropathic pain. *Neuroscience* (2002) **112**(2):409–16. doi:10.1016/s0306-4522(02)00078-7
193. Gonzalez-Darder JM, Barbera J, Abellan MJ. Effects of prior anaesthesia on autotomy following sciatic transection in rats. *Pain* (1986) **24**(1):87–91. doi:10.1016/0304-3959(86)90029-1
194. Dokka S, Shi X, Leonard S, Wang L, Castranova V, Rojanasakul Y. Interleukin-10-mediated inhibition of free radical generation in macrophages. *Am J Physiology-Lung Cell Mol Physiol* (2001) **280**(6):L1196–202. doi:10.1152/ajplung.2001.280.6.L1196
195. Linke B, Schreiber Y, Picard-Willems B, Slattery P, Nüsing RM, Harder S, et al. Activated platelets induce an anti-inflammatory response of monocytes/macrophages through cross-regulation of PGE2 and cytokines. *Mediators Inflamm* (2017) **2017**:1–14. doi:10.1155/2017/1463216
196. Zheng W, Huang W, Liu S, Levitt RC, Candiotti KA, Lubarsky DA, et al. IL-10 mediated by herpes simplex virus vector reduces neuropathic pain induced by HIV gp120 combined with ddC in rats. *Mol Pain* (2014) **10**:49. doi:10.1186/1744-8069-10-49
197. Plunkett JA, Yu CG, Easton JM, Bethea JR, Yezierski RP. Effects of interleukin-10 (IL-10) on pain behavior and gene expression following excitotoxic spinal cord injury in the rat. *Exp Neurol* (2001) **168**(1):144–54. doi:10.1006/exnr.2000.7604
198. Yu ML, Wei RD, Zhang T, Wang J, Cheng Y, Qin F, et al. Electroacupuncture relieves pain and attenuates inflammation progression through inducing IL-10 production in CFA-induced mice. *Inflammation* (2020) **43**(4):1233–45. doi:10.1007/s10753-020-01203-2