



OPEN ACCESS

*CORRESPONDENCE

Dhirendra Singh,
✉ dhirendra.singh246@gmail.com
Abidemi James Akindele,
✉ jakindele@unilag.edu.ng

RECEIVED 28 March 2025

ACCEPTED 13 August 2025

PUBLISHED 03 September 2025

CITATION

Singh D, Oladimeji-Salami JA and Akindele AJ (2025) Unraveling the pharmacological and therapeutic potential of Ranolazine beyond antianginal drug use: a new insight. *Exp. Biol. Med.* 250:10604. doi: 10.3389/ebm.2025.10604

COPYRIGHT

© 2025 Singh, Oladimeji-Salami and Akindele. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Unraveling the pharmacological and therapeutic potential of Ranolazine beyond antianginal drug use: a new insight

Dhirendra Singh^{1*}, Joy Awulika Oladimeji-Salami² and Abidemi James Akindele^{3*}

¹Department of Pharmacology, M.M College of Pharmacy, Maharishi Markandeshwar (Deemed to be University), Mullana, Ambala, Haryana, India, ²Special Duties Department, National Biotechnology Development Agency, Abuja, Nigeria, ³Department of Pharmacology, Therapeutics and Toxicology, Faculty of Basic Medical Sciences, College of Medicine, University of Lagos, Lagos, Nigeria

Abstract

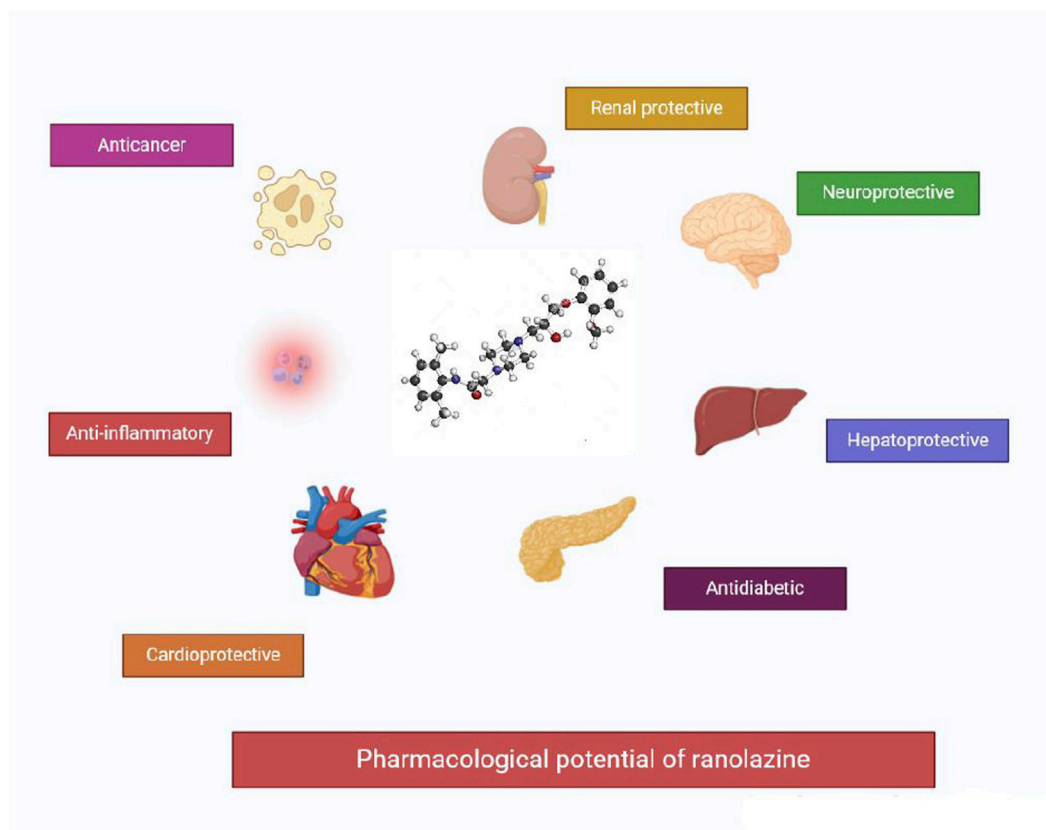
Ranolazine (RAN) is an acetanilide and piperazine derivative that selectively blocks the late sodium current in cardiac cells and is prescribed in adults as an add-on medication for the symptomatic management of patients with stable angina pectoris who are insufficiently managed or intolerant of first-line antianginal treatments. RAN was first approved by the U.S. Food and Drug Administration (FDA) in 2006 and the European Medicine Agency in 2008 for the treatment of chronic stable angina. RAN has no substantial effect on hemodynamic indicators, including heart rate and blood pressure. RAN also slows fatty acid oxidation, which increases glucose oxidation, lowers lactic acid generation, and optimizes heart performance. Besides its antianginal effect, RAN has recently revealed additional pharmacological properties such as neuroprotective, hepatoprotective, renal protective, cardioprotective, and antidiabetic effects and other beneficial pharmacological activities. We choose to write this current review paper to address the many hidden pharmacological and therapeutic potentials of RAN beyond its antianginal activity.

KEYWORDS

anticancer, cardioprotective, renalprotective, neuroprotective, hepatoprotective

Impact statement

Drug re-purposing, finding new therapeutic applications for old or existing drugs, provides the avenue to increase the therapeutic options for the treatment of disease conditions with the possible benefit of enhanced efficacy and safety profile. Beyond its antianginal action, Ranolazine exhibits a variety of pharmacological actions which can be explored for therapeutic benefits. This review extensively sheds light on a number of these pharmacological actions to broaden knowledge and spheres of potential therapeutic applications of Ranolazine.



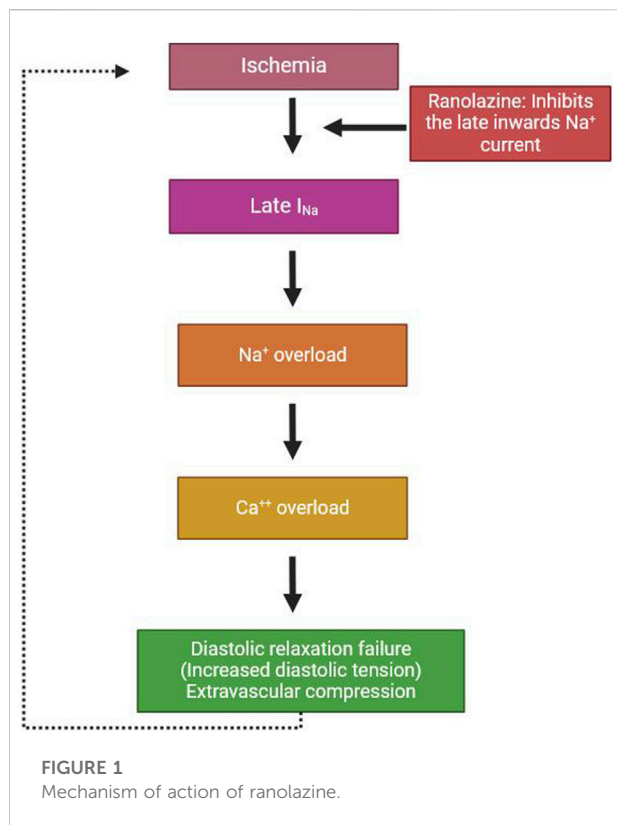
GRAPHICAL ABSTRACT

Pharmacological potential of ranolazine.

Introduction

Ranolazine (RAN) is N-(2, 6-dimethyl phenyl)-4(2-hydroxy-3-[2-methoxyphenoxy] - propyl)-1-piperazine acetamide dihydrochloride. It is an active piperazine whose anti-ischemic effect was originally attributed to the selective inhibition of fatty acid oxidation with a consequent shift of metabolism to more energy-efficient glucose oxidation [1]. An alternative mechanism of action proposed in the past for RAN was the inhibition of β_1 and β_2 adrenoceptors [2]. However, this mechanism (which is associated with sympathetic nervous system regulation of heart rate and contractility) is less prominent compared to RAN's primary action on cardiac ion channels. It is a less significant involvement at the therapeutic concentration of RAN for the treatment of angina, with the main mechanism being linked to inhibition of the late sodium current in cardiac myocytes. This effect reduces intracellular calcium overload and improves myocardial relaxation and oxygen efficiency. At the clinical level, RAN decreases the current of sodium and potassium ion channels. It has been well studied that inhibition of the late phase

of the inward sodium current occurs during cardiac repolarization [3]. In pathological conditions, a rise in calcium ion concentrations contributes to increased sodium-calcium interaction, which induces an increase in the cytosolic calcium concentration [4]. Calcium overload is thought to be the factor that induces reduced left ventricular relaxation during moderate ischemia as well as reperfusion. Increased left ventricular diastolic wall stress compromises myocardial tension circulation, which continues to rise still. Moreover, calcium overload has harmful impacts on myocardial electrical activity, predisposing to ventricular tachycardia [5]. Although this mechanism has been well studied mainly in rodents, the anti-ischemic activity of RAN due to late Na-channel suppression of myocardial perfusion lacks evidence to support this mechanism in patients with ischemic heart disorders. RAN slows the delayed rectifying K^+ current at therapeutic doses and enhances the Q-T interval [6]. The total effect of RAN on the action potential period is equilibrium between the combined effects of rectifier potassium current as well as late sodium current suppression, which prolongs the QT interval by 2–6 ms [7]. Figure 1 shows the mechanism of action of RAN.



The pharmacologically induced attenuation of the late sodium current enhances cardiac diastolic relaxation by decreasing diastolic wall stress. This ultimately results in an improvement of segmental myocardial ischemia.

RAN was first used in therapeutic settings over 25 years ago. It is widely used to treat some disorders and is safe and effective in many cases. Many preclinical and clinical experiments show that RAN may exert cellular protective effects by specifically suppressing the late sodium inward current (late i_{Na}). In the past few years, RAN has been associated with numerous positive properties, such as anticancer, renoprotective, hepatoprotective, neuroprotective, cardioprotective, analgesic, and anti-inflammatory activity, and other benefits independent of its antianginal function.

RAN modulates several cellular pathways like TNF- α , NF- κ B, Capase-3, IL-1 β , IL-6, PPAR- γ , Bax bcl-2, Notch2/Hes1, AKT-eNOS, COX-2, and ERK, which is activity independent of its cardiac protective mechanism.

Search strategies

The literature search was done on multiple electronic databases. These include Web of Science, PubMed, Scopus, and Google Scholar. Appropriate search terms and combinations were used, including ranolazine, pharmacokinetics, neuroprotective,

hepatoprotective, renoprotective, cardioprotective, and antidiabetic effects.

Pharmacokinetics

RAN is available as an oral tablet for therapeutic use and as an intravenous formulation for experimental application. Initially, oral RAN was evaluated as an instant release (IR) formulation. RAN IR has an overall terminal removal half-life of 1.4–1.9 h and a 10-fold peak-trough gap of 240–400 mg three times per day [8]. RAN is now commonly available as a sustained-release (SR) formulation with a more extended absorption phase, with a maximal plasma concentration (C_{max}) usually seen 4–6 h after oral administration and an estimated apparent total elimination half-life of 7 h after steady state. The peak-trough difference at 500–1000 mg twice/day is only 1.6-fold, which is much improved over that of the IR formulation [9–11]. The steady state is usually reached within 3 days of twice-daily dosing. RAN plasma amounts that are clinically beneficial for chronic angina range from 2 to 6 μ mol/L [12, 13]. The oral bioavailability of RAN is 30%–55% and is not influenced by food. RAN is approximately 65% bound to serum protein, mainly α 1-acid glycoprotein [14]. RAN is mainly cleared by the liver metabolic enzyme cytochrome P450 (CYP) 3A4 (70–85%) and is a substrate of P-glycoprotein. Additional processes include CYP2D6 metabolism (10–15 percent), glucuronidation (<5 percent), and renal excretion of unchanged RAN (<5 percent) [8].

Anticancer effects

Driffort et al. found that RAN repressed the pro-invasive shape of human breast cancer MDA-MB-231 cells and decreased the localized extracellular matrix degradation activity [15]. Qiu et al. and Lee et al. validated similar findings and discovered that the anti-invasive action might occur independently of proliferation [16, 17]. Qiu and his group found that RAN's anti-invasive activity was dose-related, with concentrations as low as 2.5 μ M during hypoxia [16]. Guzel et al. found that, in human colorectal cancer cells, (i) hypoxia markedly increased Matrigel invasion and (ii) therapeutic dosages of RAN decreased invasiveness without compromising proliferative ability or cell survival [18].

Rizaner and colleagues demonstrated that for robust metastatic rat prostate cancer Mat-LyLu cells, RAN (i) hindered Matrigel migration under both normoxic and hypoxic circumstances and (ii) decreased the proportion of cells in the lung metastases showing Nav1.7 [19]. Pemmireddy and team examined the anticancer action of RAN on 1,2-Dimethyl hydrazine (DMH)-induced colon cancer in mice and found that RAN substantially reduced colon cancer in

mice, most likely because of cancer cell growth deregulations [20]. Using the Dunning model of rat prostate cancer, Bugan and coworkers demonstrated in double-blind tests that gavage administration of 2.5–5 μM RAN inhibited lung metastasis by as much as 63% [21]. Guth et al. demonstrated that RAN (i) inhibited tumor development and (ii) boosted anti-cancer immunity, as shown by reduced tumor CD8⁺ T-cells Tim3 content, enhanced macrophages, and lowered blood myeloid immunosuppressive monocytes in the TRAMPC1 genetic mice model of prostate cancer [22]. Lastly, Lasheras-Otero et al. demonstrated that RAN inhibited liver metastases in a mouse model of melanoma [23].

Cardioprotective effects

Tocchetti et al. revealed that RAN could avert doxorubicin-induced cardiac failure in mice and HL-1 cardiomyocytes via lowering ROS production [24]. Furthermore, RAN has been shown to mitigate cardiac dysfunction induced by trastuzumab, which is believed to mediate its activity by inhibiting the generation of ROS [25]. De Lorenzo and teammates found that RAN mitigated not just the cardiotoxic adverse effects of trastuzumab but also of pertuzumab and trastuzumab-emtansine (TDM1) when employed in combinatorial therapies both *in vitro* and *in vivo* [26]. Cappetta et al. conducted an experiment using RAN and stated that it could protect cardiomyocytes from doxorubicin-caused oxidative damage [27]. RAN could attenuate MTX-caused oxidative damage in H9c2 cardiomyocytes by reducing MDA, LOOH, AOPPs, and XO activity, maintaining T-SH, CAT, and TAC levels, and prohibiting the HIF-1 α inflammatory cascade [28]. Jiang et al. reported that therapy with RAN in Phospholamban (PLN) knockout hiPSCs-CMs could significantly repair Ca²⁺ handling abnormalities and cellular energy metabolism, thus alleviating the PLN knockout phenotype of HF [29].

In high glucose-treated cardiac fibroblasts, RAN decreased pyroptosis, prevented collagen deposition, and enhanced heart function via enhancing miR-135b expression [30]. Furthermore, RAN protected against diabetic cardiomyopathy-induced apoptosis in rats via activation of the NOTCH1/NRG1 signaling cascade [31]. Tawfik and team showed that RAN administration ameliorated the isoprenaline-mediated myocardial damage in both nondiabetic and diabetic rats by improving histopathological scores, reducing apoptotic markers, and modulating AMPK activity [32]. Le DE and his team proved that RAN increased both resting and stress-induced cardiac adenosine levels and caused small-vessel vasodilation, which improved ischemia in dogs [33]. RAN also showed a positive effect on cardiomyocytes subjected to ischemia/reperfusion, but only when used during ischemia, and this effect is accomplished through improving calcium regulation during ischemia [34].

Tantray et al confirmed that RAN had a protective role in myocardial infarction, similar to ischemic preconditioning facilitators, via promoting myocardial Nitric oxide, Adenosine, Bradykinin, and K⁺ATPase levels in an isolated heart [35]. In anaesthetized rabbits subjected to ischemia and reperfusion, RAN lowered infarct size and raised salvage area index, activating a process similar to PreC and PostC that required activation of the RISK axis [36]. Feng and co-workers demonstrated that chronic RAN treatment effectively reduced the increased concentrations of NE and BNP-45 caused by CHF and improved LV function in CHF rats [37]. RAN increased cardiac function and decreased the level of heart injury in rats with congestive heart failure, which is likely due to the activation of AKT phosphorylation [38]. RAN attenuates pressure overload-mediated cardiac hypertrophy and systolic and diastolic activity by restoring Na⁺ and Ca²⁺ handling, preventing downstream hypertrophic pathways, and reducing ER stress [39].

In an animal model of heart failure, RAN ameliorated cardiac remodeling and improved systolic and diastolic performance by normalizing Ca²⁺ storage [40]. Coppini and colleagues showed that acute RAN treatment lowered intracellular Na⁺ and Ca²⁺ levels as well as CaMKII activity, which contributed to the decrease in hypertrophic cardiomyopathy-associated cardiac remodeling and myocardial dysfunction [41]. Moreover, RAN treatment decreased oxidative stress and alleviated diastolic dysfunction in rats fed a high-salt diet to develop hypertension [42].

Williams and co-workers demonstrated that RAN was efficient in lowering diastolic dysfunction in spontaneously hypertensive rats, and its mechanism of action was associated with suppression of the enhanced late sodium current in the SHR, resulting in decreased Ca²⁺ overload [43]. Le et al. proposed that RAN elevated adenosine concentrations in coronary veins in anaesthetized dogs, both at rest and during dobutamine-caused myocardial ischemia, mostly via enhancing the function of the cytosolic-5'-nucleotidase enzyme [33].

In individuals with CCS, RAN has been proposed as a way to increase myocardial perfusion and lessen mechanical compression of coronary microcirculation [44]. RAN enhanced coronary flow reserve in 58 patients with angina and myocardial ischemia but no obstructive coronary artery disease. This was likely because it improved abnormal coronary autoregulation, which decreased the baseline diastolic coronary flow rate and elevated the hyperemic diastolic coronary flow rate [45]. Furthermore, angina was found to improve when RAN was given in comparison to a placebo in a small trial involving women who had angina, signs of myocardial ischemia, but no obstructive coronary artery disease (CAD). There was also a trend towards improvement in the anomalies of myocardial perfusion detected by cardiac magnetic resonance imaging (CMR imaging). Additionally, compared to women with CFR >3.0, those with CFR \leq 3.0 had a markedly

increased myocardial perfusion reserve index (MPRI) while using RAN versus placebo [46]. RAN therapy also increases arginine plasma values and reduces oxidative stress in a randomized controlled study of 20 patients with unstable angina pectoris and acute cardiac ischemia [47].

Chou and colleagues discovered that RAN notably reduced action potential time, Cai transient time, and Cai decay duration, improved conduction inhomogeneity, and repressed arrhythmogenic alternans induction in db/db mouse hearts with acute IR damage [48]. Wolfes et al. studied the impact of RAN paired with various selective NCX-blockers in an isolated whole-heart model of AF in rabbits and discovered that both combinations extended aERP and aPRR and thereby reduced the development of AF [49]. In levosimendan-produced atrial fibrillation, RAN has a prominent antiarrhythmic effect, and the primary mechanism is a slight delay in repolarization and refractory period, which preserves the atrial myocardium against premature excitement and atrial fibrillation in rabbits [50]. Additionally, RAN appeared to have a dose-dependent antiarrhythmic impact on pacing-induced reentrant ventricular arrhythmias during the late phase of myocardial infarction in anaesthetized rabbits [51].

Markandeya et al. revealed that RAN inhibited late INa, which shortened APD and abolished triggered activity in Lmna (N195K/N195K) ventricular myocytes [52]. RAN has been found to enhance redox balance and mitochondrial activity in the atrium of rats suffering from acetylcholine-CaCl₂-mediated atrial fibrillation [53]. RAN reduced delayed repolarization, aberrant electrical activity, and greater late sodium currents in elderly rats continuously exposed to low testosterone, all of which encouraged maladaptive electrical remodeling in ventricular myocytes [54].

Mustroph and colleagues investigated the beneficial effect of RAN on ethanol-induced atrial fibrillation and discovered that it efficiently suppressed atrial fibrillation by altering the activity of the CaMKII-dependent NaV1.5 channel [55]. RAN also inhibited electrical remodeling, causing atrial fibrillation in HL-1 atrial myocytes through modification of the PI3K/Akt signaling axis [56]. Opacic and his group emphasized that RAN effectively lengthened the atrial effective refractory time and lowered the atrial conduction rate at baseline and after 2 days of AF in a goat model of lone AF [57]. RAN was also compared with vernakalant for cardioversion of acutely produced AF in 15 rabbit hearts. AF was produced with atrial burst pacing and acetylcholine/isoproterenol. RAN besides vernakalant showed equal efficacy in preventing AF [58].

Similarly, recent research in horses found that, in comparison with single medications, the combination of dofetilide and RAN improved the antiarrhythmic effects on acutely generated AF, influencing cardioversion time, susceptibility at AF, and AF latency [59]. The combination of RAN and ivabradine has been evaluated in AF in pigs and the combined effect of these two drugs reduced ventricular rate via decreasing conduction at

the AV node (increased A-H period) and minimizing the dominant AF frequency [60].

RAN was tested to assess its effects in a canine model of heart disease. It blocks atrial fibrillation in animals by lengthening the atrial refractory duration and atrial conduction time. No pro-arrhythmic influence was apparent on the ventricle [61]. Also, RAN administration avoided VT in the porcine model of catecholaminergic polymorphic ventricular tachycardia and decreased the T-wave length [62]. RAN has also been found to be non-inferior to lidocaine and sotalol in avoiding ischemia-reperfusion-induced ventricular tachycardia in a rat model [63]. Malavaki and team examined the vasorelaxant action of RAN and nicardipine on the rabbit aorta. Researchers found that RAN has a synergistic interaction with nicardipine to trigger vasorelaxation in rabbit aortas [64]. RAN inhibited the occurrence and minimized the duration of action potentials in HL-1 cells, resulting in an antiarrhythmic response [65].

In another study, RAN reduced HOCI-LDL-associated alterations in cardiac contractility and electrophysiology, including arrhythmias in primary cardiomyocytes [66]. Del-Canto et al. found that RAN ameliorated the electrophysiological effects responsible for the stretch-induced modification of HL-1 cell fibrillatory activation patterns by altering the rise in activation rate and preserving the magnitude of activation [67]. RAN modified the ECG abnormalities, diminished Ca²⁺ sparks and abnormal waves, lowered the *in vitro* events and the frequency of arrhythmias noticed in isolated cardiomyocytes of hypothyroid mice [68]. Two preclinical studies of RAN demonstrate promise in preventing long QT syndrome in rats. RAN suppressed QT prolongation, prevented early after depolarizations, and reduced the duration of torsades de pointes [69, 70].

RAN showed antiarrhythmic efficacy against AT (Atrial Tachycardia) elicited by rapid burst stimulation in anaesthetized rabbits [71]. Nuno and his team examined the anti-atrial fibrillatory effect and pharmacological safety characteristics of RAN in halothane-anesthetized dogs. Researchers found that RAN had little effect on ventricular early repolarization *in vivo*, but it did extend late repolarization with no danger of re-entrant arrhythmias [72].

Wolfes and colleagues evaluated the impact of RAN in combination with several specific NCX-blockers in an isolated whole-heart AF model. Both combinations increased the atrial effective refractory time while decreasing the frequency of AF episodes [49]. Aidonidis et al. investigated whether co-treatment of RAN-AMIO would show additive antiarrhythmic effects. RAN notably improved the propagation duration of fast atrial depolarizations and enhanced the AMIO-mediated mild elevations in aPRR [73]. Miranda and co-workers explored the influence of RAN on healthy cardiomyocytes as well as a cellular model of type 3 long QT syndromes (LQT3). RAN had a small effect on sarcomere shortening in healthy ENDO and EPI cells, and it reduced arrhythmias caused by INaL to the same rate as ENDO and EPI cells [74].

Eleclazine and RAN reduced the AF window and AF burden in association with the inhibition of both endogenous and enhanced atrial late I_{Na} with half maximal inhibitory concentrations (IC_{50}) of 1.14 and 9.78 μM and 0.94 and 8.31 μM , respectively [75]. RAN normalized AV-conduction in *Scn5a1798insD/+* mice by preventing the mutation-induced increase in intracellular sodium ($[Na^+]_i$) and calcium ($[Ca^{2+}]_i$) concentrations [76]. RAN also inhibited TASK-1 channels, and inhibition of TASK-1 may contribute to the observed antiarrhythmic effects of RAN [3]. RAN suppressed CaT alternans and decreased the Ca^{2+} -voltage coupling gain in a dog HF model, reducing arrhythmogenic cardiac alternans [77]. RAN has continued to yield amazing outcomes, such as the cessation of acutely caused AF in horses via cardioversion [78]. RAN partially prevented action potential and QT interval prolongation in 4-week-old *Scn5a^{+/ΔQKP}* mice and suppressed arrhythmias [79].

Ke and colleagues examined how Ca^{2+} homeostasis was affected in CKD mice and discovered that RAN, by controlling CaMKII, PLB, and late Na^+ current, reduced the length of the QT interval and the development of cardiac arrhythmogenesis [80]. Huang et al. investigated the role of FGF23 in activating the I_{Na} -Late, resulting in calcium imbalance and increasing PV arrhythmogenesis, and found that RAN-reduced FGF23 enhanced beating rates, calcium fluctuations, and mitochondrial ROS in PV cardiomyocytes [81].

In human atrial myocytes, RAN alone or when combined with low-dose dronedarone prolonged APD, increased cellular hyperpolarization, and decreased SR Ca^{2+} leakage [82]. RAN has been observed to possess a similar effect to mexiletine in terms of action potential period shortening, with less paradoxical action potential duration prolongation in LQT3 mutant cells [83]. In the rabbit heart model, RAN perfusion substantially decreased the number of breakthrough-type excitations (BEs) in the ischemic border zone (BZ) and mitigated ischemia-caused shortening of action potentials in the BZ without influencing conduction velocity, most likely because of I_{Kr} repression [84]. RAN also decreased VT load and implanted cardioverter-defibrillator (ICD) shocks in 11/12 individuals receiving drug-refractory shocks [85]. RAN also proved to be effective, well-tolerated, and safe in reducing ventricular arrhythmia episodes and ICD interventions in patients with recurrent antiarrhythmic drug-refractory events [86]. After analyzing a group of AF patients on RAN, Black-Maier et al. discovered that the medication is linked to decreased AF DF but not altered organization index or fibrillatory wave amplitude [87]. RAN, a late I (Na) blocker, appeared to possess antiarrhythmic effects, according to continuous ECG monitoring of patients admitted for acute coronary syndrome within the first week [88].

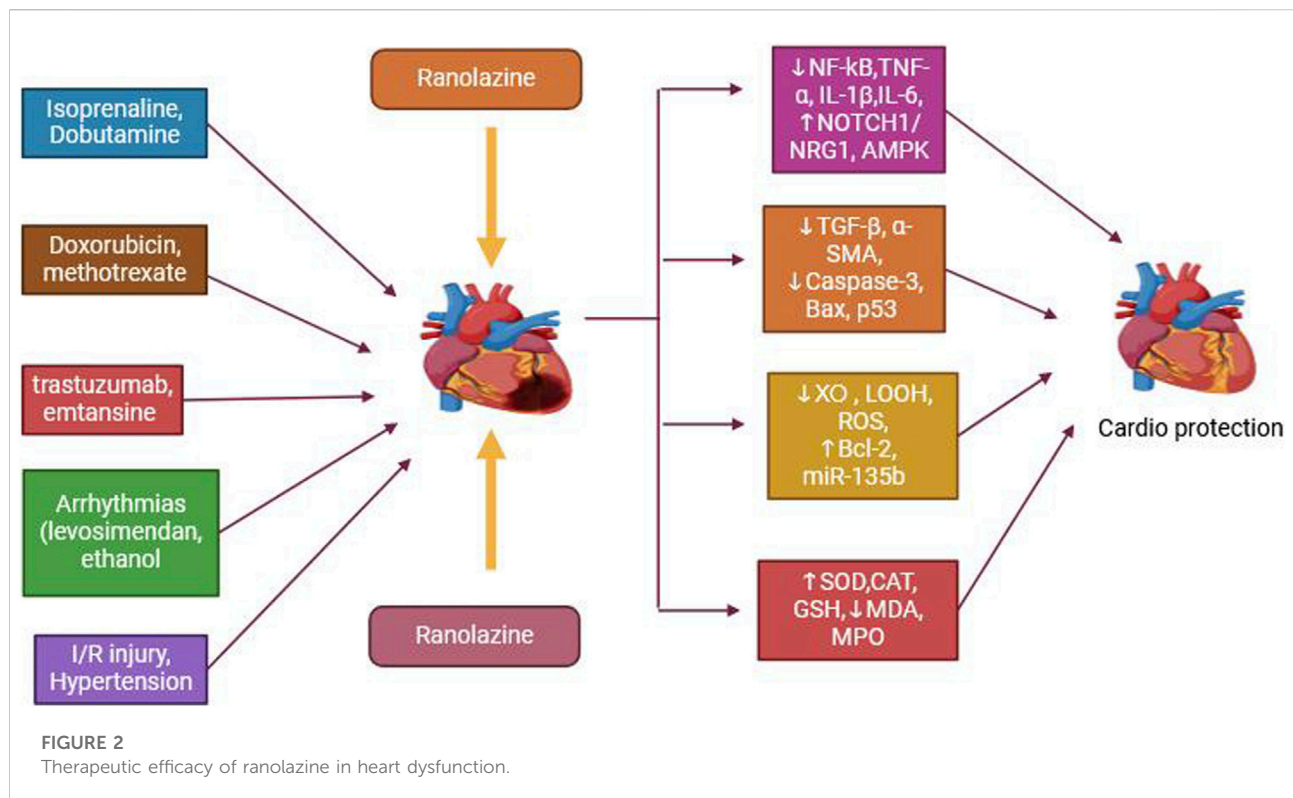
RAN was tested in patients having coronary artery disorder and paroxysmal AF who used to have a double chamber

pacemaker able to detect AF. RAN 375 mg twice each day compared with placebo shortened average AF duration and mean AF length. There was no substantial variation in QTc. The 500 mg and 750 mg arms combined showed a reduction in AF recurrence with borderline statistical significance [89]. RAN has also been demonstrated to result in a greater conversion rate of AF to normal sinus rhythm when administered in combination with amiodarone than amiodarone alone in randomized clinical research including 121 patients [90]. Tsanaxidis et al. found that a single 1000 mg daily treatment of RAN when given with amiodarone leads to a faster recovery to sinus rhythm and a better sinus conversion rate than amiodarone alone. The addition of RAN had no detrimental effect on left ventricular activity [91]. The additive value of RAN to amiodarone in AF has been confirmed by two meta-analyses. The use of RAN accelerates the time for AF cardioversion. It also helps avoid new-onset AF in people with disabilities rhythm of sinus [92, 93].

The HARMONY study demonstrated that combining moderate dosages of oral RAN with decreased doses of dronedarone effectively ameliorated the AF burden in individuals with paroxysmal AF and was tolerated satisfactorily [94]. Many other small trials have found that RAN decreases conversion time from atrial fibrillation to sinus rhythm. It also increases heart function following coronary artery bypass grafting (CABG) [95–98]. Another clinical study explored the impact of RAN on AF in postoperative atrial fibrillation (POAF). Patients having heart valve and/or heart bypass surgery have been involved. The addition of RAN to normal treatment markedly decreased the frequency of POAF. There was no effect on the stay in the intensive care facility or cardiovascular death, but the rate of cardiovascular readmission decreased by 30 days [99].

In patients experiencing acute coronary syndrome without ST-segment acceleration, RAN has been found to minimize the rate of non-sustained ventricular tachycardias and atrial fibrillation (AF) [100]. In another small study of eight patients with long QT syndrome type 3 (LQTS3), RAN was demonstrated to successfully decrease the QT period, hence reducing the frequency of ventricular arrhythmias [101]. Figure 2 shows the therapeutic efficacy of RAN in heart dysfunction. RAN inhibits $TNF-\alpha$, $IL-1\beta$, $NF-\kappa B$, Caspase-3, Bax, ROS, and Ca^{2+} levels and activates Notch, AMPK, and miR-135b Bcl-2, resulting in improved outcomes for cardiac arrhythmia, cardiac fibrosis, cardiac injury, and myocardial infarction.

Cempaka Putri et al. [102] conducted a systematic review and meta-analysis on the efficacy of using RAN to improve diastolic performance and exercise capacity in heart failure with preserved ejection fraction. It was established that RAN was significantly efficacious in improving diastolic performance in heart failure patients with preserved ejection fraction, with no significant effect on blood pressure, heart rate, and ventricular repolarization rate (shortening of the QT interval).



Neuroprotective effects

Piano and colleagues investigated the protective effect of RAN on microglia cells stimulated by LPS and found that RAN counteracts the neurotoxic effect of LPS-activated microglia on 661W neuronal cells [103]. RAN dramatically enhanced cell survival and growth in cultured astrocytes at any tested dose while decreasing LDH loss, Smac/Diablo activity, and Caspase-3 action, demonstrating a decreased rate of cell death [104].

Akgul and co-workers examined the beneficial effect of RAN in a brain I/R model of rats and concluded that RAN helped in cerebral recovery by increasing Bcl-2 and NA levels and decreasing AChE, TNF- α , and ACP levels [105]. Kahlig and team studied the antiepileptic action of RAN in hippocampus neurons and discovered that at therapeutic doses, RAN lowered the action potential firing rate of hippocampal neurons in response to recurrent depolarizing current injections by stabilizing the inactivated states of Na⁺ channels [106]. Peters et al. investigated the possibility of RAN as an anticonvulsant and found that RAN affected Nav1.2 channels, lowering macroscopic currents and slowing the recovery of rapid and slow inactivation of the Nav1.2 channel in hamster ovary cells stably expressing the rat Nav1.2 channel [107].

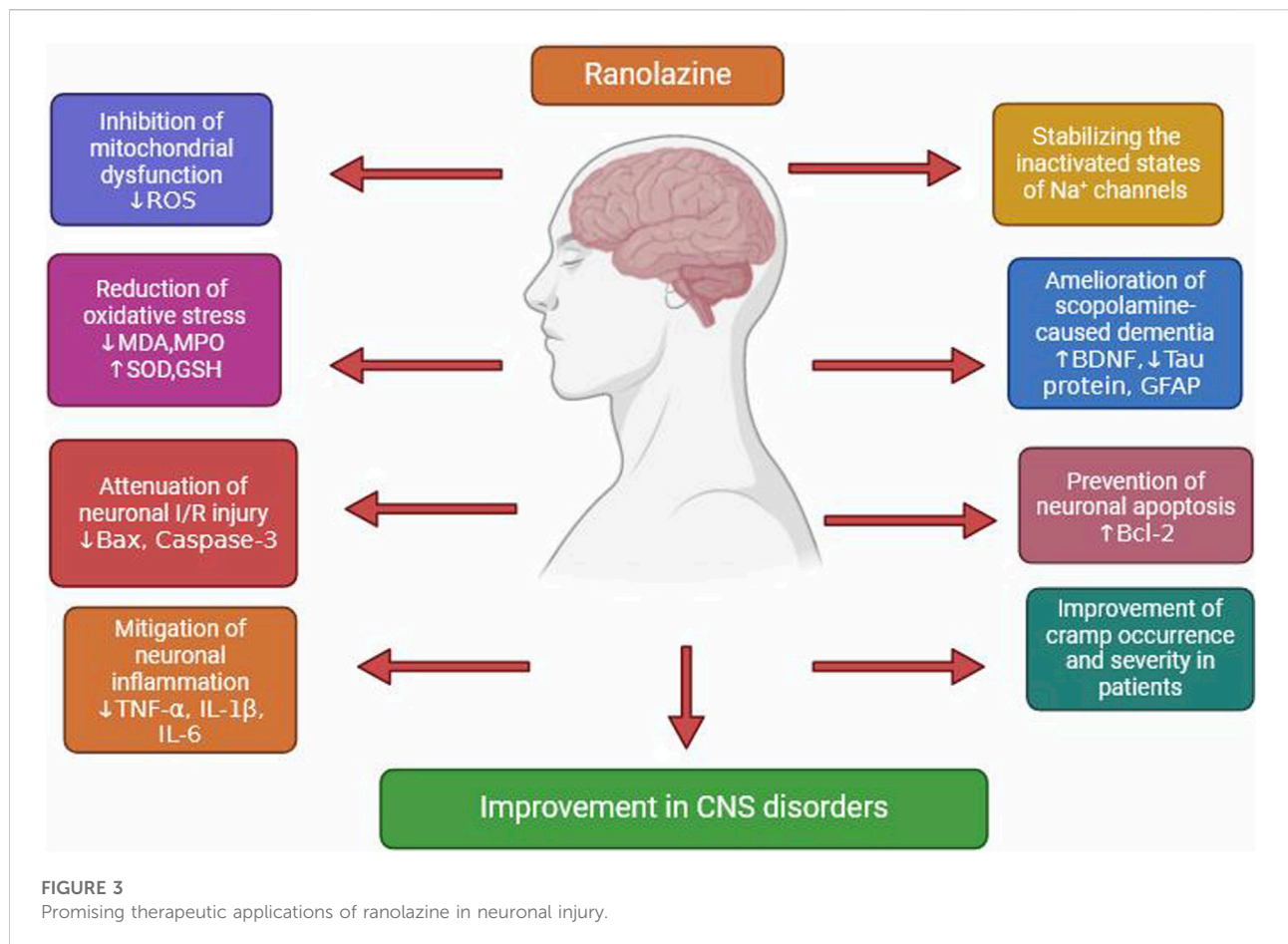
In a rat model of DOX-induced neurotoxicity, RAN reduced brain inflammation, improved BBB integrity, alleviated brain mitochondrial dysfunction, inhibited apoptosis, and preserved

microglial structure and hippocampal plasticity [108]. Samir et al. revealed that RAN has a unique neuroprotective function against scopolamine-caused dementia in rats via antioxidative, anti-inflammatory, and anti-apoptotic actions as well as regulation of GFAP, BDNF, and Tau protein levels [109]. In diabetic neuropathy rats, RAN and pioglitazone have separately altered evoked-pain activity, lowered sciatic TNF- α and IL-1 β levels, decreased levels of Nav1.7 channels, and enhanced expression of the spinal PPAR- γ gene [110]. Chandrashekhar and colleagues conducted an open-label dose-ascending trial of RAN in 14 people with amyotrophic lateral sclerosis, examining muscular cramp symptoms. It was discovered that RAN improved cramp occurrence and severity, which supports its study into muscular cramps [111]. Figure 3 shows promising therapeutic applications of RAN in neuronal injury.

RAN primarily activates anti-apoptotic and neuronal survival pathways such as Bcl-2. It also suppresses Caspase-3, TNF- α , IL-1 β , IL-6, ROS, and other factors that promote neuronal death.

Renal protective effects

RAN substantially reduces renal ischemia-reperfusion damage in rats, which was accomplished by modulating the



inflammatory reactions via a noteworthy drop in renal tissue level of HMG box1, IL-1 β , downregulation of the Notch2/Hes1 signaling pathway, and anti-oxidant action [112]. According to Abbas and teammates, RAN dramatically reduces renal ischemia-reperfusion damage in rats by increasing Bcl2 protein levels, decreasing Bax and TNF-alpha levels, and inhibiting the oxidative stress biomarker F2-isoprostane and Notch2/Hes1 signaling cascade [113]. Nayaka and Vaish revealed that RAN therapy dramatically lowered blood glucose levels, preserved renal functions, and maintained near-normal renal structure due to its glycemic management and anti-inflammatory and anti-oxidative effects against STZ-caused diabetic nephropathy in rats [114]. Ma and associates investigated the protective effect of RAN in contrast-induced acute renal injury (CI-ARI). Pre-treatment of RAN in CI-ARI mice showed no effect on total blood pressure but significantly enhanced renal perfusion, decreased contrast-associated microcirculation disruption, accelerated renal capillary thickness, and ameliorated renal vascular permeation [115]. Yusuf et al. investigated administering RAN as a preventative for patients with low renal failure having PCI and discovered that it might prevent the development of CIN [116].

Pain and inflammation

RAN inhibited DRG neuron hyperexcitability by interfering with inactivated Na (+) channels, and these activities could lead to its anti-allodynic action in animal models of neuropathic pain [117]. Furthermore, at a dose routinely employed in clinical settings, RAN was discovered to be efficacious in preventing the fast firing of DRG neurons with WT Nav1.7 channels, reducing neuropathic and inflammatory pain [118].

RAN has been demonstrated to attenuate pain behavior in animal models of acquired neurotic pain; however, the drug's effects on cold-induced pain were more potent than mechanical allodynia, and the reduction in pain was only temporary, lasting only 30–90 min based on oral or *i.p* delivery [119]. Casey et al. assessed the analgesic efficacy of RAN in complete Freund's adjuvant-mediated inflammatory pain in rats. They found that RAN exhibited a dose-dependent analgesic effect [120]. According to Gould et al. RAN at 30 mg/kg efficiently ameliorated the painful mechanical allodynia related to demyelination injury, which was induced by the administration of doxorubicin [121].

Naveena and colleagues investigated the anti-inflammatory efficacy of RAN in acute and sub-acute inflammation models in rats and found that RAN substantially lowered paw oedema volume and histological sections revealed a reduction in granulation tissue development [122]. Lenz and coworkers claimed that Na⁺ suppression by RAN resulted in lower expression of adhesion molecules and pro-inflammatory cytokines as well as reduced adherence of leukocytes to activated endothelium *in vitro* and *in vivo* [123].

Antidiabetic activity

Jordá et al. found that RAN improved insulin consequences in primary culture astrocytes by increasing anti-inflammatory facilitators like PPAR- γ , decreasing pro-inflammatory agents like COX-2, and boosting the action of Mn-SOD and components of the AKT-eNOS and ERK signaling cascade [124]. Bashir and colleagues investigated the antidiabetic efficacy of RAN against STZ-caused diabetes in rats. It was observed that RAN improved plasma fasting glucose levels and also exhibited a positive effect on the lipid profile [125].

Non-clinical investigations showed that RAN reduced fasting and non-fasting glucose levels and preserved pancreatic β -cells in STZ-induced diabetic mice [126]. In animal models of diabetes, RAN lowered postprandial and basal glucagon concentrations, resulting in a drop in hyperglycemia, demonstrating that RAN's glucose-lowering actions might be achieved via the blocking of sodium channels in pancreatic alpha cells [127]. Guerra-Ojeda explored the potential beneficial effects of RN on insulin activity in the rabbit aorta. They discovered that RAN improved vascular sensitivity to insulin, reducing tissue resistance to the hormone by raising the activities of p-eNOS/eNOS and pAKT/AKT [128]. Cassano et al. assessed the effects of RAN on glucose metabolism and cognitive performance in a T2DM model of Wistar rats and concluded that RAN improved glucose metabolism, enhanced learning and long-term memory, and modified the pro-inflammatory characteristics of diabetic mice [129]. Another study revealed the protective impact of RAN on hippocampal neurodegeneration and astrocyte activation in an STZ T2DM rat model and found that RAN reduced T2DM-induced neuronal injury and loss [130].

A post-hoc examination of the MERLIN-TIMI 36 trials indicated a 0.64 percent drop in HbA1c in diabetic patients who took RAN relative to those who did not. Fasting plasma glucose was also notably decreased by an average of 25.7 mg/dL [131]. Pettus et al. recently verified the MERLIN-TIMI 36 trial results. They investigated the use of high-dose RAN for glycemic control in addition to glimepiride background treatment (4 mg/day) in type 2 diabetes patients with an average baseline HbA1c level of 8.1% [132].

The CARISA research showed that RAN is effective at reducing HbA1c levels in patients with unstable angina. In

this assay, HbA1c was not a given result, and further stratification of results based on insulin or oral antihyperglycemic use was not possible [133]. A later randomized analysis of 465 T2D patients with an average HbA1c~8 controlled by lifestyle alone at the start indicated that RAN resulted in higher declines in HbA1c than placebo at 24 weeks (mean difference = 0.56, $p < 0.0001$) [134]. In addition to its anti-ischemic and antianginal properties, RAN demonstrated the capacity to reduce HbA1c in individuals with coronary artery disease and T2DM in two clinical investigations [135]. In a group of patients with T2D and CCS, RAN, in addition to usual anti-ischemic and glucose-lowering medication, also showed effectiveness in restoring endothelial function and glycemic status, as measured by Hb1Ac and short-term GV indices [136].

Muscle disorder

When 10 μ M RAN was applied for treating C2C12 myoblasts throughout cell growth, transformation, and the development of new myotubes, it increased the levels of myogenic regulator factors (Myf5 and MyoD), suppressed cell progression factor, decreased ROS, and preserved mitochondrial homeostasis [137]. Tomczyk and colleagues evaluated the positive effects of RAN on skeletal muscle function and metabolism in dyslipidemic rats. They learned that RAN-mediated suppression of FFA oxidation in ApoE/LDLR $-/-$ mice resulted in reduced exercise performance and total adenine nucleotide pool [138].

Torcinaro et al. explored the efficacy of RAN in preventing skeletal muscle dysfunctions associated with aging and discovered that RAN administration dramatically enhanced the muscular strength of elderly mice via up-regulating antioxidant and mitochondrial genes, and by increasing NADH-dehydrogenase function [139]. Novak and collaborators revealed that RAN improved muscle functioning compared to mexiletine without major side effects in a mouse model of myotonia congenita [140]. An open treatment study with RAN at a dose of 2×500 mg in 13 patients with chloride channel myotonia showed a significantly reduced EMG myotonia, and according to patient reports, significantly reduced muscle stiffness, and, to a lesser extent, a reduction in muscle weakness and reduced myotonia in clinical tests [141].

Lorusso et al. recently investigated the efficacy of RAN in an open-label trial of 10 patients having paramyotonia congenita and concluded that RAN dramatically reduced both subjective symptoms and clinical myotonia [142]. A phase 2 study is underway to assess the efficacy of RAN in MC, paramyotonia congenita, and Type 1 myotonic dystrophy. Patients with the above conditions were randomized to receive RAN 500 mg twice daily for 2 weeks followed by 1000 mg two times daily for 2 more weeks, compared to placebo. Primary outcomes are quality of life measurements for health and neuromuscular

disease, and EMG to assess for changes in muscle potentials and performance. It is a phase 2 trial to mainly assess the safety profile of the drug in these neuromuscular conditions (NCT02251457).

Pulmonary hypertension

Lee and colleagues investigated the preventive function of RAN against monocrotaline-caused PAH in rat models and found that RAN attenuated ventricular hypertrophy, B-type natriuretic peptide values, fibrosis activation, and cardiovascular mortality [143]. Rocchetti et al. have established that RAN inhibited constitutive elevation of the late sodium current, thereby delaying the development of myocardial remodeling in an experimental rat model of PAH induced by monocrotaline [144]. Teixeira-Fonseca et al. proved that RAN attenuated right ventricular hypertrophy while improving P wavelength and QT period in a monocrotaline-caused PH rat model [145]. In an *in vivo* study, acute treatment of RAN dramatically decreased isoproterenol-caused ventricular tachycardia/ventricular fibrillation and related cardiovascular mortality in rats with pre-existing pulmonary arterial hypertension (PAH) and heart remodeling [146]. Furthermore, a pilot experiment at a single center revealed that 8 of the 11 recruited patients completed all the research exams. The WHO FC, RV function, and exercise tolerance findings revealed improvement without any changes to the invasive hemodynamic measures, and the RV size in PAH patients was decreased after 3 months of RAN medication [147].

A recent double-blind, randomized, placebo-controlled RAN trial (n = 9 RAN, n = 6 placebo) revealed that RAN therapy enhanced RV ejection fraction but not 6-min wall distance (6MWD), N-terminal pro-brain natriuretic peptide, or quality-of-life expectancy measures in patients having precapillary pulmonary hypertension [148]. Finch and colleagues observed that the approved antianginal drug RAN improved cardiopulmonary hemodynamics, functional status, and exercise tolerance in both short-term and long-term (average time on drug approximately 2 years) plans in a cohort of patients with PH-HFpEF [149]. A Phase Ib investigation including 12 PAH patients showed no statistical significance in terms of adverse events between the control and RAN groups after a 12-week follow-up period. This outcome demonstrated the safety of the RAN therapy but did not accomplish the therapeutic aim, partly because the study medication did not reach a therapeutic serum level [150].

Peripheral arterial disease

An animal model demonstrated that injecting RAN into the femoral artery causes a long-lasting dilatation of the artery,

equivalent to that produced by nitroglycerin. This outcome might be attributed to α_1 -adrenergic receptor inhibition, which does not affect heart rate and systemic blood pressure [151]. In a pilot research study including 45 patients with irregular claudication, RAN 1000 mg BID elicited an improvement in peak walking time in comparison with placebo. Though RAN did not ameliorate the ankle-brachial index at rest, patients with extremely irregular claudication had approximately 40 percent improvement in walking time relative to placebo compared to cilostazol [152].

Hepatoprotective effects

Saed and his colleagues assessed the efficacy of RAN in attenuating obesity-induced NAFLD and hyperglycemia and concluded that RAN therapy enhanced glucose tolerance and lowered hepatic triacylglycerol levels in obese mice through increasing the activity of mRNA, which plays a role in modulating lipogenesis [153]. Al Batran stated that in a mouse model of nonalcoholic fatty liver disease, RAN significantly improved glucose oxidation via increasing PDH function [154]. Pzolat and colleagues investigated the preventive effects of RAN against MTX-induced liver injury in rats and found that RAN could attenuate MTX toxicity by reducing MDA and MPO values, enhancing SOD, CAT, and GSH levels, and improving mononuclear inflammation, vascular congestion, and fibrosis [155].

Testicular injury

Bilge et al. evaluated the protective effect of RAN in a testis torsion rat model induced by I/R and demonstrated that RAN protected against testicular damage by reducing MDA levels and improving histopathological scores [156].

Other activities

A recent study revealed that prolonged RAN treatment enhanced energy metabolism by enhancing muscle ATP content and slowing muscular strength reduction in a mouse model of amyotrophic lateral sclerosis (ALS) [157]. Marchio et al. studied the impact of RAN on vascular function and adrenergic response in human saphenous veins. They observed that RAN reduced adrenergic vasoconstriction by acting as an α_1 antagonist and enhancing the huge conductance Ca^{2+} -activated K^+ channel [158].

Molecular mechanisms of non-cardiac effects of Ranolazine

The non-cardiac effects of RAN have been associated with various molecular mechanisms. These include ion channel (late

sodium and calcium) modulation, adrenergic receptor antagonism, and metabolic effects, which collectively result in improved cellular ion homeostasis, reduced oxidative stress, and mild vasodilation in non-cardiac tissues [159]. RAN selectively inhibits the late phase of the inward sodium current and elicits a mild blocking effect on L-type calcium channels. The impacts of these effects include a reduction in intracellular sodium, consequent decrease in calcium overload via the sodium–calcium exchanger (stabilization of cellular ion homeostasis and reduction of cellular stress in tissues), and weak vasodilatory properties with consequences on vascular smooth muscle tone and peripheral circulation [159, 160]. RAN elicits antagonistic action at α -1 and β -1 adrenergic receptors present in vascular, nervous, and other tissues. This antagonistic action contributes to the modulation of vascular tone and sympathetic nervous system effects, devoid of significant changes in heart rate or blood pressure [160].

RAN invokes inhibition of delayed rectifier potassium current, which, beyond cardiac tissue, could influence electrophysiological properties in other excitable tissues [160]. RAN partially inhibits fatty acid oxidation at higher concentrations, leading to alteration of metabolic processes in non-cardiac tissues; this may lead to improvement of cellular energy efficiency under stress conditions [161].

Conclusion

RAN is a well-known selective $I_{Na,L}$ inhibitor and the most commonly utilized antianginal agent. This amazing substance is mostly used to treat chronic angina (chest pain). RAN is an add-on medicine for the relief of symptoms of individuals suffering from stable angina pectoris and those who are poorly controlled or intolerant to first-line antianginal therapy. However, an exciting surge of interest is rising around the possibility of RAN being repurposed for a varied array of health conditions. This review article investigates RAN's varied pharmacological actions, shedding light on its prospective possibilities outside the field of antianginal drugs. The review demonstrates its promise in treating an astounding variety of illnesses, from anticancer activity and neuroprotection to renal and liver protection, renal antidiabetic advantages, and anti-inflammatory capabilities.

The repurposing of RAN offers clinical promise in various health conditions, including pulmonary hypertension,

arrhythmia, heart failure, metabolic disease, and oncology, in view of its unique ion channel modulation, metabolic effects, and anti-inflammatory properties. These benefits of RAN, coupled with its safety profile, offer translational opportunities for diverse therapeutic benefits.

Future perspective

RAN exhibits pleiotropic properties, demonstrating several mechanisms of action and protective benefits against various disease models already established. Given that inflammation and oxidative stress are the fundamental contributors to almost all human diseases, medications that might impede these processes are expected to be beneficial in various medical conditions. The review focuses on the many pharmacological properties of RAN, as it has been demonstrated to produce these effects.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

Conflict of interest

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

References

- McCormack JG, Barr RL, Wolff AA, Lopaschuk GD. Ranolazine stimulates glucose oxidation in normoxic, ischemic, and reperfused ischemic rat hearts. *Circulation* (1996) **93**(1):135–42. doi:10.1161/01.cir.93.1.135
- Létienne R, Vié B, Puech A, Vieu S, Le Grand B, John GW. Evidence that ranolazine behaves as a weak β 1- and β 2-adrenoceptor antagonist in the rat [correction of cat] cardiovascular system. *Naunyn Schmiedeberg's Arch Pharmacol* (2001) **363**(4):464–71. doi:10.1007/s002100000378
- Ratte A, Wiedmann F, Kraft M, Katus HA, Schmidt C. Antiarrhythmic properties of ranolazine: inhibition of atrial fibrillation associated TASK-1 potassium channels. *Front Pharmacol* (2019) **10**:1367. doi:10.3389/fphar.2019.01367
- Tani M, Neely JR. Role of intracellular Na^+ in Ca^{2+} overload and depressed recovery of ventricular function of reperfused ischemic rat hearts. Possible involvement of H^+-Na^+ and Na^+-Ca^{2+} exchange. *Circ Res* (1989) **65**(4):1045–56. doi:10.1161/01.res.65.4.1045

5. Belardinelli L, Shryock JC, Fraser H. Inhibition of the late sodium current as a potential cardioprotective principle: effects of the late sodium current inhibitor ranolazine. *Heart* (2006) **92**(Suppl. 4):iv6–iv14. doi:10.1136/hrt.2005.078790
6. Gupta T, Khera S, Kolte D, Aronow WS, Iwai S. Antiarrhythmic properties of ranolazine: a review of the current evidence. *Int J Cardiol* (2015) **187**:66–74. doi:10.1016/j.ijcard.2015.03.324
7. Chaitman BR. Ranolazine for the treatment of chronic angina and potential use in other cardiovascular conditions. *Circulation* (2006) **113**(20):2462–72. doi:10.1161/CIRCULATIONAHA.105.597500
8. Cocco G, Rousseau MF, Bouvy T, Cheron P, Williams G, Detry JM, et al. Effects of a new metabolic modulator, ranolazine, on exercise tolerance in angina pectoris patients treated with beta-blocker or diltiazem. *J Cardiovasc Pharmacol* (1992) **20**(1):131–8. doi:10.1097/00005344-199207000-00017
9. Jerling M, Huan BL, Leung K, Chu N, Abdallah H, Hussein Z. Studies to investigate the pharmacokinetic interactions between ranolazine and ketoconazole, diltiazem, or simvastatin during combined administration in healthy subjects. *The J Clin Pharmacol* (2005) **45**(4):422–33. doi:10.1177/0091270004273992
10. Jerling M, Abdallah H. Effect of renal impairment on multiple-dose pharmacokinetics of extended-release ranolazine. *Clin Pharmacol & Ther* (2005) **78**(3):288–97. doi:10.1016/j.clpt.2005.05.004
11. Gordon M. *Medical review of safety (ranolazine)*. Rockville, MD: U.S. Food and Drug Administration (2003). Available online at: http://www.fda.gov/ohrms/dockets/ac/03/briefing/4012B2_02_Division%20Dir%20%20Memo.htm (Accessed August 22, 2023).
12. Chaitman BR, Skettino SL, Parker JO, Hanley P, Meluzin J, Kuch J, et al. Antiischemic effects and long-term survival during ranolazine monotherapy in patients with chronic severe angina. *J Am Coll Cardiol* (2004) **43**(8):1375–82. doi:10.1016/j.jacc.2003.11.045
13. Chaitman BR, Pepine CJ, Parker JO, Skopal J, Chumakova G, Kuch J, et al. Effects of ranolazine with atenolol, amlodipine, or diltiazem on exercise tolerance and angina frequency in patients with severe chronic angina: a randomized controlled trial. *JAMA* (2004) **291**(3):309–16. doi:10.1001/jama.291.3.309
14. Chaitman BR. Efficacy and safety of a metabolic modulator drug in chronic stable angina: review of evidence from clinical trials. *J Cardiovasc Pharmacol Ther* (2004) **9**(Suppl. 1):S47–64. doi:10.1177/107424840400900105
15. Drifftort V, Gillet L, Bon E, Marionneau-Lambot S, Oullier T, Joulin V, et al. Ranolazine inhibits Nav1.5-mediated breast cancer cell invasiveness and lung colonization. *Mol Cancer* (2014) **13**:264. doi:10.1186/1476-4598-13-264
16. Qiu S, Fraser SP, Pires W, Djamgoz MBA. Anti-invasive effects of minoxidil on human breast cancer cells: combination with ranolazine. *Clin Exp Metastasis* (2022) **39**(4):679–89. doi:10.1007/s10585-022-10166-7
17. Lee A, Fraser SP, Djamgoz MBA. Propranolol inhibits neonatal Nav1.5 activity and invasiveness of MDA-MB-231 breast cancer cells: effects of combination with ranolazine. *J Cell Physiol* (2019) **234**(12):23066–81. doi:10.1002/jcp.28868
18. Guzel RM, Ogmen K, Ilieva KM, Fraser SP, Djamgoz MBA. Colorectal cancer invasiveness *in vitro*: predominant contribution of neonatal Nav1.5 under normoxia and hypoxia. *J Cell Physiol* (2019) **234**(5):6582–93. doi:10.1002/jcp.27399
19. Rizaner N, Uzun S, Fraser SP, Djamgoz MBA, Altun S. Riluzole: anti-invasive effects on rat prostate cancer cells under normoxic and hypoxic conditions. *Basic & Clin Pharmacol & Toxicol* (2020) **127**(4):254–64. doi:10.1111/bcpt.13417
20. Pemmireddy R, Alvala R, Sama V, Sriramoju A. Effect of ranolazine on 1, 2 - dimethyl hydrazine induced colon cancer in mice. *Asian J Pharm Pharmacol* (2019) **5**(6):1183–90. doi:10.31024/ajpp.2019.5.6.15
21. Bugar I, Kukuc S, Karagoz Z, Fraser SP, Kaya H, Dodson A, et al. Anti-metastatic effect of ranolazine in an *in vivo* rat model of prostate cancer, and expression of voltage-gated sodium channel protein in human prostate. *Prostate Cancer Prostatic Dis* (2019) **22**(4):569–79. doi:10.1038/s41391-019-0128-3
22. Guth A, Monk E, Agarwal R, Bergman BC, Zemski-Berry KA, Minic A, et al. Targeting fat oxidation in mouse prostate cancer decreases tumor growth and stimulates anti-cancer immunity. *Int J Mol Sci* (2020) **21**(24):9660. doi:10.3390/ijms21249660
23. Lasheras-Otero I, Feliu I, Maillou A, Moreno H, Redondo-Muñoz M, Aldaz P, et al. The regulators of peroxisomal acyl-carnitine shuttle CROT and CRAT promote metastasis in melanoma. *J Invest Dermatol* (2023) **143**(2):305–16.e5. doi:10.1016/j.jid.2022.08.038
24. Tocchetti CG, Carpi A, Coppola C, Quintavalle C, Rea D, Campesan M, et al. Ranolazine protects from doxorubicin-induced oxidative stress and cardiac dysfunction. *Eur J Heart Fail* (2014) **16**(4):358–66. doi:10.1002/ehf.50
25. Riccio G, Antonucci S, Coppola C, D'Avino C, Piscopo G, Fiore D, et al. Ranolazine attenuates trastuzumab-induced heart dysfunction by modulating ROS production. *Front Physiol* (2018) **9**:38. doi:10.3389/fphys.2018.00038
26. De Lorenzo C, Paciello R, Riccio G, Rea D, Barbieri A, Coppola C, et al. Cardiotoxic effects of the novel approved anti-ErbB2 agents and reverse cardioprotective effects of ranolazine. *OncoTargets Ther* (2018) **11**:2241–50. doi:10.2147/OTT.S157294
27. Cappetta D, Esposito G, Coppini R, Piegari E, Russo R, Ciuffreda LP, et al. Effects of ranolazine in a model of doxorubicin-induced left ventricle diastolic dysfunction. *Br J Pharmacol* (2017) **174**(21):3696–712. doi:10.1111/bph.13791
28. Dogan Z, Durmus S, Ergun DD, Gelisgen R, Uzun H. Ranolazine exhibits anti-inflammatory and antioxidant activities in H9c2 cardiomyocytes. *Eur Rev Med Pharmacol Sci* (2023) **27**(7):2953–63. doi:10.26355/eurrev_202304_31927
29. Jiang Y, Li X, Guo T, Lu WJ, Ma S, Chang Y, et al. Ranolazine rescues the heart failure phenotype of PLN-deficient human pluripotent stem cell-derived cardiomyocytes. *Stem Cell Rep* (2022) **17**(4):804–19. doi:10.1016/j.stemcr.2022.02.016
30. Ren L, Chen X, Nie B, Qu H, Ju J, Bai Y. Ranolazine inhibits pyroptosis *via* regulation of miR-135b in the treatment of diabetic cardiac fibrosis. *Front Mol Biosci* (2022) **9**:806966. doi:10.3389/fmolb.2022.806966
31. Chen X, Ren L, Liu X, Sun X, Dong C, Jiang Y, et al. Ranolazine protects against diabetic cardiomyopathy by activating the NOTCH1/NRG1 pathway. *Life Sci* (2020) **261**:118306. doi:10.1016/j.lfs.2020.118306
32. Tawfik MK, Ameen AM. Cardioprotective effect of ranolazine in nondiabetic and diabetic male rats subjected to isoprenaline-induced acute myocardial infarction involves modulation of AMPK and inhibition of apoptosis. *Can J Physiol Pharmacol* (2019) **97**(7):661–74. doi:10.1139/cjpp-2018-0571
33. Le DE, Davis CM, Wei K, Zhao Y, Cao Z, Nugent M, et al. Ranolazine may exert its beneficial effects by increasing myocardial adenosine levels. *Am J Physiology-Heart Circulatory Physiol* (2020) **318**(1):H189–H202. doi:10.1152/ajpheart.00217.2019
34. Calderón-Sánchez EM, Domínguez-Rodríguez A, López-Haldón J, Jiménez-Navarro MF, Gómez AM, Smani T, et al. Cardioprotective effect of ranolazine in the process of ischemia-reperfusion in adult rat cardiomyocytes. *Revista Española de Cardiología (English Edition)* (2016) **69**(1):45–53. doi:10.1016/j.rec.2015.02.027
35. Tantray J, Sharma AK, Singh S, Zaid M, Bhat M, Gill K, et al. Ranolazine exert its beneficial effects in myocardial infarction like ischemic preconditioning mediators by increasing myocardial nitric oxide, adenosine, Bradykinin and K⁺ATPase Levels. *Res Square* (2024). doi:10.21203/rs.3.rs-3825042/v1
36. Efentakis P, Andreadou I, Bibli SI, Vasileiou S, Dagres N, Zoga A, et al. Ranolazine triggers pharmacological preconditioning and postconditioning in anesthetized rabbits through activation of RISK pathway. *Eur J Pharmacol* (2016) **789**:431–8. doi:10.1016/j.ejphar.2016.08.001
37. Feng G, Yang Y, Chen J, Wu Z, Zheng Y, Li W, et al. Ranolazine attenuated heightened plasma norepinephrine and B-Type natriuretic peptide-45 in improving cardiac function in rats with chronic ischemic heart failure. *Am J Transl Res* (2016) **8**(2):1295–301.
38. Wang GT, Li H, Yu ZQ, He XN. Effects of ranolazine on cardiac function in rats with heart failure. *Eur Rev Med Pharmacol Sci* (2019) **23**(21):9625–32. doi:10.26355/eurrev_201911_19456
39. Nie J, Duan Q, He M, Li X, Wang B, Zhou C, et al. Ranolazine prevents pressure overload-induced cardiac hypertrophy and heart failure by restoring aberrant Na⁺ and Ca²⁺ handling. *J Cell Physiol* (2019) **234**(7):11587–601. doi:10.1002/jcp.27791
40. Teng S, Ren Z, Zhao K. Vagal stimulation facilitates improving effects of ranolazine on cardiac function in rats with chronic ischemic heart failure. *Curr Mol Med* (2018) **18**(1):36–43. doi:10.2174/1566524018666180608085330
41. Coppini R, Mazzoni L, Ferrantini C, Gentile F, Pioner JM, Laurino A, et al. Ranolazine prevents phenotype development in a mouse model of hypertrophic cardiomyopathy. *Circ Heart Fail* (2017) **10**(3):e003565. doi:10.1161/CIRCHEARTFAILURE.116.003565
42. De Angelis A, Cappetta D, Piegari E, Rinaldi B, Ciuffreda LP, Esposito G, et al. Long-term administration of ranolazine attenuates diastolic dysfunction and adverse myocardial remodeling in a model of heart failure with preserved ejection fraction. *Int J Cardiol* (2016) **217**:69–79. doi:10.1016/j.ijcard.2016.04.168
43. Williams S, Pourrier M, McAfee D, Lin S, Fedida D. Ranolazine improves diastolic function in spontaneously hypertensive rats. *Am J Physiology-Heart Circulatory Physiol* (2014) **306**(6):H867–81. doi:10.1152/ajpheart.00704.2013
44. Venkataraman R, Belardinelli L, Blackburn B, Heo J, Iskandrian AE. A study of the effects of ranolazine using automated quantitative analysis of serial myocardial perfusion images. *JACC: Cardiovasc Imaging* (2009) **2**(11):1301–9. doi:10.1016/j.jcmg.2009.09.006
45. Tagliamonte E, Rigo F, Cirillo T, Astarita C, Quaranta G, Marinelli U, et al. Effects of ranolazine on noninvasive coronary flow reserve in patients with myocardial ischemia but without obstructive coronary artery disease. *Echocardiography* (2015) **32**(3):516–21. doi:10.1111/echo.12674

46. Mehta PK, Goykhman P, Thomson LE, Shufelt C, Wei J, Yang Y, et al. Ranolazine improves angina in women with evidence of myocardial ischemia but no obstructive coronary artery disease. *JACC: Cardiovasc Imaging* (2011) 4(5): 514–22. doi:10.1016/j.jcmg.2011.03.007
47. Schwemer TF, Deutscher N, Diermann N, Böger R, Schwedhelm E, Blankenberg S, et al. Effect of ranolazine on plasma arginine derivatives and urinary isoprostane 8-iso-PGF_{2α} in patients with myocardial infarction in the randomized RIMINI-trial. *Sci Rep* (2019) 9(1):5708. doi:10.1038/s41598-019-42239-1
48. Chou CC, Lee HL, Chang GJ, Wo HT, Yen TH, Wen MS, et al. Mechanisms of ranolazine pretreatment in preventing ventricular tachyarrhythmias in diabetic db/db mice with acute regional ischemia-reperfusion injury. *Sci Rep* (2020) 10(1): 20032. doi:10.1038/s41598-020-77014-0
49. Wolfes J, Ellermann C, Broer N, Rath B, Willy K, Leitz PR, et al. Antiarrhythmic effect of ranolazine in combination with selective NCX-inhibition in an experimental model of atrial fibrillation. *Pharmaceuticals (Basel)*. (2020) 13(10):321. doi:10.3390/ph13100321
50. Ellermann C, Kohnke A, Decherer DG, Kochhäuser S, Reinke F, Fehr M, et al. Ranolazine prevents levosimendan-induced atrial fibrillation. *Pharmacology* (2018) 102(3–4):138–41. doi:10.1159/000490572
51. Moschovidis V, Simopoulos V, Stravela S, Dipla K, Hatziefthimiou A, Stamatou R, et al. Dose-dependent effects of ranolazine on reentrant ventricular arrhythmias induced after subacute myocardial infarction in rabbits. *J Cardiovasc Pharmacol Ther* (2020) 25(1):65–71. doi:10.1177/1074248419858113
52. Markandeya YS, Tsubouchi T, Hacker TA, Wolff MR, Belardinelli L, Balijepalli RC. Inhibition of late sodium current attenuates ionic arrhythmia mechanism in ventricular myocytes expressing LaminA-N195K mutation. *Heart Rhythm* (2016) 13(11):2228–36. doi:10.1016/j.hrthm.2016.08.007
53. Zou D, Geng N, Chen Y, Ren L, Liu X, Wan J, et al. Ranolazine improves oxidative stress and mitochondrial function in the atrium of acetylcholine-CaCl₂ induced atrial fibrillation rats. *Life Sci* (2016) 156:7–14. doi:10.1016/j.lfs.2016.05.026
54. Banga S, Mishra M, Heinze-Milne SD, Jansen HJ, Rose RA, Howlett SE. Chronic testosterone deficiency increases late inward sodium current and promotes triggered activity in ventricular myocytes from aging male mice. *Am J Physiology-Heart Circulatory Physiol* (2023) 325(2):H264–H277. doi:10.1152/ajpheart.00505.2022
55. Mustroph J, Baier MJ, Unsinn D, Provaznik Z, Kozakov K, Lebek S, et al. Ethanol-Induced atrial fibrillation results from late I_{Na} and can be prevented by ranolazine. *Circulation* (2023) 148(8):698–700. doi:10.1161/CIRCULATIONAHA.123.064561
56. Ko TH, Jeong D, Yu B, Song JE, Le QA, Woo SH, et al. Inhibition of late sodium current via PI3K/Akt signaling prevents cellular remodeling in tachypacing-induced HL-1 atrial myocytes. *Pflügers Archiv - Eur J Physiol* (2023) 475(2):217–31. doi:10.1007/s00424-022-02754-z
57. Opačić D, van Hunnik A, Zeemering S, Dhalla A, Belardinelli L, Schotten U, et al. Electrophysiological effects of ranolazine in a goat model of lone atrial fibrillation. *Heart Rhythm* (2021) 18(4):615–22. doi:10.1016/j.hrthm.2020.11.021
58. Frommeyer G, Sterneberg M, Decherer DG, Kochhäuser S, Bögeholz N, Fehr M, et al. Comparison of vernakalant and ranolazine in atrial fibrillation. *J Cardiovasc Med* (2017) 18(9):663–8. doi:10.2459/JCM.0000000000000545
59. Carstensen H, Kjær L, Haugaard MM, Flethøj M, Hesselkilde EZ, Kanters JK, et al. Antiarrhythmic effects of combining dofetilide and ranolazine in a model of acutely induced atrial fibrillation in horses. *J Cardiovasc Pharmacol* (2018) 71(1): 26–35. doi:10.1097/FJC.0000000000000541
60. Daniels JD, Hill JA. Funny and late: targeting currents governing heart rate in atrial fibrillation. *J Cardiovasc Electrophysiol* (2015) 26(3):336–8. doi:10.1111/jce.12597
61. Burashnikov A, Di Diego JM, Barajas-Martínez H, Hu D, Zygmunt AC, Cordeiro JM, et al. Ranolazine effectively suppresses atrial fibrillation in the setting of heart failure. *Circ Heart Fail* (2014) 7(4):627–33. doi:10.1161/CIRCHEARTFAILURE.114.001129
62. Alves Bento AS, Bacic D, Saran Carneiro J, Nearing BD, Fuller H, Justo FA, et al. Selective late I_{Na} inhibition by GS-458967 exerts parallel suppression of catecholamine-induced hemodynamically significant ventricular tachycardia and T-wave alternans in an intact porcine model. *Heart Rhythm* (2015) 12(12):2508–14. doi:10.1016/j.hrthm.2015.07.025
63. Kloner RA, Dow JS, Bhandari A. First direct comparison of the late sodium current blocker ranolazine to established antiarrhythmic agents in an ischemia/reperfusion model. *J Cardiovasc Pharmacol Ther* (2011) 16(2):192–6. doi:10.1177/1074248410386485
64. Malavaki C, Hatziefthimiou A, Daskalopoulou SS, Stefanidis I, Karatzaferi C, Aidonidis I. Ranolazine enhances nicardipine-induced relaxation of alpha-1-adrenoceptor-mediated contraction on isolated rabbit aorta. *Acta Cardiol* (2015) 70(2):157–62. doi:10.1080/ac.70.2.3073506
65. Strege P, Beyder A, Bernard C, Crespo-Diaz R, Behfar A, Terzic A, et al. Ranolazine inhibits shear sensitivity of endogenous Na⁺ current and spontaneous action potentials in HL-1 cells. *Channels (Austin)* (2012) 6(6):457–62. doi:10.4161/chan.22017
66. Koyani CN, Scheruebel S, Jin G, Kolesnik E, Zorn-Paully K, Mächler H, et al. Hypochlorite-modified LDL induces arrhythmia and contractile dysfunction in cardiomyocytes. *Antioxidants (Basel)* (2021) 11(1):25. doi:10.3390/antiox11010025
67. Del-Canto I, Gómez-Cid L, Hernández-Romero I, Guillem MS, Fernández-Santos ME, Atienza F, et al. Ranolazine-mediated attenuation of mechanoelectric feedback in atrial myocyte monolayers. *Front Physiol* (2020) 11:922. doi:10.3389/fphys.2020.00922
68. Souza DS, Marques LP, Costa AD, Cruz JS, Rhana P, Santos-Miranda A, et al. Experimental hypothyroidism induces cardiac arrhythmias and ranolazine reverts and prevents the phenotype. *Life Sci* (2022) 308:120945. doi:10.1016/j.lfs.2022.120945
69. Wu L, Shryock JC, Song Y, Li Y, Antzelevitch C, Belardinelli L. Antiarrhythmic effects of ranolazine in a Guinea pig *in vitro* model of long-QT syndrome. *The J Pharmacol Exp Ther* (2004) 310(2):599–605. doi:10.1124/jpet.104.066100
70. Parikh A, Mantravadi R, Kozhevnikov D, Roche MA, Ye Y, Owen LJ, et al. Ranolazine stabilizes cardiac ryanodine receptors: a novel mechanism for the suppression of early afterdepolarization and torsades de pointes in long QT type 2. *Heart Rhythm* (2012) 9(6):953–60. doi:10.1016/j.hrthm.2012.01.010
71. Aidonidis I, Simopoulos V, Dipla K, Hatziefthimiou A, Stamatou R, Skoularigis I, et al. Effects of ranolazine and its combination with amiodarone on rapid pacing-induced reentrant atrial tachycardia in rabbits. *J Innov Card Rhythm Management* (2021) 12(3):4421–7. doi:10.19102/icrm.2021.120304
72. Nunoi Y, Kambayashi R, Goto A, Hagiwara-Nagasawa M, Chiba K, Izumi-Nakaseko H, et al. *In vivo* characterization of anti-atrial fibrillatory potential and pharmacological safety profile of I_{Na, L} plus I_{Kr} inhibitor ranolazine using the halothane-anesthetized dogs. *Heart Vessels* (2021) 36(7):1088–97. doi:10.1007/s00380-021-01830-1
73. Aidonidis I, Simopoulos V, Stravela S, Dipla K, Stamatou R, Hatziefthimiou A, et al. Ranolazine depresses conduction of rapid atrial depolarizations in a beating rabbit heart model. *J Interv Card Electrophysiol* (2021) 62(1):153–9. doi:10.1007/s10840-020-00865-0
74. Miranda VM, Beserra SS, Campos DR. Inotropic and antiarrhythmic transmural actions of ranolazine in a cellular model of type 3 long QT syndrome. *Arq Bras Cardiol* (2020) 114(4):732–5. doi:10.36660/abc.20190220
75. Chu Y, Yang Q, Ren L, Yu S, Liu Z, Chen Y, et al. Late sodium current in atrial cardiomyocytes contributes to the induced and spontaneous atrial fibrillation in rabbit hearts. *J Cardiovasc Pharmacol* (2020) 76(4):437–44. doi:10.1097/FJC.0000000000000883
76. Rivaud MR, Marchal GA, Wolswinkel R, Jansen JA, van der Made I, Beekman L, et al. Functional modulation of atrioventricular conduction by enhanced late sodium current and calcium-dependent mechanisms in Scn5a1798insD/+ mice. *EP Europace* (2020) 22(10):1579–89. doi:10.1093/europace/eaab127
77. Fukaya H, Plummer BN, Piktet JS, Wan X, Rosenbaum DS, Laurita KR, et al. Arrhythmogenic cardiac alternans in heart failure is suppressed by late sodium current blockade by ranolazine. *Heart Rhythm* (2019) 16(2):281–9. doi:10.1016/j.hrthm.2018.08.033
78. Carstensen H, Hesselkilde EZ, Haugaard MM, Flethøj M, Carlson J, Pehrson S, et al. Effects of dofetilide and ranolazine on atrial fibrillation rate in a horse model of acutely induced atrial fibrillation. *J Cardiovasc Electrophysiol* (2019) 30(4): 596–606. doi:10.1111/jce.13849
79. Montnach J, Chizelle FF, Belbachir N, Castro C, Li L, Loussouarn G, et al. Arrhythmias precede cardiomyopathy and remodelling of Ca²⁺ handling proteins in a novel model of long QT syndrome. *J Mol Cell Cardiol* (2018) 123:13–25. doi:10.1016/j.jmcc.2018.08.019
80. Ke HY, Chin LH, Tsai CS, Lin FZ, Chen YH, Chang YL, et al. Cardiac calcium dysregulation in mice with chronic kidney disease. *J Cell Mol Med* (2020) 24(6): 3669–77. doi:10.1111/jcmm.15066
81. Huang SY, Chen YC, Kao YH, Hsieh MH, Lin YK, Chung CC, et al. Fibroblast growth factor 23 dysregulates late sodium current and calcium homeostasis with enhanced arrhythmogenesis in pulmonary vein cardiomyocytes. *Oncotarget* (2016) 7(43):69231–42. doi:10.18632/oncotarget.12470
82. Hartmann N, Mason FE, Braun I, Pabel S, Voigt N, Schotola H, et al. The combined effects of ranolazine and dronedarone on human atrial and ventricular electrophysiology. *J Mol Cell Cardiol* (2016) 94:95–106. doi:10.1016/j.jmcc.2016.03.012
83. Paci M, Passini E, Severi S, Hyttinen J, Rodriguez B. Phenotypic variability in IQT3 human induced pluripotent stem cell-derived cardiomyocytes and their

response to antiarrhythmic pharmacologic therapy: an *in silico* approach. *Heart Rhythm* (2017) **14**(11):1704–12. doi:10.1016/j.hrthm.2017.07.026

84. Ogawa T, Honjo H, Yamazaki M, Kushiya Y, Sakuma I, Kodama I, et al. Ranolazine facilitates termination of ventricular tachyarrhythmia associated with acute myocardial ischemia through suppression of late I_{Na} -Mediated focal activity. *Circ J* (2017) **81**(10):1411–28. doi:10.1253/circj.CJ-17-0128

85. Bunch TJ, Mahapatra S, Murdock D, Molden J, Weiss JP, May HT, et al. Ranolazine reduces ventricular tachycardia burden and ICD shocks in patients with drug-refractory ICD shocks. *Pacing Clin Electrophysiol* (2011) **34**(12):1600–6. doi:10.1111/j.1540-8159.2011.03208.x

86. Curnis A, Salghetti F, Cerini M, Vizzardi E, Sciatti E, Vassanelli F, et al. Ranolazine therapy in drug-refractory ventricular arrhythmias. *J Cardiovasc Med* (2017) **18**(7):534–8. doi:10.2459/JCM.0000000000000521

87. Black-Maier EW, Pokorney SD, Barnett AS, Liu P, Shrader P, Ng J, et al. Ranolazine reduces atrial fibrillatory wave frequency. *Europace* (2017) **19**(7):1096–100. doi:10.1093/europace/euw200

88. Scirica BM, Morrow DA, Hod H, Murphy SA, Belardinelli L, Hedgepeth CM, et al. Effect of ranolazine, an antianginal agent with novel electrophysiological properties, on the incidence of arrhythmias in patients with non ST-segment elevation acute coronary syndrome: results from the metabolic efficiency with ranolazine for less ischemia in non ST-Elevation acute coronary syndrome thrombolysis in myocardial infarction 36 (MERLIN-TIMI 36) randomized controlled trial. *Circulation* (2007) **116**(15):1647–52. doi:10.1161/CIRCULATIONAHA.107.724880

89. Leftheriotis D, Flevari P, Theodorakis G, Rigopoulos A, Ikonomidis I, Panou F, et al. The effects of ranolazine on paroxysmal atrial fibrillation in patients with coronary artery disease: a preliminary observational study. *J Atr Fibrillation* (2014) **6**(5):940. doi:10.4022/jafib.940

90. Koskinas KC, Fragakis N, Katritsis D, Skeberis V, Vassilikos V. Ranolazine enhances the efficacy of amiodarone for conversion of recent-onset atrial fibrillation. *Europace* (2014) **16**(7):973–9. doi:10.1093/europace/eut407

91. Tsanaxidis N, Aidonidis I, Hatziefthimiou A, Daskalopoulou SS, Giamouzis G, Triposkiadis F, et al. Ranolazine added to amiodarone facilitates earlier conversion of atrial fibrillation compared to amiodarone-only therapy. *Pacing Clin Electrophysiol* (2017) **40**(4):372–8. doi:10.1111/pace.13048

92. De Vecchis R, Ariano C, Giasi A, Cioppa C. Antiarrhythmic effects of ranolazine used both alone for prevention of atrial fibrillation and as an add-on to intravenous amiodarone for its pharmacological cardioversion: a meta-analysis. *Minerva cardioangiologica* (2018) **66**(3):349–59. doi:10.23736/S0026-4725.17.04349-3

93. Guerra F, Romandini A, Barbarossa A, Belardinelli L, Capucci A. Ranolazine for rhythm control in atrial fibrillation: a systematic review and meta-analysis. *Int J Cardiol* (2017) **227**:284–91. doi:10.1016/j.ijcard.2016.11.103

94. Reiffel JA, Camm AJ, Belardinelli L, Zeng D, Karwatowska-Prokopczuk E, Olmsted A, et al. The HARMONY trial: combined ranolazine and dronedarone in the management of paroxysmal atrial fibrillation: mechanistic and therapeutic synergism. *Circ Arrhythmia Electrophysiol* (2015) **8**(5):1048–56. doi:10.1161/CIRCEP.115.002856

95. Simopoulos V, Tagarakis GI, Daskalopoulou SS, Daskalopoulos ME, Lenos A, Chrysagis K, et al. Ranolazine enhances the antiarrhythmic activity of amiodarone by accelerating conversion of new-onset atrial fibrillation after cardiac surgery. *Angiology* (2014) **65**(4):294–7. doi:10.1177/0003319713477911

96. Miles RH, Passman R, Murdock DK. Comparison of effectiveness and safety of ranolazine versus amiodarone for preventing atrial fibrillation after coronary artery bypass grafting. *The Am J Cardiol* (2011) **108**(5):673–6. doi:10.1016/j.amjcard.2011.04.017

97. Burashnikov A, Antzelevitch C. Ranolazine versus amiodarone for prevention of postoperative atrial fibrillation. *Future Cardiol* (2011) **7**(6):733–7. doi:10.2217/fca.11.67

98. Tagarakis GI, Aidonidis I, Daskalopoulou SS, Simopoulos V, Liouras V, Daskalopoulos ME, et al. Effect of ranolazine in preventing postoperative atrial fibrillation in patients undergoing coronary revascularization surgery. *Curr Vasc Pharmacol* (2014) **11**(6):988–91. doi:10.2174/15701611106140128123506

99. Hammond DA, Smotherman C, Jankowski CA, Tan S, Osian O, Kraemer D, et al. Short-course of ranolazine prevents postoperative atrial fibrillation following coronary artery bypass grafting and valve surgeries. *Clin Res Cardiol* (2015) **104**(5):410–7. doi:10.1007/s00392-014-0796-x

100. Gong M, Zhang Z, Fragakis N, Korantzopoulos P, Letsas KP, Li G, et al. Role of ranolazine in the prevention and treatment of atrial fibrillation: a meta-analysis of randomized clinical trials. *Heart Rhythm* (2017) **14**(1):3–11. doi:10.1016/j.hrthm.2016.10.008

101. Chorin E, Hu D, Antzelevitch C, Hochstadt A, Belardinelli L, Zeltser D, et al. Ranolazine for congenital Long-QT syndrome type III: experimental and long-term

clinical data. *Circ Arrhythmia Electrophysiol* (2016) **9**(10):e004370. doi:10.1161/CIRCEP.116.004370

102. Putri DKSC, Andrianto A, Al-Farabi MJ, Saputra PBT, Nugraha RA. Efficacy of ranolazine to improve diastolic performance in heart failure with preserved ejection fraction: a systematic review and meta-analysis. *Eur Cardiol* (2023) **18**:e02. doi:10.15420/ecr.2022.10

103. Piano I, Votta A, Colucci P, Corsi F, Vitolo S, Cerri C, et al. Anti-inflammatory reprogramming of microglia cells by metabolic modulators to counteract neurodegeneration; a new role for ranolazine. *Sci Rep* (2023) **13**(1):20138. doi:10.1038/s41598-023-47540-8

104. Aldasoro M, Guerra-Ojeda S, Aguirre-Rueda D, Mauricio MD, Vila JM, Marchio P, et al. Effects of ranolazine on astrocytes and neurons in primary culture. *PLoS One* (2016) **11**(3):e0150619. doi:10.1371/journal.pone.0150619

105. Akgul E, Gunduz MK, Parlar AI, Guner Y, Eroglu M, Ozhan A, et al. The anti-apoptotic effect of ranolazine on cerebral protection during cardiopulmonary bypass and carotid artery surgery. *Acta Cardiol Sin* (2024) **40**(1):77–86. doi:10.6515/ACS.202401_40(1).20230814C

106. Kahlig KM, Hirakawa R, Liu L, George ALJ, Belardinelli L, Rajamani S. Ranolazine reduces neuronal excitability by interacting with inactivated states of brain sodium channels. *Mol Pharmacol* (2014) **85**(1):162–74. doi:10.1124/mol.113.088492

107. Peters CH, Sokolov S, Rajamani S, Ruben PC. Effects of the antianginal drug, ranolazine, on the brain sodium channel Na(V)1.2 and its modulation by extracellular protons. *Br J Pharmacol* (2013) **169**(3):704–16. doi:10.1111/bph.12150

108. Chunchai T, Arinno A, Ongnok B, Pantiya P, Khuanjing T, Prathumsap N, et al. Ranolazine alleviated cardiac/brain dysfunction in doxorubicin-treated rats. *Exp Mol Pathol* (2022) **127**:104818. doi:10.1016/j.yexmp.2022.104818

109. Samir SM, Hassan HM, Elmowafy R, ElNashar EM, Alghamdi MA, AlSheikh MH, et al. Neuroprotective effect of ranolazine improves behavioral discrepancies in a rat model of scopolamine-induced dementia. *Front Neurosci* (2024) **17**:1267675. doi:10.3389/fnins.2023.1267675

110. Elkholy SE, Elaidy SM, El-Sherbeen NA, Toraih EA, El-Gawly HW. Neuroprotective effects of ranolazine versus pioglitazone in experimental diabetic neuropathy: targeting Nav1.7 channels and PPAR- γ . *Life Sci* (2020) **250**:117557. doi:10.1016/j.lfs.2020.117557

111. Chandrashekar S, Hamasaki AC, Clay R, McCalley A, Herbelin L, Pasnoor M, et al. Open-label pilot study of ranolazine for cramps in amyotrophic lateral sclerosis. *Muscle & Nerve* (2022) **66**(1):71–5. doi:10.1002/mus.27560

112. Abbas LM, Al-Mudhafar RH, Al-Mudhafar DH, Hadi NR. Ranolazine protects the kidney from ischemia/reperfusion injury in adult male rats by modulation of inflammatory and oxidative pathways and suppression of Notch2/Hes1 signalling pathway. *Syst Rev Pharm* (2021) **12**(1):12. doi:10.31838/srp.2021.1.72

113. Abbas LM, Hameed AMA, Abbas WJ, Abdulsatar M, Hadi NR. The anti-apoptotic, anti-inflammatory and anti-oxidant effects of ranolazine on renal ischemia-reperfusion injury in adult Male rats. *Int J Pharm Res* (2021) **13**(01). doi:10.31838/ijpr/2021.13.01.658

114. Nayaka R, Vaish R. Renoprotective potential of ranolazine in ameliorating diabetic nephropathy in a rat model of streptozotocin-induced diabetes. *J Pharm Negative Results* (2022) **13**:1599–605. doi:10.47750/pnr.2022.13.S02.251

115. Ma C, Chen T, Ti Y, Yang Y, Qi Y, Zhang C, et al. Ranolazine alleviates contrast-associated acute kidney injury through modulation of calcium-independent oxidative stress and apoptosis. *Life Sci* (2021) **267**:118920. doi:10.1016/j.lfs.2020.118920

116. Yusuf J, Prakash G, Safal S, Mehta V, Mukhopadhyay S. Efficacy of nicorandil and ranolazine in prevention of contrast-induced nephropathy in patients with mild-to-moderate renal dysfunction: a randomized controlled trial. *Coron Artery Dis* (2024) **35**(3):186–92. doi:10.1097/MCA.0000000000001347

117. Hirakawa R, El-Bizri N, Shryock JC, Belardinelli L, Rajamani S. Block of Na⁺ currents and suppression of action potentials in embryonic rat dorsal root ganglion neurons by ranolazine. *Neuropharmacology* (2012) **62**(7):2251–60. doi:10.1016/j.neuropharm.2012.01.021

118. Estacion M, Waxman SG, Dib-Hajj SD. Effects of ranolazine on wild-type and mutant hNav1.7 channels and on DRG neuron excitability. *Mol Pain* (2010) **6**:35. doi:10.1186/1744-8069-6-35

119. Gould HJ, Garrett C, Donahue RR, Paul D, Diamond I, Taylor BK. Ranolazine attenuates behavioral signs of neuropathic pain. *Behav Pharmacol* (2009) **20**(8):755–8. doi:10.1097/FBP.0b013e3283323c90

120. Casey GP, Roberts JS, Paul D, Diamond I, Gould HJ, 3rd. Ranolazine attenuation of CFA-induced mechanical hyperalgesia. *Pain Med* (2010) **11**(1):119–26. doi:10.1111/j.1526-4637.2009.00763.x

121. Gould HJ, Soignier RD, Cho SR, Hernandez C, Diamond I, Taylor BK, et al. Ranolazine attenuates mechanical allodynia associated with demyelination injury. *Pain Med* (2014) **15**(10):1771–80. doi:10.1111/pme.12516

122. Naveena R, Hashikar NK, Davangeri R, Majagi SI. Effect of anti-inflammatory activity of ranolazine in rat model of inflammation. *Indian J Med Res* (2018) **148**(6):743–7. doi:10.4103/ijmr.IJMR_1504_16
123. Lenz M, Salzmänn M, Ciotu CI, Kaun C, Krychtiuk KA, Rehberger Likozar A, et al. Pharmacologic modulation of intracellular Na⁺ concentration with ranolazine impacts inflammatory response in humans and mice. *Proc Natl Acad Sci USA* (2022) **119**(29):e2207020119. doi:10.1073/pnas.2207020119
124. Jordá A, Aldasoro M, Campo-Palacio I, Vila JM, Aldasoro C, Campos-Campos J, et al. Facilitation of insulin effects by ranolazine in astrocytes in primary culture. *Int J Mol Sci* (2022) **23**(19):11969. doi:10.3390/ijms231911969
125. Bashir S, Kalabharathi HL. Ranolazine improves glucose and lipid homeostasis in streptozotocin induced diabetes mellitus in albino wistar rats. *Int J Basic Clin Pharmacol* (2016) **14**:77–80. doi:10.18203/2319-2003.ijbcp20162456
126. Ning Y, Zhen W, Fu Z, Jiang J, Liu D, Belardinelli L, et al. Ranolazine increases β -cell survival and improves glucose homeostasis in low-dose streptozotocin-induced diabetes in mice. *The J Pharmacol Exp Ther* (2011) **337**(1):50–8. doi:10.1124/jpet.110.176396
127. Dhalla AK, Yang M, Ning Y, Kahlig KM, Krause M, Rajamani S, et al. Blockade of Na⁺ channels in pancreatic α -cells has antidiabetic effects. *Diabetes* (2014) **63**(10):3545–56. doi:10.2337/db13-1562
128. Guerra-Ojeda S, Jordá A, Aldasoro C, Vila JM, Valles SL, Arias-Mutis OJ, et al. Improvement of vascular insulin sensitivity by ranolazine. *Int J Mol Sci* (2023) **24**(17):13532. doi:10.3390/ijms241713532
129. Cassano V, Leo A, Tallarico M, Nesci V, Cimellaro A, Fiorentino TV, et al. Metabolic and cognitive effects of ranolazine in type 2 diabetes mellitus: data from an *in vivo* model. *Nutrients* (2020) **12**(2):382. doi:10.3390/nu12020382
130. Cassano V, Tallarico M, Armentaro G, De Sarro C, Iannone M, Leo A, et al. Ranolazine attenuates brain inflammation in a rat model of type 2 diabetes. *Int J Mol Sci* (2022) **23**(24):16160. doi:10.3390/ijms232416160
131. Morrow DA, Scirica BM, Chaitman BR, McGuire DK, Murphy SA, Karwatowska-Prokopczuk E, et al. Evaluation of the glycometabolic effects of ranolazine in patients with and without diabetes mellitus in the MERLIN-TIMI 36 randomized controlled trial. *Circulation* (2009) **119**(15):2032–9. doi:10.1161/CIRCULATIONAHA.107.763912
132. Pettus J, McNabb B, Eckel RH, Skyler JS, Dhalla A, Guan S, et al. Effect of ranolazine on glycaemic control in patients with type 2 diabetes treated with either glimepiride or metformin. *Diabetes Obes Metab* (2016) **18**(5):463–74. doi:10.1111/dom.12629
133. Chisholm JW, Goldfine AB, Dhalla AK, Braunwald E, Morrow DA, Karwatowska-Prokopczuk E, et al. Effect of ranolazine on A1C and glucose levels in hyperglycemic patients with non-ST elevation acute coronary syndrome. *Diabetes Care* (2010) **33**(6):1163–8. doi:10.2337/dc09-2334
134. Eckel RH, Henry RR, Yue P, Dhalla A, Wong P, Jochelson P, et al. Effect of ranolazine monotherapy on glycemic control in subjects with type 2 diabetes. *Diabetes Care* (2015) **38**(7):1189–96. doi:10.2337/dc14-2629
135. Kosiborod M, Arnold SV, Spertus JA, McGuire DK, Li Y, Yue P, et al. Evaluation of ranolazine in patients with type 2 diabetes mellitus and chronic stable angina: results from the TERISA randomized clinical trial (type 2 diabetes evaluation of ranolazine in subjects with chronic stable angina). *J Am Coll Cardiol* (2013) **61**(20):2038–45. doi:10.1016/j.jacc.2013.02.011
136. Nusca A, Bernardini F, Mangiacapra F, Maddaloni E, Melfi R, Ricottini E, et al. Ranolazine improves glycemic variability and endothelial function in patients with diabetes and chronic coronary syndromes: results from an experimental study. *J Diabetes Res* (2021) **2021**:1–9. doi:10.1155/2021/4952447
137. Ileana T, Anna M, Pamela S, Fernanda V, Stefano B, Livio L. Ranolazine promotes muscle differentiation and reduces oxidative stress in C2C12 skeletal muscle cells. *Endocrine* (2017) **58**(1):33–45. doi:10.1007/s12020-016-1181-5
138. Tomczyk M, Braczko A, Jablonska P, Mika A, Przyborowski K, Jedrzejewska A, et al. Enhanced muscle strength in dyslipidemic mice and its relation to increased capacity for fatty acid oxidation. *Int J Mol Sci* (2021) **22**(22):12251. doi:10.3390/ijms222212251
139. Torcinaro A, Cappetta D, De Santa F, Telesca M, Leigh B, Berrino L, et al. Ranolazine counteracts strength impairment and oxidative stress in aged sarcopenic mice. *Metabolites* (2022) **12**(7):663. doi:10.3390/metabo12070663
140. Novak KR, Norman J, Mitchell JR, Pinter MJ, Rich MM. Sodium channel slow inactivation as a therapeutic target for myotonia congenita. *Ann Neurol* (2015) **77**(2):320–32. doi:10.1002/ana.24331
141. Arnold WD, Kline D, Sanderson A, Hawash AA, Bartlett A, Novak KR, et al. Open-label trial of ranolazine for the treatment of myotonia congenita. *Neurology* (2017) **89**(7):710–3. doi:10.1212/WNL.0000000000004229
142. Lorusso S, Kline D, Bartlett A, Freimer M, Agriesti J, Hawash AA, et al. Open-label trial of ranolazine for the treatment of paramyotonia congenita. *Muscle & Nerve* (2019) **59**(2):240–3. doi:10.1002/mus.26372
143. Lee JC, Kim KC, Choe SY, Hong YM. Reduced immunoreactivities of B-type natriuretic peptide in pulmonary arterial hypertension rats after ranolazine treatment. *Anat Cell Biol* (2016) **49**(1):7–14. doi:10.5115/acb.2016.49.1.7
144. Rocchetti M, Sala L, Rizzetto R, Staszewsky LI, Alemanni M, Zambelli V, et al. Ranolazine prevents INaL enhancement and blunts myocardial remodeling in a model of pulmonary hypertension. *Cardiovasc Res* (2014) **104**(1):37–48. doi:10.1093/cvr/cvu188
145. Teixeira-Fonseca JL, de Lima Conceição MR, Leal-Silva P, Roman-Campos D. Ranolazine exerts atrial antiarrhythmic effects in a rat model of monocrotaline-induced pulmonary hypertension. *Basic & Clin Pharmacol & Toxicol* (2023) **132**(5):359–68. doi:10.1111/bcpt.13845
146. Liles JT, Hoyer K, Oliver J, Chi L, Dhalla AK, Belardinelli L. Ranolazine reduces remodeling of the right ventricle and provoked arrhythmias in rats with pulmonary hypertension. *The J Pharmacol Exp Ther* (2015) **353**(3):480–9. doi:10.1124/jpet.114.221861
147. Khan SS, Cuttita MJ, Beussink-Nelson L, Kozyleva A, Sanchez C, Mkrdichian H, et al. Effects of ranolazine on exercise capacity, right ventricular indices, and hemodynamic characteristics in pulmonary arterial hypertension: a pilot study. *Pulm Circ* (2015) **5**(3):547–56. doi:10.1086/682427
148. Han Y, Forfia P, Vaidya A, Mazurek JA, Park MH, Ramani G, et al. Ranolazine improves right ventricular function in patients with precapillary pulmonary hypertension: results from a double-blind, randomized, placebo-controlled trial. *J Card Fail* (2021) **27**(2):253–7. doi:10.1016/j.cardfail.2020.10.006
149. Finch KT, Stratton EA, Farber HW. Ranolazine for the treatment of pulmonary hypertension associated with heart failure with preserved ejection fraction: a pilot study. *The J Heart Lung Transplant* (2016) **35**(11):1370–3. doi:10.1016/j.healun.2016.07.015
150. Gombert-Maitland M, Schilz R, Mediratta A, Addetia K, Coslet S, Thomeas V, et al. Phase I safety study of ranolazine in pulmonary arterial hypertension. *Pulm Circ* (2015) **5**(4):691–700. doi:10.1086/683813
151. Nieminen T, Tavares CA, Pegler JR, Belardinelli L, Verrier RL. Ranolazine injection into coronary or femoral arteries exerts marked, transient regional vasodilation without systemic hypotension in an intact porcine model. *Circ Cardiovasc Interventions* (2011) **4**(5):481–7. doi:10.1161/CIRCINTERVENTIONS.111.962852
152. Ma A, Garland WT, Smith WB, Skettino S, Navarro MT, Chan AQ, et al. A pilot study of ranolazine in patients with intermittent claudication. *Int Angiol* (2006) **25**(4):361–9.
153. Saed CT, Tabatabaei Dakhili SA, Greenwell AA, Chan JSF, Yang K, Gopal K, et al. The antianginal ranolazine fails to improve glycaemia in obese liver-specific pyruvate dehydrogenase deficient male mice. *Basic & Clin Pharmacol & Toxicol* (2023) **133**(2):194–201. doi:10.1111/bcpt.13906
154. Batran RA, Gopal K, Aburasayn H, Eshreif A, Almutairi M, Greenwell AA, et al. The antianginal ranolazine mitigates obesity-induced nonalcoholic fatty liver disease and increases hepatic pyruvate dehydrogenase activity. *JCI Insight* (2019) **4**(1):e124643. doi:10.1172/jci.insight.124643
155. Polat ME, Sari E, Tanriverdi LH, Gunata M, Aladag M, Sahin AF, et al. A potential hepatoprotective effect of ranolazine against methotrexate-induced liver injury in rats. *Authorea* (2023). doi:10.22541/au.168490817.77526721/v1
156. Keseroglu BB, Ozer E, Karakan T, Ozgur BC, Surer H, Ogus E, et al. Protective effects of Ranolazine on testicular torsion and detorsion injury in rats. *Andrologia* (2020) **52**(7):e13616. doi:10.1111/and.13616
157. Scaricamazza S, Salvatori I, Giacomazzo G, Loeffler JP, Renè F, Rosina M, et al. Skeletal-muscle metabolic reprogramming in ALS-SOD1^{G93A} mice predates disease onset and is a promising therapeutic target. *iScience* (2020) **23**(5):101087. doi:10.1016/j.isci.2020.101087
158. Marchio P, Guerra-Ojeda S, Aldasoro M, Valles SL, Martín-González I, Martínez-León JB, et al. Relaxant and antiadrenergic effects of ranolazine in human saphenous vein. *Eur J Cardio-Thoracic Surg* (2020) **58**(2):277–85. doi:10.1093/ejcts/ezaa034
159. Rouhana S, Virsolvy A, Fares N, Richard S, Thireau J. Ranolazine: an old drug with emerging potential; lessons from pre-clinical and clinical investigations for possible repositioning. *Pharmaceuticals (Basel)* (2021) **15**(1):31. doi:10.3390/ph15010031
160. Rayner-Hartley E, Sedlak T. Ranolazine: a contemporary review. *J Am Heart Assoc* (2016) **5**(3):e003196. doi:10.1161/JAHA.116.003196
161. Kaplan A, Amin G, Abidi E, Altara R, Booz GW, Zouein FA. Role of ranolazine in heart failure: from cellular to clinic perspective. *Eur J Pharmacol* (2022) **919**:174787. doi:10.1016/j.ejphar.2022.174787